

Dear Author,

Please, note that changes made to the HTML content will be added to the article before publication, but are not reflected in this PDF.

Note also that this file should not be used for submitting corrections.



ELSEVIER

Contents lists available at ScienceDirect

Forensic Science International

journal homepage: www.elsevier.com/locate/forsciint

Technical Note

A review of necrophagous insects colonising human and animal cadavers in south-east Queensland, Australia

Q1 Julianne F. Farrell^{a,*}, Andrew E. Whittington^b, Myron P. Zalucki^aQ2^a The University of Queensland, School of Biological Sciences, St. Lucia, Qld 4072, Australia^b Bournemouth University, Talbot Campus, Fern Barrow, Poole BH12 5BB, United Kingdom

ARTICLE INFO

Article history:

Received 16 May 2014

Received in revised form 26 July 2015

Accepted 30 July 2015

Available online xxx

Keywords:

Forensic entomology

Post-mortem

Diptera

Insect colonisation

Human cadavers

Queensland

ABSTRACT

A review of insects collected from decomposing human remains in south-east Queensland yielded 32 species in three orders (Diptera, Coleoptera, Hymenoptera) and 11 families (Calliphoridae, Sarcophagidae, Muscidae, Phoridae, Sepsidae, Chironomidae, Dermestidae, Cleridae, Histeridae, Staphylinidae, Encyrtidae). There were 15 cases where remains were located indoors and five cases where remains were outdoors, in both terrestrial and aquatic environments. Coleoptera were strongly associated with outdoors remains, while dipteran species composition was similar in both indoor and outdoor habitats. Some Diptera were only associated with indoors remains, while others were similarly restricted to remains recovered outdoors. Hymenopteran parasitoids were active in both habitats. Comparative collections were made from other vertebrate remains, including road-kill and farmed animals throughout south-east Queensland (Qld) and northern New South Wales (NSW) during the same period.

© 2015 Published by Elsevier Ireland Ltd.

1. Introduction

Q4 A common legal challenge for forensic entomology relates to the scientific validity of substituting vertebrate remains for human remains in baseline studies [1]. Due to the legal and ethical issues of using human cadavers for research, forensic entomologists have used a variety of vertebrate animals (Table 1) to observe insect succession and generate the baseline data used for post mortem interval estimates in medicolegal cases. Pigs are by far the most internationally accepted human.

This study was initiated to determine the range of necrophagous insects attracted to human remains in south-east Queensland (Qld), and to compare the species observed with those that inhabit other vertebrate remains in the same region. Of the 20 cases reviewed at the Brisbane mortuary, there were 15 cases where remains were located indoors and five cases where remains were outdoors, in both terrestrial and aquatic environments. Coleoptera were strongly associated with outdoors remains, while dipteran species composition was similar in both indoor and outdoor

habitats. Some Diptera were only associated with indoors remains, while others were similarly restricted to remains recovered outdoors. Hymenopteran parasitoids were active in both habitats.

2. Materials and methods

Access to decomposing human remains was obtained through the Queensland Health Forensic and Scientific Services (QHFSS) mortuary in Brisbane.

Approvals for entomological research were granted by Queensland Health, the Queensland Police Service, and The University of Queensland, with the following permits issued:

- I. Queensland Health Human Research Ethics Committee approval number FSS-HEC AU/1/OFF9012. 39
- II. Approval as a 'Genuine Researcher' under S.53(7) Queensland Coroners Act 2003. 42
- III. The University of Queensland Medical Research Ethics Committee, project number 2011001124. 44

During the period December 2011–January 2014, necrophagous insects inhabiting human remains found in outdoor and indoor environments in south-east Queensland (Qld) were collected. Remains had been stored in a cool room running at 4 °C after delivery to the mortuary and prior to autopsy. The time between

Q3 * Corresponding author. Tel.: +61 411 257 831.

E-mail addresses: juliannefarrell17@gmail.com (J.F. Farrell), m.zalucki@uq.edu.au (A.E. Whittington), awhittington@bournemouth.ac.uk (M.P. Zalucki).

<http://dx.doi.org/10.1016/j.forsciint.2015.07.053>

0379-0738/© 2015 Published by Elsevier Ireland Ltd.

Table 1
Summary of vertebrate remains used in decomposition studies to obtain insect succession data.

Author	Location	Animal model
Payne 1965 [3]	South Carolina, USA	Pigs
Richards and Goff 1997 [4]	Hawaii, USA	Pigs
Anderson et al. 2002 [5]	Alberta, Canada	Pigs
Archer 2003 [6]	Victoria, Australia	Pigs
Matuszewski et al. 2008 [7]	Western Poland	Pigs
Sharanowski et al. 2008 [8]	Saskatoon, Canada	Pigs
Eberhardt and Elliot 2008 [9]	Auckland, New Zealand	Pigs
Voss et al. 2008 [10]	WA, Australia	Pigs
Voss et al. 2009, 2011 [11,12]	WA, Australia	Pigs
Anderson 2011 [13]	Alberta, Canada	Pigs
Fuller 1934 [14]	ACT, Australia	Rodents
Kočárek 2003 [15]	Opava, Czech Republic	Rodents
Azwandi et al. 2013 [16]	Selangor, Malaysia	Rodents
Johnson 1975 [17]	Illinois, USA	Rabbits
Tantawi et al. 1996 [18]	Alexandria, Egypt	Rabbits
Bourel et al. 1999 [19]	Ambleteuse, France	Rabbits
Bachmann and Simmons 2010 [20]	Lancashire, UK	Rabbits
Azwandi et al. 2013 [16]	Selangor, Malaysia	Rabbits
Ahmad and Ahmad 2009 [21]	Kedah, Malaysia	Monkeys
Ahmad et al. 2011 [22]	Kedah, Malaysia	Monkeys
Azwandi et al. 2013 [16]	Selangor, Malaysia	Monkeys
Braak 1986 [23]	Transvaal, South Africa	Antelope
O'Flynn and Moorhouse 1979 [24]	Queensland, Australia	Macropods
Reed 1958 [25]	Tennessee, USA	Dogs
Jiron and Cartin 1981 [26]	Costa Rica	Dogs
O'Flynn 1983 [27]	Queensland, Australia	Dogs
O'Flynn and Moorhouse 1979 [24]	Queensland, Australia	Cats
O'Flynn 1983 [27]	Queensland, Australia	Cats
O'Flynn 1983 [27]	Queensland, Australia	Sheep
Johnson 1975 [17]	Illinois, USA	Squirrels
Lang et al. 2006 [28]	Tasmania, Australia	Possums
Bornemissza 1957 [29]	Perth, Australia	Guinea pigs
Cornaby 1974 [30]	Costa Rica	Reptiles
Nelder et al. 2009 [31]	Alabama, USA	Reptiles
Rodriguez and Bass 1983 [32]	Tennessee, USA	Humans
Rodriguez and Bass 1985 [33]	Tennessee, USA	Humans
Schoenly et al. 2002, 2007 [2,34]	Tennessee, USA	Humans

52 delivery of the remains to the mortuary and insect collection was
53 usually 2–3 days.

54 Dipteran adults, pupae, larvae and eggs, and adult Coleoptera
55 were collected from the cadavers, their clothing and inside body
56 bags. Collections were made immediately prior to, and during
57 autopsy. Most of the eggs and larvae found were placed onto fresh
58 kangaroo mince for rearing, while some were killed in hot water
59 and preserved in 70% alcohol. Pupae were rinsed in water, dried
60 and placed into 40 ml disposable plastic containers with perforat-
61 ed lids to continue development. Adults were collected and killed
62 by freezing if not already dead in the clothing and body bag.

63 Eggs, larvae and pupae were reared under ambient conditions
64 in Toowoomba, Qld then identified as adults using published
65 taxonomic keys.

66 Specimens were collected in the presence of mortuary staff, and
67 sometimes investigating police. Decedent information recorded
68 included stage of decomposition, age, gender, date last known to be
69 alive, date delivered to the mortuary, date of insect collection,
70 location and type of death scene, and manner of death.

71 Collected insects were identified to species wherever possible
72 using the taxonomic keys referenced above, reference to collec-
73 tions held by The Queensland Museum (QM) and the Australian
74 National Insect Collection (ANIC), or with assistance from
75 entomologists (Bryan Cantrell, Sasha Voss, Jocelyn King).

76 Based on an insect succession study conducted over two years
77 using pigs, a list of forensically important Diptera for south-east
78 Qld was created [35]. Comparative collections were conducted on
79 an opportunistic basis on more than 80 road-kills and other dead
80 vertebrate remains from central Qld through to central NSW
81 during the same period as the mortuary study. Vertebrate remains
82 examined included kangaroos and wallabies (*Macropus* spp.
83 $n \geq 40$), feral pigs (*Sus scrofa* $n \geq 20$), sheep (*Ovis* sp. $n = 5$), cattle
84 (*Bos taurus* $n = 2$), red foxes (*Vulpes vulpes* $n = 3$), rats (*Rattus*
85 *norvegicus* $n = 2$) and rabbits (*Oryctolagus cuniculus* $n = 5$). Decom-
86 position stages ranged from fresh to skeletonised.

87 The same collection, rearing and identification procedures were
88 followed. These data were compared with insect taxa collected
89 from 20 human remains at the Queensland Health Forensic and
90 Scientific Services mortuary in Brisbane.

91 3. Results

92 From a total of 20 cases, there were four females and sixteen
93 males with an age range of 29 to over 70 years. Manner of death
94 included six non-suspicious/natural causes, seven suicides, one
95 accidental poisoning, one probable homicide and five undeter-
96 mined. The periods between the discovery of the cadaver and time
97 when last known to be alive ranged between 2 days and 6 weeks.
98 The majority of cases originated from urban areas in the Brisbane
99 and Gold Coast regions (Table 2).

100 In total, 32 insect species representing 3 orders (Diptera,
101 Coleoptera, Hymenoptera) and 11 families (Calliphoridae, Sarcophagidae,
102 Muscidae, Phoridae, Sepsidae, Chironomidae, Dermestidae,
103 Cleridae, Histeridae, Staphylinidae, Encyrtidae) were
104 collected and identified. Within the 20 cases, 15 remains were
105 indoors, and five were outdoors (Table 3), including one suicide in a
106 car with open windows in bushland, and one homicide dumped in
107 a river. In all cases, marbling and skin slippage were observed on
108 the remains, regardless of the stage of decomposition.

109 A total of 20 species were collected from the 15 indoors cases,
110 while 24 species were collected from the five outdoors cases. There
111 was greater diversity seen in the outdoors cases, with 11 families
112 represented, compared with six families from the indoors cases
113 (Table 3). Insects listed as unidentified were usually too damaged
114 by body fluids, clothing or body bags for reliable identification
115 beyond family level. *Ch. ruffifacies*, *S. crassipalpis*, *L. cuprina* and *Ch.*
116 *megacephala* were the most frequently collected species from
117 indoors cases, while *Ch. saffrana*, *Ch. ruffifacies*, *Ch. nigripes* and *Ch.*
118 *megacephala* were most frequently encountered on cases originat-
119 ing outdoors. The Sarcophagids were most prevalent in cases
120 originating indoors. In very few of the indoors cases covered in this
121 study were the dwellings closed or sufficiently well screened to
122 impede insect access.

123 Calliphorids were present in 15 of the 20 cases (75%), with the
124 next most frequently encountered family being the Sarcophagids,
125 present in 9 of the 20 cases (45%). Overall, *Ch. ruffifacies* occurred
126 most frequently, being found in 55% of the cases, followed by *Ch.*
127 *saffrana* (present at 40% of cases), *Ch. megacephala* (present at 35%
128 of cases) and *S. crassipalpis* (present at 35% of cases).

129 Hymenopteran parasitic wasps were collected after emergence
130 from dipteran pupae in two mortuary cases.

131 The inspections conducted on road-kill and farmed livestock
132 resulted in the observation or collection of at least 42 species from
133 four orders and 16 families. It is possible that some less common or
134 cryptic species were missed, as searches were often not as
135 exhaustive as those done on human remains due to time or safety
136 constraints. Some carcasses (sheep, cattle) were visited more than
137 once.

138 The most common dipteran species collected or observed from
139 over 80 road-kill or other dead animal remains were *Ch. ruffifacies*,

Table 2
Summary of decedent's personal details from the Brisbane mortuary cases.

Case No	Gender	Age (years)	Weight (kg)	Manner of death	Clothes	Decomp	Habitat
2011-01	♀	>70	Unknown	Non-suspicious	Partial	Early	Indoors
2011-02	♀	60–70	>75	Non-suspicious	Naked	Early	Indoors
2011-03	♂	>70	~60	Non-suspicious	Partial	Early	Indoors
2011-04	♂	64	>100	Non-suspicious	Partial	Advanced	Indoors
2011-05	♂	46	~70	Suicide – CO poisoning	Entirely	Advanced	Outdoors
2012-01	♂	29	50	Suicide-hanging	Entirely	Advanced	Outdoors
2012-02	♀	50	75	Suicide – drug overdose	Partial	Early	Indoors
2012-03	♂	35	43	Non-suspicious	Entirely	Early	Indoors
2014-04	♂	49	78	Drug overdose	Partial	Advanced	Indoors
2012-05	♂	>60	Unknown	Alcoholism	Entirely	Advanced	Indoors
2012-06	♂	69	63	Non-suspicious	Partial	Early	Indoors
2012-07	♂	46	86	Suicide	Entirely	Early	Indoors
2012-08	♀	38	49	Unknown	Entirely	Moderate	Indoors
2012-09	♂	60	59	Suicide	Entirely	Advanced	Indoors
2012-10	♂	68	81	Non-suspicious	Naked	Moderate	Indoors
2012-11	♂	35	Unknown	Suicide – gas	Partial	Advanced	Indoors
2013-01	♂	40	78	Alcoholism	Entirely	Early	Outdoors
2013-QPS	♂	31	~75	Homicide	Partial	Early	Aquatic
2014-01	♂	54	~65	Undetermined	Partial	Advanced	Outdoors
2014-02	♂	59	Unknown	Non-suspicious	Partial	Advanced	Indoors

C. augur, *Ch. saffranaea*, and *Ch. varipes* with *Ch. rufifacies* by far outnumbering all other species. *Saprinus cyaneus*, *Necrobia rufipes*, *Creophilus erythrocephalus* and *Dermestes maculatus* were the most commonly encountered Coleopterans, with the first three species listed being maggot predators.

The list of species from non-human vertebrates is comparable with the most frequently collected species from human remains within the same broad geographical region of south-east Qld (Table 3). Habitat descriptions have been simplified into four categories – grazing describes agricultural land containing grass, shrub and tree vegetation; cropping describes agricultural land largely composed of monoculture crop or fallow paddocks with few shrubs or trees; forest describes national parks or managed plantations composed mainly of shrubs and/or trees; and urban describes domestic and commercial buildings, and their surroundings.

In Table 4, the monthly/seasonal presence of necrophagous Diptera are shown. The collections, from both human and non-human remains were largely opportunistic, so do not necessarily indicate all the months that a particular species is likely to be present in the region.

4. Discussion

In the first trials to test the widely-held assumption that pig remains are reliable analogues for human remains, insect taxa attracted to both porcine and human remains were compared at the University of Tennessee [2,34]. Over 99% of the total taxa caught by sweep nets and pitfall traps were common to both pigs and humans [34]. Between-subject comparisons revealed negligible preferences by forensically important insects for human over porcine remains [2].

While our preliminary study covered a limited time period and included a limited number of cases of decomposing human remains in the Brisbane mortuary, there is strong similarity of arthropod species attracted to both human and other vertebrate remains in the same geographical region.

A review of those insects collected yielded 32 species in 3 orders and 11 families. They were composed of 12 species (37.5%) common to both indoors and outdoors, 8 species (25%) restricted to indoors and 12 species (37.5%) collected only from outdoors cases.

In comparison, Goff [36] examined 35 cadavers in Hawaii and collected 22 species, comprising 5 species (23%) from both

habitats, 14 species (64%) from indoors, and 21 species (95%) from outdoors. Possible explanations for the differences in the percentage presence observed include individual habitat preferences of species specific to Hawaii or Qld, microhabitat around the buildings or outdoor death scenes, or the difference in post mortem intervals recorded in both surveys. Recorded PMI's in Hawaii varied between 2 and 21 days [36], while the cases from the Brisbane mortuary were generally recorded from the date last known to be alive, and ranged between 2 days and over 6 weeks.

The most frequently collected species from both human and other vertebrate remains in south east Qld were *Ch. rufifacies*, *Ch. megacephala*, and the closely related *Ch. saffranaea*, *C. augur*, and *Ch. varipes* (Diptera); and *N. rufipes* and *S. cyaneus* (Coleoptera). These should be considered forensically important species in the subtropical areas of Qld and northern NSW. Although *Ch. rufifacies* has been extensively studied [27,37,38], for the other species listed, published development data is sparse or non-existent, and should be the subject of further work to enable their use in PMI estimates. The early arrivals of *Chrysomya* spp. confirm observations made by others that they behave as primary blowflies in summer in Qld [24]. When comparing species collected from human and non-human remains, the grazing areas attracted a greater diversity of Diptera species to road-kill or other vertebrate remains than did urban, cropped or forested sites (Table 3). This may be a result of the more frequent occurrence of road-kill (mostly macropods) in grazed and forested areas, where timber provides cover and daytime shade for the macropods. With little shelter available, road-kill is less frequently encountered in cropped areas. In contrast, human remains found indoors attracted a greater diversity of species (Table 3).

Sukontason et al. [39] investigated the insect inhabitants of 30 human cadavers and found *Ch. megacephala* and *Ch. rufifacies* to be the most common species in indoor, outdoor, and forested areas in northern Thailand and are regarded as forensically important. Similarly, in Malaysia, Kumara et al. [40] investigated 50 cases, while Kavitha et al. [41] examined 80 cases of human remains. Both found *Ch. megacephala* and *Ch. rufifacies* to be the most frequently collected species. Based on these reports, it would be reasonably safe to suggest that *Ch. megacephala* and *Ch. rufifacies* would dominate carrion and be forensically important species in central and northern Queensland as well as in southern Qld. There are no published works on carrion-associated arthropods in central and northern Qld, however, *Ch. rufifacies*, *Ch. megacephala*, *Ch.*

Table 3
Frequency of insects collected from 20 cases of decomposing human remains and 82 non-human vertebrate remains in various habitats.

Family	Genus/species ^a		Mortuary cases (n = 20)		Other vertebrate remains (n = 82)			
			Indoor	Outdoor	Habitat			
					Grazing	Cropping	Forest	Urban
Calliphoridae	<i>Chrysomya ruffifacies</i> (Macquart, 1842)	a	8	3	12	11	1	1
	<i>Chrysomya ruffifacies</i>	l						
	<i>Chrysomya saffranaea</i> (Bigot, 1877)	a	4	4	4		3	
	<i>Chrysomya saffranaea</i>	l						
	<i>Chrysomya megacephala</i> (Fabricius, 1794)	a	5	2	2	2		
	<i>Chrysomya megacephala</i>	l						
	<i>Lucilia cuprina</i> (Wiedemann, 1830)	a	6	1	1			
	<i>Lucilia cuprina</i>	l						
	<i>Chrysomya nigripes</i> Aubertin, 1932	a	2	3			1	
	<i>Chrysomya nigripes</i>	l						
	<i>Chrysomya varipes</i> (Macquart, 1851)	a	3	1	5	5		
	<i>Chrysomya varipes</i>	l						
	<i>Calliphora augur</i> (Fabricius, 1775)	a	2		5	1		
	<i>Calliphora augur</i> l	l						
	<i>Chrysomya incisuralis</i> (Macquart, 1851)	a	1					
	<i>Chrysomya semimetallica</i> (Malloch, 1927)	l	1					
	<i>Chrysomya flavifrons</i> (Aldrich, 1925)	a		1	3	1		
	<i>Calliphora stygia</i> (Fabricius, 1782)	a				1		1
	<i>Calliphora stygia</i>	l						1
	<i>Calliphora fulvicoxa</i> Hardy, 1930	a			2			
	<i>Calliphora centralis</i> Malloch, 1927	a			1			
	<i>Calliphora hilli</i> Patton, 1925	a			1			
	<i>Calliphora fuscofemorata</i> Malloch, 1927	a	1		1			
	<i>Calliphora ochracea</i> Schiner, 1868	a			1			
	<i>Calliphora ochracea</i>	l						1
	<i>Hemipyrellia ligurriens</i> (Wiedemann, 1830)	a			2			
<i>Hemipyrellia ligurriens</i>	l							
Sarcophagidae	<i>Lucilia papuensis</i> Macquart, 1843	a	1					
	<i>Sarcophaga crassipalpis</i> Macquart, 1839	a	7					
	<i>Sarcophaga crassipalpis</i>	l						1
	<i>Sarcophaga impatiens</i> Walker, 1849	a	3	1		1		5
	<i>Sarcophaga impatiens</i>	l						
	<i>Sarcophaga aurifrons</i> Macquart, 1846	l	1			2		
	<i>Sarcophaga praedatrix</i> Walker, 1849	l		1				2
Unidentified sp.	l	1	1					
Muscidae	<i>Hydrotaea chalcogaster</i> (Wiedemann, 1824)	a	1	1	6	2		
	<i>Hydrotaea chalcogaster</i>	l						
	<i>Synthesiomyia nudiseta</i> (Wulp, 1883)	l	1					
	<i>Musca vetustissima</i> Walker, 1849			1				
	<i>Musca domestica</i> Linnaeus, 1758				1	1		
Unidentified sp.			1					
Phoridae	<i>Megaselia</i> sp. Rondani, 1856	a	4	1				
	<i>Megaselia</i> sp.	l						
Sepsidae	Unidentified sp.	a		1				
Chironomidae	Unidentified sp.	l		1				
Dermestidae	<i>Dermestes maculatus</i> DeGreer, 1774		1	2	6	12	1	
	<i>Dermestes frischii</i> Kugelann, 1792				3			
	<i>Dermestes ater</i> DeGreer, 1774				2			
Cleridae	<i>Necrobia ruficollis</i> (Fabricius, 1775)			1				
	<i>Necrobia rufipes</i> (DeGreer, 1775)			1	5	11		
Histeridae	<i>Saprinus cyaneus</i> (Fabricius, 1775)			2	7	10	1	3
	<i>Saprinus pseudocyanus</i> White, 1846				5	3	1	1
	<i>Saprinus cupreus</i> Erichson, 1834				1			
Staphylinidae	<i>Aleochara</i> sp. Gravenhorst, 1802			1				1
	<i>Creophilus erythrocephalus</i> (Fabricius, 1775)				5	10		
Silphidae	<i>Diamesus osculans</i> (Vigors, 1825)					1		
Scarabaeidae	<i>Aphodius</i> sp. Illiger, 1798				2	1		
	<i>Onthophagus</i> sp. Latreille, 1802					2		
Trogidae	<i>Omorgus</i> sp. Erichson, 1847					2		
Encyrtidae	<i>Tachinaephagus zealandicus</i> Ashmead, 1904		1	1				
Acaridae	<i>Caloglyphus berlesei</i> Michael, 1903				1			
Parasitidae	Unidentified sp.				1			
Laelapidae	Unidentified sp.				1			

^a a = adult, l = immatures.

224 *saffranaea*, *S. cyaneus*, and *N. rufipes* have been regularly collected or
 225 Q5 observed from macropod and feral pig road-kill in central Qld
 226 (XXX, unpublished).

227 The two species collected from human remains and not
 228 collected from road-kill, farmed livestock or domestic pigs used
 229 in a concurrent insect succession study were *Synthesiomyia*

nudiseta (Muscidae) and *Sarcophaga crassipalpis* (Sarcophagidae). 230
 Both species are known to colonise a variety of vertebrate remains 231
 in the region (B. Cantrell, pers comm) and any individuals present 232
 were most likely missed during the carcass search. 233

As has been reported by Kumara et al. [40] in Malaysia, the 234
 Sarcophagids in Qld were more frequently encountered in indoor 235

Table 4

The immature (imm.) and adult development stages of Diptera collected from human and non-human vertebrate remains on a monthly and seasonal basis. ■ denotes human remains. ■ denotes non-human vertebrate remains.

		SUMMER			AUTUMN			WINTER			SPRING		
		Dec	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov
<i>Ch. rufifacies</i>	imm.	■	■	■			■	■		■	■	■	■
	adult	■	■		■								■
<i>Ch. saffranaea</i>	imm.	■	■		■			■		■			
	adult			■	■								■
<i>Ch. megacephala</i>	imm.						■	■		■			■
	adult			■									
<i>Ch. nigripes</i>	imm.			■									■
	adult	■			■								
<i>Ch. varipes</i>	imm.	■	■							■			■
	adult		■	■	■						■		■
<i>Ch. incisuralis</i>	adult			■									
<i>Ch. semimetallica</i>	imm.									■			
<i>Ch. flavifrons</i>	imm.						■	■		■		■	
	adult		■	■			■	■			■		
<i>Ch fuscofemorata</i>	adult						■	■					
<i>C. augur</i>	imm.									■	■		■
	adult	■	■										
<i>C. stygia</i>	imm.										■	■	
	adult										■	■	
<i>C. fulvicoxa</i>	adult										■	■	
<i>C. fuscofemorata</i>	adult										■	■	
<i>C. centralis</i>	adult										■	■	
<i>C. hilli</i>	adult										■	■	
<i>C. ochracea</i>	imm.							■		■			
<i>H. ligurriens</i>	imm.							■					
	adult		■	■									
<i>L. cuprina</i>	imm.	■	■					■		■			■
	adult						■	■			■		
<i>L. papuensis</i>	adult	■	■										
<i>S. crassipalpis</i>	imm.		■	■						■			■
<i>S. impatiens</i>	imm.	■	■								■		■
	adult	■	■										
<i>S. praedatrix</i>	imm.						■	■					■
	adult	■	■										
<i>S. aurifrons</i>	imm.						■	■					
<i>H. chalcogaster</i>	imm.		■	■					■				
	adult	■	■		■						■		
<i>S. nudiseta</i>	imm.	■	■										
<i>Megaselia</i> sp.	imm.							■					
	adult	■	■	■			■	■					■
Sepsidae sp.	adult			■									

environments, with *S. crassipalpis* being collected from 50% of the indoors cases.

Sarcophaga impatiens (active in spring, autumn, winter) was collected from both indoor and outdoor environments, and they were regularly collected during the concurrent insect succession study on the Darling Downs [35]. In one mortuary case, *S. impatiens* and *S. crassipalpis* were the only two species present, suggesting a primary colonisation. In a further three cases, *S. impatiens* probably behaved as a secondary coloniser, arriving after *Ch. megacephala*, *Ch. saffranaea*, *C. ochracea*, *L. cuprina*, *H. ligurriens* and *Megaselia* sp. In these cases, the few *S. impatiens* larvae found were younger than other species present.

In all cases that involved sarcophagids, very few individuals were found. This may be that as ovoviviparous flies, they are out-competed by the calliphorids, which have the capacity to produce large numbers of eggs and larvae. The predatory behaviour of *Ch. rufifacies* larvae may also affect sarcophagid survival in carrion.

Large numbers of *Megaselia* sp. were collected as pupae from the skin and clothing of four indoors cases, while 2 adults were collected from the body bag of one outdoors case. These limited collections indicate a distinct preference for indoors locations, and could possibly serve as a useful indication that a corpse has been moved from an indoors location sometime after death.

In the case where *T. zealandicus* was collected from remains found indoors in summer, they had colonised *Ch. megacephala* or *L. cuprina* (records inconclusive), both species having been collected as 2nd instar larvae. In the case where *T. zealandicus* were collected from indoor remains during winter, they had colonised *H. ligurriens*, collected as 2nd and 3rd instar larvae. A parasitised puparium was retained with the wasps and used for identification of the host species [42]. While the literature describing preferred hosts for *T. zealandicus* is sparse, *H. ligurriens* has not been mentioned before as a known host species (*S. Voss pers comm*).

5. Conclusion

The predominance of Calliphoridae, which were present in 15 of the 20 mortuary cases and all of the road-kill and farmed animal remains, reflects the family's close association with human and other vertebrate remains, justifying the ongoing use of blowflies in forensic investigations. Sarcophagid species were collected from 9 of the 20 mortuary cases, and in three of these cases, were the only larvae present, indicating a potential for *S. crassipalpis* and *S. impatiens* to behave as primary invaders. They appeared to behave as secondary invaders in other cases where much more developmentally advanced Calliphorid larvae were also present.

The conclusion drawn from these results is that the same forensically important insects collected from non-human vertebrate (road-kill and farmed animal) remains are likely to be found at crime scenes involving human remains in the same season and same geographical region. The best forensic indicator species for Qld are likely to be *Ch. rufifacies*, *Ch. megacephala*, *Ch. saffranaea*, *Ch. nigripes* and *S. crassipalpis*.

Taxa collected from human remains in south-east Qld are comparable on a species level to those collected from experimental pig carcasses, road-kill and dead farm animals in the same region and during the same seasons. Thus, it is reasonable to assume that pigs remain a useful human model for forensic entomology research. There should soon be opportunities to compare and test the validity of entomological data generated on non-human models with the recent opening of a 'body farm' west of Sydney in NSW, being managed by the University of Technology, Sydney (UTS) <http://www.abc.net.au/news/2014-11-19/body-farm-to-study-decomposing-human-corpse-set-up-in-sydney/5904394>.

Acknowledgements

We would like to thank the forensic pathologists and technical staff at Queensland Health Forensic & Scientific Services mortuary for their interest and support. Thanks also to Leonard Jarick, Jenny Anderson and others for allowing access to farmed livestock remains or culled feral animals on their properties.

Additionally, thanks to Bryan Cantrell for identification of Sarcophagids, Owen Seeman for the identification of mites, Jocelyn King for the identification of Histerids, and Sasha Voss for identification of the Hymenopteran parasitic wasps.

This study was undertaken by J.F. in partial fulfilment of her M.Phil. in Forensic Entomology at The University of Queensland.

References

[1] E.P. Catts, M.L. Goff, Forensic entomology in criminal investigations, *Annu. Rev. Entomol.* 37 (1992) 253–272.

[2] K.G. Schoenly, N.H. Haskell, R.D. Hall, J.R. Gbur, Comparative performance and complementarity of four sampling methods and arthropod preference tests from human and porcine remains at the Forensic Anthropology Centre in Knoxville, in Tennessee, *J. Med. Entomol.* 44 (5) (2007) 881–894.

[3] J.A. Payne, A summer carrion study of the baby pig *Sus scrofa* Linnaeus, *Ecology* 46 (5) (1965) 592–602.

[4] E.N. Richards, M.L. Goff, Arthropod succession on exposed carrion in three contrasting tropical habitats on Hawaii Island, Hawaii, *J. Med. Entomol.* 34 (3) (1997) 328–339.

[5] G.S. Anderson, N. Hobischak, C. Samborski, O. Beattie, Insect succession on carrion in the Edmonton, Alberta region of Canada, Technical Report TR-04-2002, Canadian Police Research Centre, 2002.

[6] M.S. Archer, Annual variation in arrival and departure times of carrion insects at carcasses: implications for succession studies in forensic entomology, *Aust. J. Zool.* 51 (2003) 569–576.

[7] S. Matuszewski, D. Bajerlein, S. Konwerski, K. Szpila, An initial study of insect succession and carrion decomposition in various forest habitats of Central Europe, *Forensic Sci. Int.* 180 (2008) 61–69.

[8] B.J. Sharanowski, E.G. Walker, G.S. Anderson, Insect succession and decomposition patterns on shaded and sunlit carrion in Saskatchewan in three different seasons, *Forensic Sci. Int.* 179 (2008) 219–240.

[9] T.L. Eberhardt, D.A. Elliot, A preliminary investigation of insect colonisation and succession on remains in New Zealand, *Forensic Sci. Int.* 176 (2008) 217–223.

[10] S.C. Voss, S.L. Forbes, I.R. Dadour, Decomposition and insect succession on cadavers inside a vehicle environment, *Forensic Sci. Med. Pathol.* 4 (2008) 22–32.

[11] S.C. Voss, H. Spafford, I.R. Dadour, Host location and behavioural response patterns of the parasitoid, *Tachinaephagus zealandicus* Ashmead (Hymenoptera: Encyrtidae), to host and host-habitat odours, *Ecol. Entomol.* 34 (2009) 204–213.

[12] S.C. Voss, D.F. Cook, I.R. Dadour, Decomposition and insect succession of clothed and unclothed carcasses in Western Australia, *Forensic Sci. Int.* 211 (2011) 67–75.

[13] G.S. Anderson, Comparison of decomposition rates and faunal colonization of carrion in indoor and outdoor environments, *J. Forensic Sci.* 56 (1) (2011) 136–142.

[14] M.E. Fuller, The insect inhabitants of carrion: a study in animal ecology, *CSIRO Bull.* 82 (1934) 1–62.

[15] P. Kočárek, Decomposition and Coleoptera succession on exposed carrion of small mammal in Opava, the Czech Republic, *Eur. J. Soil Biol.* 39 (2003) 31–45.

[16] A. Azwandi, H.N. Keterina, L.C. Owen, M.D. Nurizzati, B. Omar, Adult carrion arthropod community in a tropical rainforest of Malaysia: analysis of three common forensic entomology animal models, *Trop. Biomed.* 30 (3) (2013) 481–494.

[17] M.D. Johnson, Seasonal and microseral variations in the insect populations on carrion, *Am. Midl. Nat.* 93 (1) (1975) 79–90.

[18] T.I. Tantawi, E.M. El-Kady, B. Greenberg, H.A. El-Ghaffar, Arthropod succession on exposed rabbit carrion in Alexandria, Egypt, *J. Med. Entomol.* 33 (4) (1996) 566–580.

[19] B. Bourel, L. Martin-Bouyer, V. Hedouin, J.-C. Cailliez, D. Derout, D. Gosset, Necrophilous insect succession on rabbit carrion in sand dune habitats in northern France, *J. Med. Entomol.* 36 (4) (1999) 420–425.

[20] J. Bachmann, T. Simmons, The influence of preburial insect access on the decomposition rate, *J. Forensic Sci.* 55 (4) (2010) 893–900.

[21] A. Ahmad, A.H. Ahmad, A preliminary study on the decomposition and dipteran associated with exposed carcasses in an oil plantation in Bandar Baharu, Kedah, Malaysia, *Trop. Biomed.* 26 (1) (2009) 1–10.

[22] N.W. Ahmad, L.H. Lim, C.C. Dhang, H.C. Chi, A.G. Abdullah, W.-N.W. Mustafa, C.W. Kian, J. Jeffery, R. Hashim, S.M. Azirum, Comparative insect fauna succession on indoor and outdoor monkey carcasses in a semi-forested area in Malaysia, *Asian Pac. J. Trop. Biomed.* (2011) S232–S238.

[23] L.E.O. Braak, Arthropods associated with carcasses in the northern Kruger National Park, S. Afr. J. Wildl. Res. 16 (1986) 91–98.

[24] M.A. O'Flynn, D.E. Moorhouse, Species of *Chrysomya* as primary flies in carrion, *J. Aust. Entomol. Soc.* 18 (1979) 31–32.

[25] H.B. Reed, A study of dog carcass communities in Tennessee, with special reference to the insects, *Am. Midl. Nat.* 59 (1) (1958) 213–245.

[26] L.F. Jiron, V.M. Cartin, Insect succession in the decomposition of a mammal in Costa Rica, *J. N. Y. Entomol. Soc.* 89 (3) (1981) 158–165.

[27] M.A. O'Flynn, The succession and rate of development of blowflies in carrion in southern Queensland and the application of these data to forensic entomology, *J. Aust. Entomol. Soc.* 22 (1983) 137–148.

[28] M.D. Lang, G.R. Allen, B.J. Horton, Blowfly succession from possum (*Trichosurus vulpecular*) carrion in a sheep-farming zone, *Med. Vet. Entomol.* 20 (2006) 445–452.

[29] G.F. Bornemissza, An analysis of arthropod succession in carrion and the effect of its decomposition on the soil fauna, *Aust. J. Zool.* 5 (1957) 1–12.

[30] B.W. Cornaby, Carrion reduction by animals in contrasting tropical habitats, *Biotropica* 6 (1) (1974) 51–63.

[31] M.P. Nelder, J.W. McCreedie, C.S. Major, Blow flies visiting decaying alligators: is succession synchronous or asynchronous? *Psyche* 2009 (2009) 7, <http://www.hindawi.com/journals/psyche/2009/575362/> (accessed 15.08.13).

[32] W.C. Rodriguez, W.M. Bass, Insect activity and its relationship to decay rates of human cadavers in East Tennessee, *J. Forensic Sci.* 28 (1983) 423–432.

[33] W.C. Rodriguez, W.M. Bass, Decomposition of buried bodies and methods that may aid in their location, *J. Forensic Sci.* 30 (3) (1985) 836–852.

[34] K.G. Schoenly, R.D. Hall, Testing reliability of animal models in research and training programs in forensic entomology, Part II. *NJ Final Report 97-IJ-CX-0046*, 2002.

[35] J. Farrell, Necrophagous insects in southern Queensland and their potential use as forensic indicators, (Master of Philosophy Thesis), The University of Queensland, 2015.

[36] M.L. Goff, Comparison of insect species associated with decomposing remains recovered inside dwellings and outdoors on the island of Oahu, Hawaii, *J. Forensic Sci.* 36 (3) (1991) 748–753.

[37] J.H. Byrd, J.F. Butler, Effects of temperature on *Chrysomya rufifacies* (Diptera: Calliphoridae) development, *J. Med. Entomol.* 34 (3) (1997) 353–435.

[38] D.L. Baumgartner, Review of *Chrysomya rufifacies* (Diptera: Calliphoridae), *J. Med. Entomol.* 30 (2) (1993) 338–352.

[39] K. Sukontason, P. Narongchai, C. Kanchai, K. Vichairat, et al., Forensic entomology cases in Thailand: a review of cases from 2000 to 2006, *Parasitol. Res.* 101 (5) (2007) 1417–1423.

[40] T.K. Kumara, R.H.L. Disney, A.A. Hassan, M. Flores, T.S. Hwa, Z. Mohamed, M.R.C. Salmah, S. Bhupinder, Occurrence of Oriental flies associated with indoor and outdoor human remains in the tropical climate of North Malaysia, *J. Vector Ecol.* 37 (1) (2012) 62–68.

[41] R. Kavitha, W.A. Nazni, T.C. Tan, H.L. Lee, M.S. Azirum, Review of forensically important entomological specimens collected from human cadavers in Malaysia.

[42] K.L. Sukontason, R. Ngern-Klun, D. Sripakdee, K. Sukontason, Identifying fly puparia by clearing technique: application to forensic entomology, *Parasitol. Res.* 101 (2007) 1407–1416.

335
336
337
338
339
340
341
342
343
344
345
346
347
348
349
350
351
352
353
354
355
356
357
358
359
360
361
362
363
364
365
366
367
368
369
370
371
372
373
374
375
376
377
378
379
380
381
382
383
384
385
386
387
388
389
390
391
392
393
394
395
396
397
398
399
400
401
402
403
404
405
406
407
408
409
410
411
412
413
414
415
416
417
418