

1 **Spatial variability in the somatic growth of pikeperch *Sander lucioperca*, an invasive piscivorous**
2 **fish**

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10 Running head: Growth of pikeperch

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16 Introduced fishes can develop invasive populations that impact native species and ecosystems.
17 Understanding the population ecology of introduced species in their extended ranges and how this
18 compares to their native ranges is therefore important for informing their management. Here, the age
19 and somatic growth rates of the piscivorous freshwater fish pikeperch *Sander lucioperca* were analysed
20 across their invasive and native ranges to determine their spatial patterns and drivers. Analyses were
21 initially completed in their invaded range in central and western England. Populations varied spatially
22 in their growth rates; they were slowest for a population in a narrow and shallow canal, and fastest in a
23 large, impounded lowland river. A meta-analysis of parameters of the von Bertalanffy growth model
24 then revealed that across their native and invasive ranges, their theoretical ultimate lengths (L_{∞}) and
25 growth coefficients (K) were significantly related to latitude, but not longitude. Their relationships with
26 latitude were non-linear, with higher values of L_{∞} and lower values of K being evident towards their
27 northerly and southerly range limits. Faster growth rates were evident in the middle of their range (45
28 to 55 °N), suggesting temperatures here were most optimal for growth, but were in a trade-off with
29 reduced ultimate lengths. These spatial patterns suggest that whilst introduced *S. lucioperca* can
30 colonise new waters across a wide area, the expression of their life history traits will vary spatially,
31 with potential implications for how invasive populations establish and integrate into native fish
32 communities.

33

34 Keywords: Climate change, Invasion, latitude-growth relationships, scale ageing, von Bertalanffy
35 growth model, zander.

36 **Introduction**

37

38 Introductions of non-native fishes can potentially result in the establishment of sustainable populations
39 that naturally disperse and invade (Cucherousset & Olden, 2011; Gozlan, Britton, Cowx, & Copp,
40 2010). Whilst only a small proportion of introduced fishes develop invasions, these fishes can have
41 substantial impacts on native species (Gozlan, Andreou, et al., 2010; Gozlan, Britton, et al., 2010). For
42 piscivorous fishes, impacts tend to be via predation with, for example, invasive largemouth bass
43 *Micropterus salmoides* and peacock basses of the *Cichla* genus having deleterious impacts on native
44 species richness and fish abundance (Gratwicke & Marshall, 2001; Pelicice & Agostinho, 2009).
45 Managing the impact of invasive fishes in open systems is challenging (Britton, Gozlan, & Copp,
46 2011), so predictions that indicate whether an introduced species will develop an invasion are
47 fundamental to their management (Copp et al., 2014, 2016). Spatial assessments of somatic growth
48 rates assist these predictions (Britton, Harper, & Oyugi, 2010), especially as growth rates can be a
49 strong proxy of other life history traits (Oyugi et al., 2011). The expression of life history traits can
50 help to explain invasion patterns and processes for a number of non-native species (Olden, Poff, &
51 Bestgen, 2006; Vila-Gispert, Alcaraz, & García-Berthou, 2005), particularly when related to abiotic
52 parameters (Benejam, Alcaraz, Sasal, Simon-Levert, & García-Berthou, 2009). These patterns have
53 also helped highlight the regions that are most vulnerable to invasion (Cucherousset et al., 2009;
54 Ribeiro, Elvira, Collares-Pereira, & Moyle, 2008).

55

56 Pikeperch *Sander lucioperca* is a large-bodied piscivorous freshwater fish with a native range
57 extending throughout much of Europe, from Germany in the West to Central Russia in the East
58 (Maitland, 2004). The major driver of *S. lucioperca* introductions and translocations has been the
59 diversification of fish assemblages to increase sport angling opportunities (Hickley & Chare, 2004).

60 Their popularity for angling has resulted in a series of regulated and unregulated releases across
61 Western Europe, with their invasive range now including France, Spain, Portugal and Great Britain
62 (Elvira & Almodóvar, 2001; Kopp et al., 2009; Ribeiro et al., 2009). In Britain, the first recorded *S.*
63 *lucioperca* introduction was in 1878, when individuals of German origin were stocked into enclosed
64 waters in the East of England (Copp, Wesley, Kovac, Ives, & Carter, 2003). Subsequent translocations
65 to Eastern England occurred during the 1960s (Wheeler & Maitland, 1973) and it was these releases
66 that lead to their establishment and invasion (Copp et al., 2003; Hickley, 1986; Linfield & Rickards,
67 1979). Reports of anglers capturing individuals from other catchments became widespread throughout
68 the 1970s (Hickley, 1986; Wheeler, 1974), with self-sustaining populations now present throughout
69 central and western England (Copp et al., 2003; Nunn, Bolland, Harvey, & Cowx, 2007; Smith, Leah,
70 & Eaton, 1998).

71

72 Latitude can have a significant influence on the life history traits of freshwater fishes (Blanck &
73 Lamouroux, 2007), mostly via spatial differences in temperature and light intensity (Heibo,
74 Magnhagen, & Vøllestad, 2005; Rypel, 2012). Consequently, latitude is often used as an explanatory
75 variable in assessments of life history trait variation over large spatial scales. This includes fishes in
76 their European invasive range, where growth rates for invasive fishes tend to significantly decrease
77 with increasing latitude (Benejam et al., 2009; Cucherousset et al., 2009). Although used less often,
78 assessments in the longitudinal variability in growth rates can also provide insights into how growth
79 varies spatially (Britton, Davies, & Pegg, 2013). For *S. lucioperca*, many studies have been completed
80 on their life history traits, including growth rates (e.g. Ablak & Yilmaz, 2004; Argillier, Barral, & Irz,
81 2012; Błaszczuk, 2000; Copp et al., 2003; Ložys, 2004). However, these studies have been primarily
82 associated with stock assessment and aquaculture (e.g. Balik, Çubuk, Özkök, & Uysal, 2004; Nyberg,
83 Degerman, & Sers, 1996; Nyina-wamwiza, Xu, Blanchard, & Kestemont, 2005; Özvarol & İklz, 2008).

84 There has been much less focus on their spatial variability in growth parameters (Milardi, Lappalainen,
85 Malinen, Vinni, & Ruuhijärvi, 2011; Pérez-Bote & Roso, 2012), and how their age range and somatic
86 growth rates might vary across their native and invasive distributions. This is despite the ecological and
87 management utility of these data for assisting invasion risk assessments across their invasive range
88 (Copp et al., 2014, 2016).

89
90 Consequently, the aim here was to synthesise data on *S. lucioperca* somatic growth rates from across
91 their native and invasive ranges via a combination of field study and literature review. To initially test
92 how invasive *S. lucioperca* growth rates vary between populations at small spatial scales, a field study
93 focused on invasive populations in central and western England. To then test how *S. lucioperca* growth
94 rates vary across their native and invasive European ranges, a meta-analysis tested spatial patterns in
95 their somatic growth rate parameters across their entire range. As per patterns for other invasive fishes
96 (e.g. Benejam et al., 2009; Cucherousset et al., 2009), it was predicted that latitude would be a
97 significant predictor of *S. lucioperca* growth rates, with decreased growth rates with increased latitude.

98

99 **Methods**

100

101 *Field study sites*

102 The field study was mainly focused on the River Severn basin in central and western England. In this
103 basin, the distribution of *S. lucioperca* is restricted to the Warwickshire Avon (52.0874 N, 1.9481 W),
104 the lower River Severn, generally below the city of Worcester (52.3664 N, 2.3043 W), and the
105 Gloucester-Sharpness Canal that is connected to the Severn estuary at its downstream end (51.7249 N,
106 2.4733 W). In addition, the Grand Union Canal has hydrological connection with the Warwickshire

107 Avon and has invasive *S. lucioperca* present. An area of this canal close to its connection with
108 Warwickshire Avon was thus also sampled (52.2287 N, -0.9159 W).

109

110 A common feature of these invaded waters is that they are heavily regulated for navigation; the Severn
111 and Warwickshire Avon are impounded by navigation weirs, the Grand Union Canal comprises of
112 series of locks to overcome changes in the gradient of the surrounding land and the Gloucester-
113 Sharpness canal was constructed specially for navigation of goods into Gloucester docks. However, the
114 waters differ considerably in their other features. The lower River Severn is up to 40 m in width, with
115 depths to 8 m and is subject to regular winter spates when levels can increase by 5 m. The
116 Warwickshire Avon is generally of widths to 25 m and depths to 4 m, and although also prone to floods
117 in winter, these tend to be much less severe than the Severn. The Grand Union Canal is generally no
118 more than 15 m in width with depths rarely exceeding 2 m, and flood events are rare, whilst the
119 Gloucester-Sharpness canal is unusually broad and deep for a British canal, being approximately 25 m
120 wide with depths to over 5 m. *Sander lucioperca* were confirmed as present in the Warwickshire Avon
121 in 1976, the lower River Severn in 1980 and the Grand Union Canal in 1984 (Hickley, 1986; Nunn et
122 al., 2007). There is no specific evidence over the timing of their introduction into the Gloucester-
123 Sharpness canal or whether it was from a release by anglers or, perhaps more unlikely, natural dispersal
124 via the Severn estuary, although mixohaline waters have been suggested as a potential dispersal route
125 for *S. lucioperca* (Brown, Scott, & Wilson, 2007).

126

127 Growth data on these populations was then supplemented by data on the age and growth rates of *S.*
128 *lucioperca* from the River Great Ouse system (52.3276 N, 0.1769 W). With a catchment area of
129 approx. 8600 km², the River Great Ouse is one of the largest river basins in England and, in the area of
130 sampling, consists of heavily modified, impounded and regulated river channel of 20 m width and

131 depths to 2 m, with the flow regulated by numerous sluices for drainage and flood relief (Pinder,
132 Marker, Mann, Bass, & Copp, 1997). The rationale for including *S. lucioperca* from here was that the
133 fish of the Severn and Great Ouse system all originated from the same original stock that was
134 introduced into Eastern England in 1878 (Copp et al., 2003). Thus, the inclusion of the Great Ouse data
135 utilised fish of the same genetic lineage to the Severn and represented another population from an
136 impounded and regulated river channel.

137

138 *Field sample collection*

139 The age and growth analyses were completed on data obtained from scales collected from *S.*
140 *lucioperca*, as scales provide a consistent and reliable method for age and growth rate analyses
141 (Britton, 2007). The habitat characteristics of the River Severn, Warwickshire Avon ('Lower
142 Warwickshire Avon') and Gloucester Sharpness Canal (Table 1) meant their sampling for *S.*
143 *lucioperca* using typical sampling methods were generally inefficient and/ or unfeasible (e.g. electric
144 fishing, seine netting, fyke netting). Thus, scale sample collection was primarily via catch and release
145 angling. This was facilitated by the Environment Agency, the inland fishery regulatory body of
146 England, who established a network of anglers within the River Severn basin. Participating anglers
147 recorded their catches and were trained in the collection of scale samples. Correspondingly, from 2014
148 to 2017, anglers collected scales (3 to 5 scales per fish from the area below the dorsal fin and above the
149 lateral line) from captured *S. lucioperca*, with additional recording of the location, date of capture and
150 fish fork length (FL, nearest mm). Additional scale samples were collected using seine netting
151 techniques within off-channel boat marinas. For the Grand Union canal, scales were collected from *S.*
152 *lucioperca* in April 2017 where sampling used boat mounted electric fishing. In addition, some data
153 were available for *S. lucioperca* from an upstream site on the Warwickshire Avon, where electric

154 fishing was completed in May 2000 ('Middle Warwickshire Avon'; 52.1894 N, 1.7045 W). The Great
155 Ouse fish were sampled by seine netting in August 2003 and 2005.

156

157 *Age and growth determination from scales*

158 Scales were aged on a projecting microscope (x10 to x48 magnification). Ages were determined by
159 counting of annual growth marks ('annuli'), where an annulus was identified as the transition between
160 two uninterrupted zones of closely and widely spaced circuli. In order to minimise error in age
161 estimations, a confidence scoring system was utilised. In this system, the age estimate was assigned a
162 score of 1 or 2, where 1 indicated relatively high confidence (e.g. scales with clearly defined annuli and
163 high certainty in the age estimate) and 2 indicated reduced confidence (e.g. scales were poorly defined
164 annuli with high age estimate uncertainty). Scales which were assigned a score of 2 were excluded
165 from subsequent analyses to minimise the probability of using data based on low ageing accuracy.
166 Following their ageing, scales were measured for their scale radius and the distances from the scale
167 focus to the first, second and last annulus. These measurements were converted to back-calculated
168 lengths using the Fraser-Lee back-calculation equation (Francis, 1990):

169

170 where L_c is the fish body length at capture, S_i the mean scale length at annulus i , S_c the mean scale total
171 length and c is the intercept from the regression of body length on mean scale length. Back calculated
172 lengths enabled the growth increment between age 1 and 2 to be determined (interpreted as the
173 'juvenile growth rate') and the back calculated length at the last annulus (i.e. hatching date calculated
174 as April according to Lappalainen, Dörner, & Wysujack, (2003) provided a length at age that was not
175 biased by sampling date).

176

177 *Growth rate analyses of scale data*

178 The age and growth data from the scales were analysed in two ways. First, length-at-age data were
179 fitted, using non-linear, least-squares regression, to the von Bertalanffy growth model, , where L_t is
180 length at age t , L_∞ is the asymptotic length, K is the rate at which the curve approaches L_∞ and t_0 is the
181 theoretical age of the fish at zero length. 95 % confidence limits for von Bertalanffy growth parameters
182 were obtained by non-parametric bootstrap resampling over 10,000 iterations. This provided estimated
183 values of L_∞ , K and t_0 .

184

185 Secondly, analysis of standardised growth residuals compared the *S. lucioperca* growth rates across the
186 field sampling sites (other than the ‘Middle Warwickshire Avon’ where the juvenile growth data were
187 not available). The analyses were completed using both lengths at the last annulus and the juvenile
188 growth rate (Amat Trigo, Gutmann Roberts, & Britton, 2017; Beardsley & Britton, 2012). For the
189 juvenile growth rate, the mean length increment (i.e. the back-calculated length difference between 1
190 and 2 years) across all populations was used to calculate the residuals, taken as the difference between
191 the individual length increment of each fish and the mean length increment. For lengths at the last
192 annulus, residuals were calculated using modelled length values, obtained by fitting the back-calculated
193 length at last annulus to the von Bertalanffy growth model across all populations. The residual value of
194 each individual fish was then calculated as the difference between its modelled and observed value.
195 The standardized residual of each individual was determined and compared between populations using
196 ANOVA, with type II sums of squares used to account for unbalanced data due to differences in sample
197 sizes (Langsrud, 2003); Tukeys post-hoc tests were used to determine the significance of differences
198 between the populations.

199

200 *Growth rate comparisons across the ranges of Sander lucioperca*

201 The von Bertalanffy growth model parameters of L_{∞} and K for *S. lucioperca* within these field sites
202 were then compared with data from other populations from across their native and invasive ranges, as
203 gathered by literature review. This review was based on searches completed in Web of Science, and
204 supplemented by Google Scholar, starting with search terms based on the species name ('pikeperch';
205 pike-perch'; 'zander'; '*Stizostedion lucioperca*'; '*Sander lucioperca*') in 'title' searches, and then using
206 these within Boolean logic search terms with words including 'age', 'growth', 'von Bertalanffy',
207 'invasive', 'introduced', 'non-native', and their combinations. Searches were then completed using the
208 same terms but searching for 'topic' to provide any additional material that would otherwise have been
209 missed. These searches were then supplemented by data from Fishbase.org (Froese & Pauly, 2018).
210 Across these studies, data were omitted where the values were considered unreliable or were deemed to
211 be not biologically relevant (Supplementary information, table S1). These criteria were primarily where
212 the value of L_{∞} was considered very high or very low for the species in general, suggesting sampling
213 had not been representative of the population (Živkov, Trichkova, & Raikova-Petrova, 1999), or had
214 been subject to high harvest rates (i.e. an anthropogenic pressure that other populations had not been
215 subjected to). Where von Bertalanffy growth model parameters were calculated based on standard or
216 total length, these were converted to fork length using linear models from Copp et al. (2003) to enable
217 comparisons to be consistent across studies. The relationship between the location of populations, as
218 latitude and longitude, were then tested against L_{∞} and K using linear and non-linear models, where
219 regression statistics and the lowest value of Akaike's Information Criteria (AIC) were used in
220 combination to select the best fitting model. All statistical analysis and graphical outputs were
221 performed using R (Version 3.4.3; R Development Core Team 2017).

222

223 **Results**

224

225 *Field study on growth parameters of invasive Sander lucioperca*

226 There were 625 *S. lucioperca* aged in the field study, of which 472 were retained for analyses based on
227 a confidence score of 1. These retained fish ranged in length between 74 and 770 mm, with individuals
228 aged to 11 years old (Table 1). However, populations from the Grand Union Canal, the Gloucester-
229 Sharpness canal and the 'Middle Warwickshire Avon', were only present in samples to 8 years old
230 (Table 1; Table S2). For the data combined across all populations, L_{∞} was 996 mm and K was 0.13.
231 Across the populations, mean L_{∞} values ranged from 753 to 980 mm, and K between 0.12 and 0.22
232 (Table 1, Fig. 1). However, there was considerable overlap in the 95 % confidence limits of these
233 growth parameters across the populations, suggesting differences between these populations were not
234 significant.

235

236 Analysis of standardised growth residuals revealed significant differences between these populations in
237 both their juvenile growth rates (ANOVA: $F_{4,347} = 45.01$, $P < 0.01$; Fig. 2a) and lengths at the last
238 annulus (ANOVA: $F_{5,434} = 16.97$, $P < 0.01$; Fig. 2b). For juvenile growth rates, Tukey post-hoc tests
239 revealed that significant differences were due to slower growth in the Grand Union Canal population
240 compared to all other populations ($P < 0.01$) and faster growth in the River Great Ouse compared to all
241 other populations ($P < 0.01$). For length at the last annulus, Tukey post-hoc tests showed that
242 significant differences were due to faster growth in length at last annulus on the River Severn
243 population to those of the Grand Union Canal, the River Great Ouse ($P < 0.01$) and the middle
244 Warwickshire Avon ($P < 0.05$). The Grand Union Canal also showed slowest growth across all
245 populations and significant differences with the Gloucester-Sharpness Cana, the River Severn ($P <$
246 0.01) and the River Great Ouse ($P < 0.05$).

247

248 *Spatial variability in Sander lucioperca growth rates*

249 Literature review provided 34 studies with data on the von Bertalanffy growth parameters of *S.*
250 *lucioperca*, of which 22 were retained for further analysis (Supplementary information, Table S1). The
251 analysis also included data from the 6 invasive populations from the field study.

252
253 Across these 22 populations, L_{∞} ranged between 709 and 1116 mm and K between 0.03 and 0.24
254 (Supplementary information, Table S1). The L_{∞} and K parameters derived for the six invasive
255 populations from England sat within these data, with their values towards the higher values of K and
256 lower values of L_{∞} . However, von Bertalanffy growth parameter estimates were only retained from
257 three of the English populations within the field study through application of the criteria used to
258 exclude literature values from the meta-analysis (*cf.* Table S1). This resulted in the data from the lower
259 Warwickshire Avon, the Grand Union Canal and the Gloucester-Sharpness Canal not being included in
260 the analysis. For the 25 retained populations, the relationship between L_{∞} and K was significant, with
261 decreasing values of K as L_{∞} increased (linear regression: $R^2 = 0.19$, $F_{1,23} = 5.56$, $P = 0.02$, Fig. 3). The
262 relationships between latitude and both L_{∞} and K were best described by non-linear regression (AIC;
263 Table 2) and revealed significant U-shaped relationships ($P = 0.05$ and $P < 0.001$ respectively). There
264 were higher values of L_{∞} and lower values of K at the either end of their latitudinal range (Fig. 4). The
265 relationships of longitude with both L_{∞} and K were non-significant ($P > 0.05$; Table 2; Fig. 4).

266

267 **Discussion**

268

269 Across their entire range, the results suggested *S. lucioperca* were rarely present in samples above the
270 age of 10 years and L_{∞} only exceeded 900 mm in a small number of populations. Spatially, there was
271 variation in von Bertalanffy growth parameters across their entire range, with this at least partially
272 explained by the influence of latitude. This spatial variation in von Bertalanffy growth parameters was

273 less apparent in the field study that was completed at a smaller spatial scale. However, the standardized
274 residuals of juvenile growth rates and length at the last annulus indicated some significant differences
275 in growth rates between populations, even at this reduced spatial scale, suggesting factors other than
276 latitude were also important determinants of *S. lucioperca* growth rates.

277

278 It was predicted that latitude would have a significant influence on the growth of *S. lucioperca*, with
279 growth rates decreasing as latitude increases, given that this is a common spatial pattern for many
280 freshwater fishes in the northern hemisphere (Blanck & Lamouroux, 2007; Cucherousset et al., 2009;
281 Heibo et al., 2005), and previous studies have shown fast growth rates in some southern *S. lucioperca*
282 populations (Lappalainen et al., 2003). Whilst there was a significant relationship between both von
283 Bertalanffy growth parameters and latitude, AIC values indicated that the best fitting models were both
284 non-linear, with this contrary to the prediction. Instead, the models indicated U-shaped relationships
285 between the parameters, whereby populations were comprised of individuals of larger body sizes with
286 slower growth rates towards their northerly and southerly limits. These non-linear relationships were
287 likely to have resulted from the population growth rates having a non-linear relationship with
288 environmental factors (Lappalainen, Tarkan, & Harrod, 2008), especially water temperature. This is
289 because water temperatures tend to strongly correlate with latitude, and the strong influence of
290 temperature on fish growth rates is well established (Magnuson, Crowder, & Medvick, 1979). Thus, the
291 increases in K (growth coefficient) that were apparent from approximately 45 to 55 °N might be linked
292 to water temperatures in these areas providing more optimum thermal conditions for faster and efficient
293 growth rates. Indeed, some increases in water temperature have positive effects on *S. lucioperca*
294 growth, with the number of degree days over 10°C increasing their annual length increments (Buijse &
295 Houthuijzen, 1992; Lappalainen et al., 2005; Lappalainen, Milardi, Nyberg, & Venäläinen, 2009;
296 Ložys, 2004). However, faster growth rates tend to generally limit ultimate body sizes due to the

297 influence of, for example, the earlier onset of sexual maturity that diverts energy from somatic growth
298 to reproduction (Ložys, 2004), so potentially explaining the trade-off of higher K values but lower
299 ultimate lengths at 45 to 55 °N.

300

301 At higher latitudes, freshwater fish populations often show slower growth and larger asymptotic lengths
302 (Blanck & Lamouroux, 2007), and thus the results of the relationships of L_{∞} and K with latitude
303 outside of 45 to 55 °N were consistent with this. For example, the slower growth rates that were
304 apparent towards the southerly limits of their range might relate to sub-optimal, warm summer
305 temperatures that prevented their efficient growth, and thus depressed the values of K (Lappalainen et
306 al., 2008). There might have also been a genetic component in the spatial growth patterns of *S.*
307 *luciperca*, given that significant population genetic variation has been detected across their range
308 (Eschbach et al., 2014). This could not, however, be tested here.

309

310 Density dependent factors could also have been influencing the growth rates of these *S. luciperca*
311 populations, with this potentially related to differences in prey availability. In other piscivorous fishes,
312 growth rates were 1.3 times faster at low population densities than high densities in immature walleye
313 *Sander vitreus* (Venturelli, Lester, Marshall, & Shuter, 2010). However, Haugen et al. (2007) revealed
314 conflicting interactions between density-dependent and density-independent factors affecting the
315 growth of pike, *Esox lucius* a large-bodied piscivorous species. Variation in the annual growth rates of
316 a 0+ *S. luciperca* population has been explained by the higher availability of prey species in warmer
317 years (Mooij, Lammens, & Densen, 1994). The onset of piscivory in *S. luciperca* can also influence
318 juvenile growth rates (Buijse & Houthuijzen, 1992), with the realised lengths of piscivorous fishes
319 early in life generally being an important determinant of their ultimate sizes (Mittelbach & Persson,
320 1998). Additionally, *S. luciperca* express faster growth rates in eutrophic waters (Argillier et al.,

321 2012), with Keskinen & Marjomäki (2003) revealing *S. lucioperca* growth was positively correlated
322 with total phosphorus and turbidity, and negatively with size of the water body. In our study, there were
323 some significant differences apparent in juvenile growth rates in the field component. It can thus be
324 hypothesised that differences in the growth rates between the Grand Union Canal and the River Great
325 Ouse to all other populations were at least partially related to differences in prey availability, the
326 physical characteristics of the water body and the onset of obligate piscivory between these
327 populations. However, it was beyond the scope of this study to decouple the relative influences of these
328 factors on *S. lucioperca* growth rates. It was also possible that sample year affected the juvenile growth
329 rates of these populations via, for example, differences in prey availability and temperature.
330 Additionally, these samples were obtained by different sampling methods, increasing the risk of some
331 sampling bias in fish through selective sampling. Thus, the interpretation of these field data warrant
332 some caution due to these issues.

333

334 In the field component of this study, the age and growth rate data were derived only from scales. This
335 was because *S. lucioperca* is now considered a recreationally important fishery resource in most
336 invaded waters in England (Hickley & Chare, 2004), with catch-and-release angling almost universally
337 utilised by anglers. This is despite legislation that controls the distribution of the species in England
338 (Hickley & Chare, 2004), with some evidence suggesting significant ecological impacts on native fish
339 communities following their introduction (Fickling & Lee, 1983; Hickley, 1986; Smith et al., 1998).
340 The use of scales in ageing studies can be problematic, especially in older fishes where the aggregation
341 of annuli on the scale edge can result in ageing errors, with these usually being under-estimations of
342 age (Amat Trigo et al., 2017; Britton et al., 2013). Nevertheless, in a comparative study on the use of
343 calcified structures for age determination in *S. lucioperca* from Turkey, Bostanci (2008) found that
344 scales were typically clear and straight forward to interpret, with the only exception being scales

345 collected from large, long-lived individual fish. Scales were also used as a reliable ageing method for
346 British populations of *S. lucioperca* by Copp et al. (2003) and Britton (2007). The use of the confidence
347 scoring system in ageing the scales should have also increased the reliability of the data used in
348 analyses, with those scales that were difficult to age with high certainty not being used. It should be
349 noted that the scales aged with high uncertainty were not just those from large, slow growing fish, but
350 include scales from smaller fish, where the annuli on the scales were too indistinct to enable a reliable
351 age estimate. There was also no bias in the rejected scale data with respect to the population or the
352 sampling method.

353

354 The use of literature review to compile a meta-analysis of *S. lucioperca* growth data enabled the study
355 to look at growth patterns across a large spatial area. Similar approaches have been recently used for
356 invasive fishes such as roach *Rutilus rutilus* (Tarkan & Vilizzi, 2015) and carp *Cyprinus carpio* (Vilizzi
357 & Copp, 2017). Whilst effective at describing growth over environmental gradients and large spatial
358 areas, differences in how data were collected and/ or analysed between the studies also potentially
359 introduces some discrepancies into analyses. For example, in our study, standard, fork and total length
360 were all used in the reporting of L_{∞} , and so all values were converted to fork length to enable reliable
361 comparisons. In doing so, however, they might have slightly affected the relationship of L_{∞} versus K ,
362 given K values could not be altered in same manner. However, the adjusted difference in lengths was
363 relatively minor (generally < 20 mm) and so did not have a material effect of the relationship of L_{∞}
364 versus K . In addition, there are a number of analytical methods to derive von Bertalanffy growth
365 parameters from length at age data, such as use of two or three parameter growth models that can result
366 in different estimates (Pardo et al., 2013). However, these analytical issues could not be easily
367 controlled in our meta-analysis and, thus, it was assumed that the published values were accurate for
368 the sampled fish.

370 The increasing water temperatures that are generally predicted to occur via climate change will
371 potentially have profound impacts on water resources and river ecosystems (Johnson et al., 2009;
372 Wilby et al., 2006). As a result, there will be major changes in freshwater fish distribution and
373 community structure (Graham & Harrod, 2009; Ruiz-Navarro, Gillingham, & Britton, 2016a).
374 Ecological impacts of freshwater invaders are likely to be enhanced with this warming, such as through
375 altered competitive interactions and increased predation pressure on native species (Rahel & Olden,
376 2008). However, predicting the response of specific invaders to warming is inherently difficult due to
377 these being influenced through complex direct and indirect effects (Britton, Cucherousset, Davies,
378 Godard, & Copp, 2010; Kuczynski, Legendre, & Grenouillet, 2018). For example, in temperate
379 freshwaters, it is likely that all fishes (plus other taxa) will respond to warming by altering their
380 distributions, life history traits and phenology (Comte & Grenouillet, 2013; Ruiz-Navarro et al., 2016a;
381 Ruiz-Navarro, Gillingham, & Britton, 2016b). This is likely to lead to range changes and altered
382 population abundances (Ruiz-Navarro et al., 2016b). For *S. lucioperca* to invade temperate regions, low
383 water temperatures are not considered a constraint due to them being primarily a cold-water species
384 capable of reproducing at relatively low temperatures. Indeed, the temperate climate of England has not
385 prevented population establishment and invasion in the last 50 years (Hickley, 1986; Copp et al., 2003).
386 Thus, it is unlikely that climate change will have a substantial influence on their ability to invade new
387 temperate regions, unless the warming results in temperatures that are too high for their survival.
388 However, given the significant relationships between latitude and their von Bertalanffy growth
389 parameters, then it is likely that as warming proceeds then the impact for *S. lucioperca* is likely to be
390 through altered growth rates. In England, for example, it is likely that their ultimate lengths will reduce
391 and growth rates increase, and potentially result in more abundant populations comprised of smaller
392 individuals. This is in line with predictions for a number of native fishes (Ruiz-Navarro et al., 2016b).

394 In summary, this meta-analysis of the von Bertalanffy growth parameters of *S. lucioperca* suggested
395 that whilst their introductions can result in invasive populations within a wide spatial area and in
396 climates that range from temperate to Mediterranean, the expression of their life history traits will vary
397 considerably. Growth rates will be faster in their mid-range (approximately 45 to 55 °N) than at their
398 northerly and southerly range limits, most likely due to the influence of temperature, although it is
399 acknowledged that other factors will influence their growth at the population level, such as prey
400 availability. These results highlight the extent to which their growth data varies spatially and can be
401 applied to their invasion management by providing a more robust basis for risk assessments that utilise
402 data on their life history traits.

403

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405

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409

410 **References**

411 Ablak, Ö., & Yilmaz, M. (2004). Growth properties of pikeperch (*Sander lucioperca* (L., 1758)) living
412 in Hirfanlı Dam Lake. *Turkish Journal of Veterinary and Animal Sciences*, 28(3), 455–463.

413 Amat Trigo, F., Gutmann Roberts, C., & Britton, J. R. (2017). Spatial variability in the growth of
414 invasive European barbel *Barbus barbus* in the River Severn basin, revealed using anglers as
415 citizen scientists. *Knowledge & Management of Aquatic Ecosystems*, (418), 17.

416 <https://doi.org/10.1051/kmae/2017009>

- 417 Argillier, C., Barral, M., & Irz, P. (2012). Growth and diet of the pikeperch *Sander lucioperca* (L.) in
418 two French reservoirs. *Archives of Polish Fisheries*, 20(3), 191–200.
- 419 Balik, I., Çubuk, H., Özkök, R., & Uysal, R. (2004). Size composition, growth characteristics and stock
420 analysis of the pikeperch, *Sander lucioperca* (L. 1758), population in Lake Eğirdir. *Turkish*
421 *Journal of Veterinary and Animal Sciences*, 28(4), 715–722.
- 422 Beardsley, H., & Britton, J. R. (2012). Contribution of temperature and nutrient loading to growth rate
423 variation of three cyprinid fishes in a lowland river. *Aquatic Ecology*, 46(1), 143–152.
424 <https://doi.org/10.1007/s10452-011-9387-3>
- 425 Benejam, L., Alcaraz, C., Sasal, P., Simon-Levert, G., & García-Berthou, E. (2009). Life history and
426 parasites of the invasive mosquitofish (*Gambusia holbrooki*) along a latitudinal gradient.
427 *Biological Invasions*, 11(10), 2265–2277. <https://doi.org/10.1007/s10530-008-9413-0>
- 428 Blanck, A., & Lamouroux, N. (2007). Large-scale intraspecific variation in life-history traits of
429 European freshwater fish. *Journal of Biogeography*, 34(5), 862–875.
430 <https://doi.org/10.1111/j.1365-2699.2006.01654.x>
- 431 Błaszczuk, P. (2000). Growth rate of zander (*Stizostedion lucioperca* L., 1758) in the water of
432 Miedzyodrze in 1996-1998. *Acta Ichthyologica et Piscatoria*, 30(2), 35–46.
- 433 Bostanci, D. (2008). A comparison of calcified structures for aging of pikeperch (*Sander lucioperca*) in
434 Bafra Fish Lake, Turkey. *Journal of Freshwater Ecology*, 23(3), 485–486.
435 <https://doi.org/10.1080/02705060.2008.9664230>
- 436 Britton, J. R. (2007). Reference data for evaluating the growth of common riverine fishes in the UK.
437 *Journal of Applied Ichthyology*, 23(5), 555–560. [https://doi.org/10.1111/j.1439-](https://doi.org/10.1111/j.1439-0426.2007.00845.x)
438 [0426.2007.00845.x](https://doi.org/10.1111/j.1439-0426.2007.00845.x)
- 439 Britton, J. R., Cucherousset, J., Davies, G. D., Godard, M. J., & Copp, G. H. (2010). Non-native fishes
440 and climate change: predicting species responses to warming temperatures in a temperate

441 region. *Freshwater Biology*, 55(5), 1130–1141. <https://doi.org/10.1111/j.1365->
442 2427.2010.02396.x

443 Britton, J R, Gozlan, R. E., & Copp, G. H. (2011). Managing non-native fish in the environment. *Fish*
444 *and Fisheries*, 12(3), 256–274. <https://doi.org/10.1111/j.1467-2979.2010.00390.x>

445 Britton, J. R., Harper, D. M., & Oyugi, D. O. (2010). Is the fast growth of an equatorial *Micropterus*
446 *salmoides* population explained by high water temperature?: Growth of non-native *Micropterus*
447 *salmoides*. *Ecology of Freshwater Fish*, 19(2), 228–238. <https://doi.org/10.1111/j.1600->
448 0633.2010.00407.x

449 Britton, J R, Davies, G. D., & Pegg, J. (2013). Spatial variation in the somatic growth rates of European
450 barbel *Barbus barbus*: a UK perspective. *Ecology of Freshwater Fish*, 22(1), 21–29.
451 <https://doi.org/10.1111/j.1600-0633.2012.00588.x>

452 Brown, J. A., Scott, D. M., & Wilson, R. W. (2007). Do estuaries act as saline bridges to allow
453 invasion of new freshwater systems by non-indigenous fish species? In *Gherardi F. (eds)*
454 *Biological invaders in inland waters: Profiles, distribution, and threats* (Vol. 2, pp. 401–414).
455 Springer, Dordrecht.

456 Buijse, A. D., & Houthuijzen, R. P. (1992). Piscivory, Growth, and Size-Selective Mortality of Age 0
457 Pikeperch (*Stizostedion lucioperca*). *Canadian Journal of Fisheries and Aquatic Sciences*,
458 49(5), 894–902. <https://doi.org/10.1139/f92-100>

459 Comte, L., & Grenouillet, G. (2013). Do stream fish track climate change? Assessing distribution shifts
460 in recent decades. *Ecography*, 36(11), 1236–1246. <https://doi.org/10.1111/j.1600->
461 0587.2013.00282.x

462 Copp, G. H., Godard, M. J., Russell, I. C., Peeler, E. J., Gherardi, F., Tricarico, E., ... Britton, J. R.
463 (2014). A preliminary evaluation of the European Non-native Species in Aquaculture Risk

464 Assessment Scheme applied to species listed on Annex IV of the EU Alien Species Regulation.
465 *Fisheries Management and Ecology*, 23(1), 12–20. <https://doi.org/10.1111/fme.12076>

466 Copp, G. H., Russell, I. C., Peeler, E. J., Gherardi, F., Tricarico, E., Macleod, A., ... Savini, D. (2016).
467 European Non-native Species in Aquaculture Risk Analysis Scheme—a summary of assessment
468 protocols and decision support tools for use of alien species in aquaculture. *Fisheries*
469 *Management and Ecology*, 23(1), 1–11. <https://doi.org/10.1111/fme.12074>

470 Copp, G. H., Wesley, K. J., Kovac, V., Ives, M. J., & Carter, M. G. (2003). Introduction and
471 establishment of the pikeperch *Stizostedion lucioperca*(L.) in Stanborough Lake (Hertfordshire)
472 and its dispersal in the Thames catchment. *The London Naturalist*, (82), 139–154.

473 Cucherousset, J., Copp, G. H., Fox, M. G., Sterud, E., van Kleef, H. H., Verreycken, H., & Záhorská,
474 E. (2009). Life-history traits and potential invasiveness of introduced pumpkinseed *Lepomis*
475 *gibbosus* populations in northwestern Europe. *Biological Invasions*, 11(9), 2171.
476 <https://doi.org/10.1007/s10530-009-9493-5>

477 Cucherousset, J., & Olden, J. D. (2011). Ecological Impacts of Nonnative Freshwater Fishes. *Fisheries*,
478 36(5), 215–230. <https://doi.org/10.1080/03632415.2011.574578>

479 Elvira, B., & Almodóvar, A. (2001). Freshwater fish introductions in Spain: facts and figures at the
480 beginning of the 21st century. *Journal of Fish Biology*, 59(sA), 323–331.
481 <https://doi.org/10.1111/j.1095-8649.2001.tb01393.x>

482 Eschbach, E., Nolte, A. W., Kohlmann, K., Kersten, P., Kail, J., & Arlinghaus, R. (2014). Population
483 differentiation of zander (*Sander lucioperca*) across native and newly colonized ranges suggests
484 increasing admixture in the course of an invasion. *Evolutionary Applications*, 7(5), 555–568.
485 <https://doi.org/10.1111/eva.12155>

- 486 Fickling, N. J., & Lee, R. L. G. (1983). A Review of the Ecological Impact of the Introduction of the
487 Zander (*Stizostedion lucioperca* L.) into Waters of the Eurasian Mainland. *Aquaculture*
488 *Research*, 14(3), 151–155. <https://doi.org/10.1111/j.1365-2109.1983.tb00065.x>
- 489 Francis, R. I. C. C. (1990). Back-calculation of fish length: a critical review. *Journal of Fish Biology*,
490 36(6), 883–902. <https://doi.org/10.1111/j.1095-8649.1990.tb05636.x>
- 491 Froese, R., & Pauly, D. (2018, June). FishBase. World Wide Web electronic publication. Retrieved
492 from www.fishbase.org
- 493 Gozlan, R. E., Andreou, D., Asaeda, T., Beyer, K., Bouhadad, R., Burnard, D., ... Esmaeili, H. R.
494 (2010). Pan-continental invasion of *Pseudorasbora parva*: towards a better understanding of
495 freshwater fish invasions. *Fish and Fisheries*, 11(4), 315–340. [https://doi.org/10.1111/j.1467-](https://doi.org/10.1111/j.1467-2979.2010.00361.x)
496 [2979.2010.00361.x](https://doi.org/10.1111/j.1467-2979.2010.00361.x)
- 497 Gozlan, R. E., Britton, J. R., Cowx, I., & Copp, G. H. (2010). Current knowledge on non-native
498 freshwater fish introductions. *Journal of Fish Biology*, 76(4), 751–786.
499 <https://doi.org/10.1111/j.1095-8649.2010.02566.x>
- 500 Graham, C. T., & Harrod, C. (2009). Implications of climate change for the fishes of the British Isles.
501 *Journal of Fish Biology*, 74(6), 1143–1205. <https://doi.org/10.1111/j.1095-8649.2009.02180.x>
- 502 Gratwicke, B., & Marshall, B. E. (2001). The relationship between the exotic predators *Micropterus*
503 *salmoides* and *Serranochromis robustus* and native stream fishes in Zimbabwe. *Journal of Fish*
504 *Biology*, 58(1), 68–75. <https://doi.org/10.1111/j.1095-8649.2001.tb00499.x>
- 505 Haugen, T. O., Winfield, I. J., Vøllestad, L. A., Fletcher, J. M., James, J. B., & Stenseth, N. C. (2007).
506 Density Dependence and Density Independence in the Demography and Dispersal of Pike Over
507 Four Decades. *Ecological Monographs*, 77(4), 483–502. <https://doi.org/10.1890/06-0163.1>
- 508 Heibo, E., Magnhagen, C., & Vøllestad, L. A. (2005). Latitudinal variation in life-history traits in
509 Eurasian perch. *Ecology*, 86(12), 3377–3386. <https://doi.org/10.1890/04-1620>

510 Hickley, P. (1986). Invasion by Zander and the Management of Fish Stocks. *Philosophical*
511 *Transactions of the Royal Society of London B: Biological Sciences*, 314(1167), 571–582.
512 <https://doi.org/10.1098/rstb.1986.0073>

513 Hickley, P., & Chare, S. (2004). Fisheries for non-native species in England and Wales: angling or the
514 environment? *Fisheries Management and Ecology*, 11(3–4), 203–212.
515 <https://doi.org/10.1111/j.1365-2400.2004.00395.x>

516 Johnson, A. C., Acreman, M. C., Dunbar, M. J., Feist, S. W., Giacomello, A. M., Gozlan, R. E., ...
517 Jones, J. I. (2009). The British river of the future: how climate change and human activity might
518 affect two contrasting river ecosystems in England. *Science of the Total Environment*, 407(17),
519 4787–4798. <https://doi.org/10.1016/j.scitotenv.2009.05.018>

520 Keskinen, T., & Marjomäki, T. J. (2003). Growth of pikeperch in relation to lake characteristics: total
521 phosphorus, water colour, lake area and depth. *Journal of Fish Biology*, 63(5), 1274–1282.
522 <https://doi.org/10.1046/j.1095-8649.2003.00249.x>

523 Kopp, D., Cucherousset, J., Syväranta, J., Martino, A., Céréghino, R., & Santoul, F. (2009). Trophic
524 ecology of the pikeperch (*Sander lucioperca*) in its introduced areas: a stable isotope approach
525 in southwestern France. *Comptes Rendus Biologies*, 332(8), 741–746.
526 <https://doi.org/10.1016/j.crv.2009.04.001>

527 Kuczynski, L., Legendre, P., & Grenouillet, G. (2018). Concomitant impacts of climate change,
528 fragmentation and non-native species have led to reorganization of fish communities since the
529 1980s. *Global Ecology and Biogeography*, 27(2), 213–222. <https://doi.org/10.1111/geb.12690>

530 Langsrud, Ø. (2003). ANOVA for unbalanced data: Use Type II instead of Type III sums of squares.
531 *Statistics and Computing*, 13(2), 163–167. <https://doi.org/10.1023/A:1023260610025>

- 532 Lappalainen, J., Dörner, H., & Wysujack, K. (2003). Reproduction biology of pikeperch (*Sander*
533 *lucioperca* (L.))—a review. *Ecology of Freshwater Fish*, 12(2), 95–106.
534 <https://doi.org/10.1034/j.1600-0633.2003.00005.x>
- 535 Lappalainen, J., Malinen, T., Rahikainen, M., Vinni, M., Nyberg, K., Ruuhijärvi, J., & Salminen, M.
536 (2005). Temperature dependent growth and yield of pikeperch, *Sander lucioperca*, in Finnish
537 lakes. *Fisheries Management and Ecology*, 12(1), 27–35. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2400.2004.00416.x)
538 [2400.2004.00416.x](https://doi.org/10.1111/j.1365-2400.2004.00416.x)
- 539 Lappalainen, J., Milardi, M., Nyberg, K., & Venäläinen, A. (2009). Effects of water temperature on
540 year-class strengths and growth patterns of pikeperch (*Sander lucioperca* (L.)) in the brackish
541 Baltic Sea. *Aquatic Ecology*, 43(1), 181–191. <https://doi.org/10.1007/s10452-007-9150-y>
- 542 Lappalainen, J., Tarkan, A. S., & Harrod, C. (2008). A meta-analysis of latitudinal variations in life-
543 history traits of roach, *Rutilus rutilus*, over its geographical range: linear or non-linear
544 relationships? *Freshwater Biology*, 53(8), 1491–1501. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2427.2008.01977.x)
545 [2427.2008.01977.x](https://doi.org/10.1111/j.1365-2427.2008.01977.x)
- 546 Linfield, R. S. J., & Rickards, R. B. (1979). The Zander in Perspective. *Aquaculture Research*, 10(1),
547 1–16. <https://doi.org/10.1111/j.1365-2109.1979.tb00249.x>
- 548 Ložys, L. (2004). The growth of pikeperch (*Sander lucioperca* L.) and perch (*Perca fluviatilis* L.)
549 under different water temperature and salinity conditions in the Curonian Lagoon and
550 Lithuanian coastal waters of the Baltic Sea. *Hydrobiologia*, 514(1–3), 105–113.
551 <https://doi.org/10.1023/B:hydr.0000018211.26378.b9>
- 552 Magnuson, J. J., Crowder, L. B., & Medvick, P. A. (1979). Temperature as an ecological resource.
553 *American Zoologist*, 19(1), 331–343. <https://doi.org/10.1093/icb/19.1.331>
- 554 Maitland, P. S. (2004). *Keys to the Freshwater Fish of Britain and Ireland, with Notes on their*
555 *Distribution and Ecology*. Ambleside, UK: Freshwater Biological Association (FBA).

556 Milardi, M., Lappalainen, J., Malinen, T., Vinni, M., & Ruuhijärvi, J. (2011). Problems in managing a
557 slow-growing pikeperch (*Sander lucioperca* (L.)) population in Southern Finland. *Knowledge*
558 *and Management of Aquatic Ecosystems*, (400), 08. <https://doi.org/10.1051/kmae/2011010>

559 Mittelbach, G. G., & Persson, L. (1998). The ontogeny of piscivory and its ecological consequences.
560 *Canadian Journal of Fisheries and Aquatic Sciences*, 55(6), 1454–1465.
561 <https://doi.org/10.1139/f98-041>

562 Mooij, W. M., Lammens, E., & Densen, W. V. (1994). Growth rate of 0+ fish in relation to
563 temperature, body size, and food in shallow eutrophic Lake Tjeukemeer. *Canadian Journal of*
564 *Fisheries and Aquatic Sciences*, 51(3), 516–526. <https://doi.org/10.1139/f94-054>

565 Nunn, A. D., Bolland, J. D., Harvey, J. P., & Cowx, I. G. (2007). Establishment of self-sustaining
566 populations of non-native fish species in the River Trent and Warwickshire Avon, UK,
567 indicated by the presence of 0+ fish. *Aquatic Invasions*, 2(3), 190–196.
568 <https://doi.org/10.3391/ai.2007.2.3.6>

569 Nyberg, P., Degerman, E., & Sers, B. (1996). Survival after catch in trap-nets, movements and growth
570 of the pikeperch (*Stizostedion lucioperca*) in Lake Hjälmaren, Central Sweden. In *Annales*
571 *Zoologici Fennici* (pp. 569–575). JSTOR. Retrieved from <http://www.jstor.org/stable/23736102>

572 Nyina-wamwiza, L., Xu, X. L., Blanchard, G., & Kestemont, P. (2005). Effect of dietary protein, lipid
573 and carbohydrate ratio on growth, feed efficiency and body composition of pikeperch *Sander*
574 *lucioperca* fingerlings. *Aquaculture Research*, 36(5), 486–492. [https://doi.org/10.1111/j.1365-](https://doi.org/10.1111/j.1365-2109.2005.01233.x)
575 [2109.2005.01233.x](https://doi.org/10.1111/j.1365-2109.2005.01233.x)

576 Olden, J. D., Poff, N. L., & Bestgen, K. R. (2006). Life-history strategies predict fish invasions and
577 extirpations in the Colorado River Basin. *Ecological Monographs*, 76(1), 25–40.
578 <https://doi.org/doi.org/10.1890/05-0330>

- 579 Oyugi, D. O., Cucherousset, J., Ntiba, M. J., Kisia, S. M., Harper, D. M., & Britton, J. R. (2011). Life
580 history traits of an equatorial common carp *Cyprinus carpio* population in relation to thermal
581 influences on invasive populations. *Fisheries Research*, *110*(1), 92–97.
582 <https://doi.org/10.1016/j.fishres.2011.03.017>
- 583 Özvarol, Z. A. B., & İkiz, R. (2008). Growth, mortality and stock analysis of the pikeperch, *Sander*
584 *luciperca* (L., 1758) population of Karacaören I Dam Lake. *Journal of Fisheries Sciences*.
585 *Com*, *2*(2), 134–145.
- 586 Pardo, S.A., Cooper, A.B. & Dulvy, N.K. (2013) Avoiding fishy growth curves. *Methods in Ecology*
587 *and Evolution*, *4*, 353-360.
- 588 Pelicice, F. M., & Agostinho, A. A. (2009). Fish fauna destruction after the introduction of a non-native
589 predator (*Cichla kelberi*) in a Neotropical reservoir. *Biological Invasions*, *11*(8), 1789–1801.
590 <https://doi.org/10.1007/s10530-008-9358-3>
- 591 Pérez-Bote, J. L., & Roso, R. (2012). Growth and length–weight relationships of *Sander luciperca*
592 (Linnaeus, 1758) in the Alcántara Reservoir, south-western Spain: comparison with other water
593 bodies in Eurasia. *Journal of Applied Ichthyology*, *28*(2), 264–268.
594 <https://doi.org/10.1111/j.1439-0426.2011.01918.x>
- 595 Pinder, L. C. V., Marker, A. F. H., Mann, R. H. K., Bass, J. A. B., & Copp, G. H. (1997). The River
596 Great Ouse, a highly eutrophic, slow-flowing, regulated, lowland river in eastern England.
597 *River Research and Applications*, *13*(3), 203–218. [https://doi.org/10.1002/\(SICI\)1099-](https://doi.org/10.1002/(SICI)1099-1646(199705)13:3<203::AID-RRR449>3.0.CO;2-F)
598 [1646\(199705\)13:3<203::AID-RRR449>3.0.CO;2-F](https://doi.org/10.1002/(SICI)1099-1646(199705)13:3<203::AID-RRR449>3.0.CO;2-F)
- 599 R Core Team. (2018). R: A language and environment for statistical computing. R Foundation for
600 Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.

601 Rahel, F. J., & Olden, J. D. (2008). Assessing the Effects of Climate Change on Aquatic Invasive
602 Species. *Conservation Biology*, 22(3), 521–533. [https://doi.org/10.1111/j.1523-](https://doi.org/10.1111/j.1523-1739.2008.00950.x)
603 [1739.2008.00950.x](https://doi.org/10.1111/j.1523-1739.2008.00950.x)

604 Ribeiro, F., Elvira, B., Collares-Pereira, M. J., & Moyle, P. B. (2008). Life-history traits of non-native
605 fishes in Iberian watersheds across several invasion stages: a first approach. *Biological*
606 *Invasions*, 10(1), 89–102. <https://doi.org/10.1007/s10530-007-9112-2>

607 Ribeiro, F., Gante, H. F., Sousa, G., Filipe, A. F., Alves, M. J., & Magalhaes, M. F. (2009). New
608 records, distribution and dispersal pathways of *Sander lucioperca* in Iberian freshwaters.
609 *Cybium*, 33(3): 255–256.

610 Ruiz-Navarro, A., Gillingham, P. K., & Britton, J. R. (2016a). Predicting shifts in the climate space of
611 freshwater fishes in Great Britain due to climate change. *Biological Conservation*, 203, 33–42.
612 <https://doi.org/10.1016/j.biocon.2016.08.021>

613 Ruiz-Navarro, A., Gillingham, P. K., & Britton, J. R. (2016b). Shifts in the climate space of temperate
614 cyprinid fishes due to climate change are coupled with altered body sizes and growth rates.
615 *Global Change Biology*, 22(9), 3221–3232. <https://doi.org/10.1111/gcb.13230>

616 Rypel, A. L. (2012). Meta-analysis of growth rates for a circumpolar fish, the northern pike (*Esox*
617 *lucius*), with emphasis on effects of continent, climate and latitude. *Ecology of Freshwater Fish*,
618 21(4), 521–532. <https://doi.org/10.1111/j.1600-0633.2012.00570.x>

619 Smith, P. A., Leah, R. T., & Eaton, J. W. (1998). A review of the current knowledge on the
620 introduction, ecology and management of zander, *Stizostedion lucioperca*, in the UK. In
621 *Stocking and Introduction of Fish* (Cowx IG, pp. 209–224). Oxford: Blackwell Science.

622 Tarkan, A.S. & Vilizzi, L. (2015). Patterns, latitudinal clines and counter-gradient variation in the
623 growth of roach *Rutilus rutilus* (Cyprinidae) in its Eurasian area of distribution. *Reviews in Fish*
624 *Biology and Fisheries*, 25, 587-602.

- 625 Venturelli, P. A., Lester, N. P., Marshall, T. R., & Shuter, B. J. (2010). Consistent patterns of maturity
626 and density-dependent growth among populations of walleye (*Sander vitreus*): application of
627 the growing degree-day metric. *Canadian Journal of Fisheries and Aquatic Sciences*, 67(7),
628 1057–1067. <https://doi.org/10.1139/F10-041>
- 629 Vila-Gispert, A., Alcaraz, C., & García-Berthou, E. (2005). Life-history traits of invasive fish in small
630 Mediterranean streams. *Biological Invasions*, 7(1), 107–116. [https://doi.org/10.1007/s10530-](https://doi.org/10.1007/s10530-004-9640-y)
631 [004-9640-y](https://doi.org/10.1007/s10530-004-9640-y)
- 632 Vilizzi, L. & Copp, G.H. (2017). Global patterns and clines in the growth of common carp *Cyprinus*
633 *carpio*. *Journal of Fish Biology*, 91, 3-40.
- 634 Wheeler, A. (1974). Changes in the freshwater fish fauna of Britain. *Changing Flora and Fauna of*
635 *Britain*. DL Hawksworth, Ed.
- 636 Wheeler, A. & Maitland, P. S. (1973). The scarcer freshwater fishes of the British Isles. *Journal of*
637 *Fish Biology*, 5(1), 49–68. <https://doi.org/10.1111/j.1095-8649.1973.tb04430.x>
- 638 Wilby, R. L., Whitehead, P. G., Wade, A. J., Butterfield, D., Davis, R. J., & Watts, G. (2006).
639 Integrated modelling of climate change impacts on water resources and quality in a lowland
640 catchment: River Kennet, UK. *Journal of Hydrology*, 330(1–2), 204–220.
641 <https://doi.org/10.1016/j.jhydrol.2006.04.033>
- 642 Živkov, M. T., Trichkova, T. A., & Raikova-Petrova, G. N. (1999). Biological reasons for the
643 unsuitability of growth parameters and indices for comparing fish growth. *Environmental*
644 *Biology of Fishes*, 54(1), 67–76. <https://doi.org/10.1023/A:1007425005491>

645 Table 1. Samples size, length and age range and Von Bertalanffy growth parameters estimates for invasive *Sander lucioperca* in England, values
 646 in parentheses represent the 95% confidence limits of each parameter estimate.

647

River	N	Length (mm)	Age (years)	L_{∞}	K	t_0
Gloucester-Sharpness Canal	18	345-660	2 - 8	980 (679, 2509)	0.13 (0.03, 0.36)	-1.10 (-3.94, 0.65)
Grand Union Canal	12	169-551	2 - 6	820 (598, 1726)	0.18 (0.05, 0.34)	-0.08 (-0.75, 0.46)
Lower Warwickshire Avon	9 26	90-695	0 - 11	870 (713, 1143)	0.16 (0.10, 0.25)	-0.82 (-1.23, -0.46)
Middle Warwickshire Avon	35	142-650	1 - 7	753 (676, 904)	0.22 (0.15, 0.30)	-0.08 (-0.54, 0.23)
Great Ouse	70	110-760	1 - 10	853 (725, 1138)	0.19 (0.11, 0.28)	-0.27 (-0.73, 0.03)
Severn	18	74-770	0 - 10	874 (806, 964)	0.18 (0.15, 0.21)	-0.43 (-0.62, -0.26)

648 Table 2. Linear and non-linear (2nd order polynomial) regression statistics for the relationship
 649 between von Bertalanffy growth parameters L_{∞} and K versus latitude and longitude.

650 *significant at $P \leq 0.05$; **significant at $P \leq 0.01$.

651

Relationship	Model	R ²	F	P	AIC
Latitude vs. L_{∞}	Linear	0.01	(1,23) 1.23	0.28	310.61
	Non-linear	0.16	(2, 22) 3.36	0.05*	307.25
Longitude vs. L_{∞}	Linear	0.04	(1,23) 2.07	0.16	309.76
	Non-linear	0.08	(2, 22) 2.13	0.14	309.50
Latitude vs. K	Linear	0.29	(1,23) 10.71	0.01**	-77.85
	Non-linear	0.43	(2, 22) 10.06	< 0.01**	-82.84
Longitude vs. K	Linear	-0.01	(1,23) 0.99	0.33	-69.34
	Non-linear	0.07	(2, 22) 1.87	0.18	-70.24

652 **Figure captions**

653

654 Figure 1. Fitted von Bertalanffy growth curve for populations of *Sander lucioperca* at sites in
655 England including the Gloucester-Sharpness canal (dotted line), the Grand Union canal (dot-
656 dash line), the lower Warwickshire Avon (dashed line), the middle Warwickshire Avon (two
657 dash line) the River Great Ouse (solid line) and the river Severn (long dash line).

658

659 Figure 2. Mean standardised growth residuals for (a) juvenile growth rate; and (b) length at
660 last annulus for the Gloucester-Sharpness canal (GSC), Grand Union Canal (GUC), the lower
661 Warwickshire Avon (L. Avon), the middle Warwickshire Avon (M. Avon), the river Great
662 Ouse (Ouse) and the river Severn (Severn). Error bars represent the upper and lower 95%
663 confidence limits.

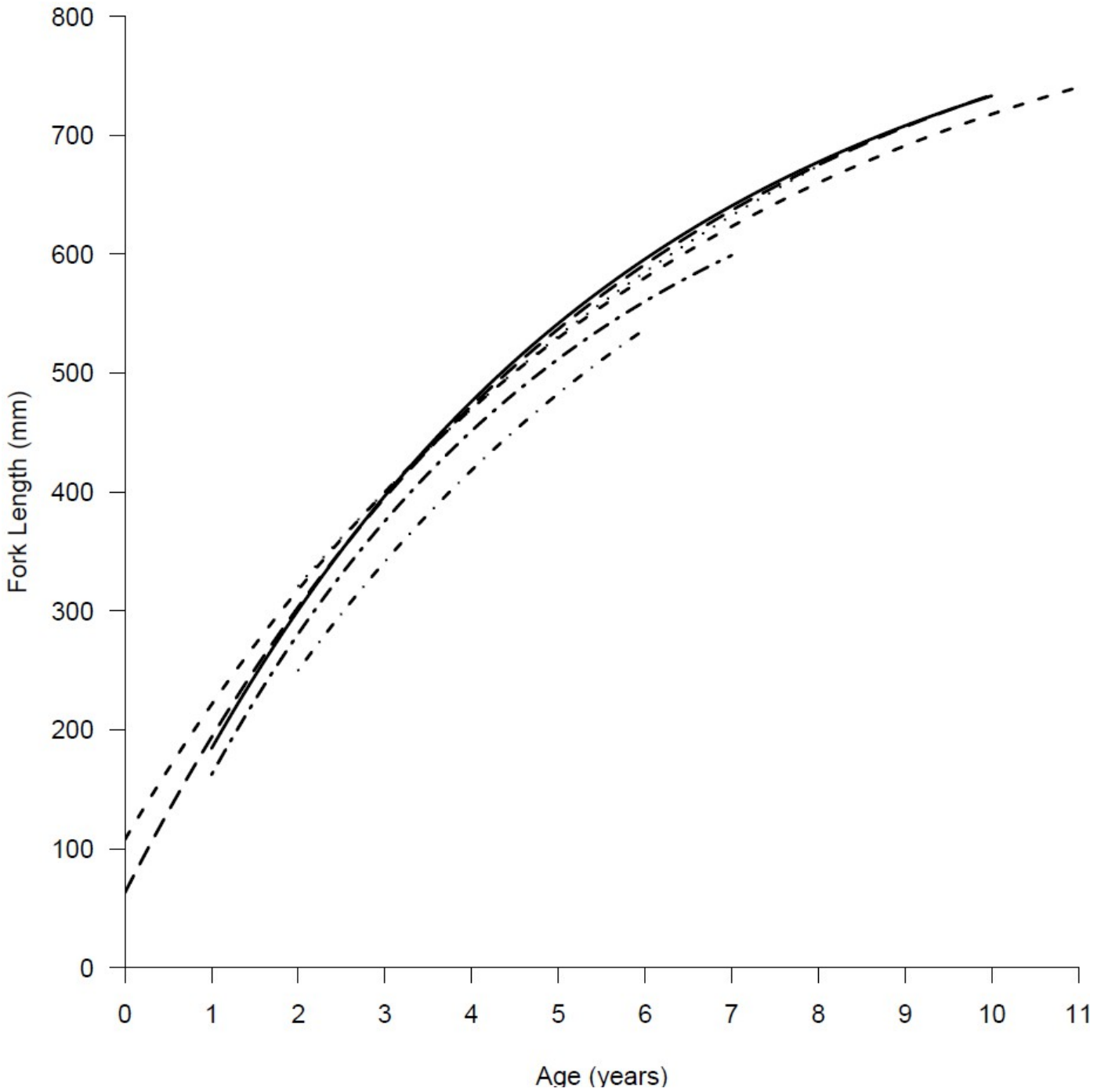
664

665 Figure 3. Relationship of L_{∞} (as fork length) and K of the von Bertalanffy growth model for
666 populations of *Sander lucioperca*, where the solid line is the significant relationship between
667 the variables according to linear regression, filled circles represent values extracted from
668 literature whilst open circles represent values from invasive populations in England derived
669 in this study.

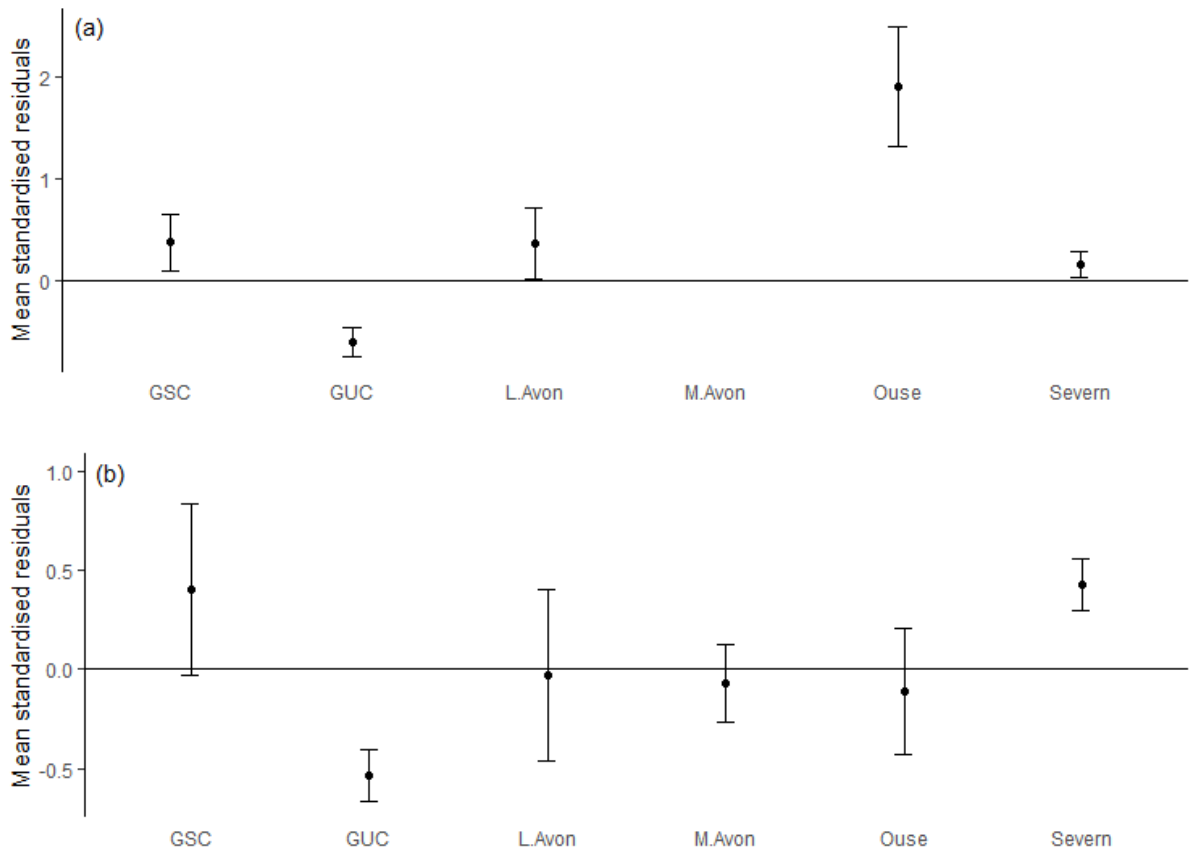
670

671 Figure 4. Relationships of latitude and longitude with L_{∞} (as fork length) and K of the von
672 Bertalanffy growth model for *Sander lucioperca*. The solid line represents the significant
673 relationship between the variables according to polynomial regression (2nd order), filled
674 circles represent values extracted from literature whilst open circles represent values from
675 invasive populations in England derived in this study and retained in the meta-analysis.

676



677 Figure 1.



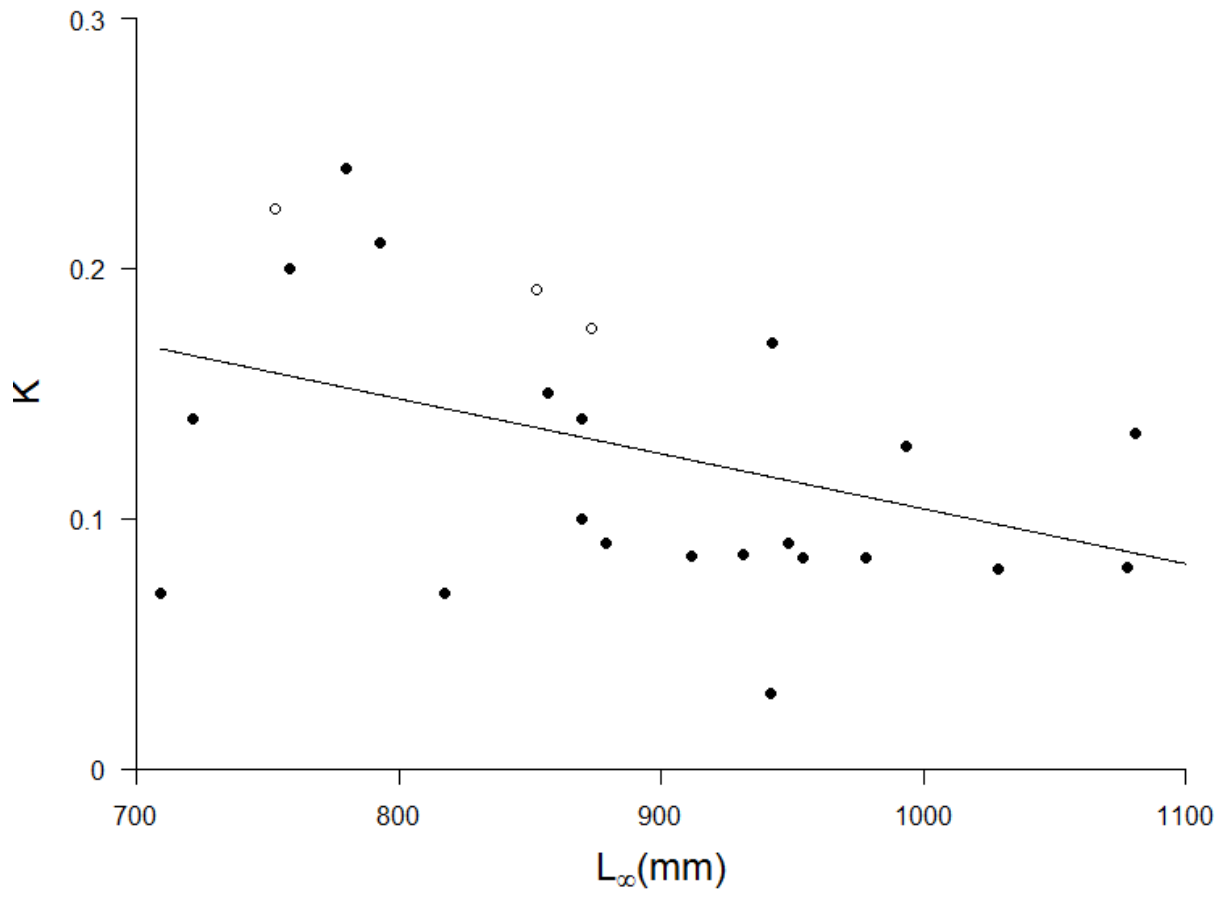
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679 Figure 2.

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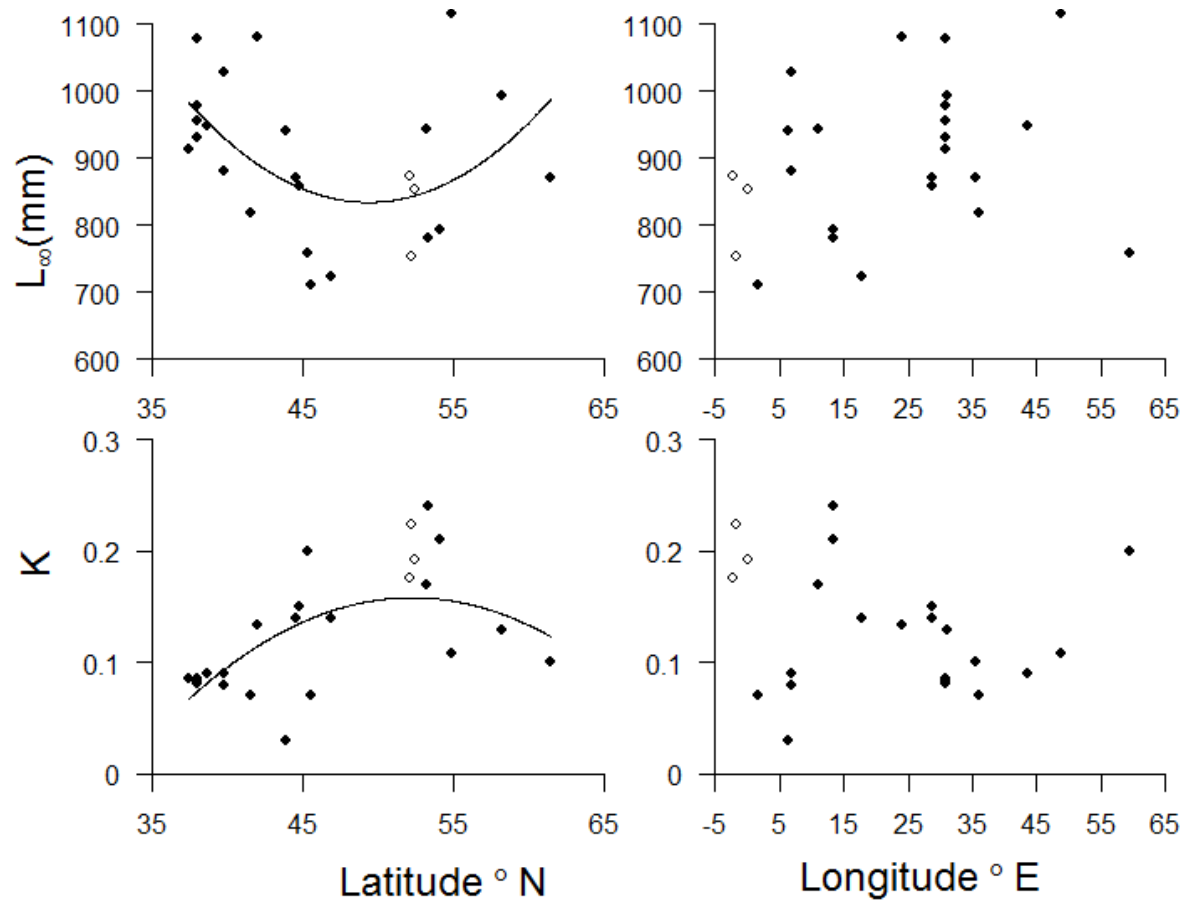
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683

684 Figure 3.

685



686

687 Figure 4.