

1   **Predicting the ecological impacts of an alien invader: experimental approaches**  
2   **reveal the trophic consequences of competition**

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15    **Abstract**

16

17    1. Ecological theory on the trophic impacts of invasive fauna on native  
18    competitors is equivocal. While increased inter-specific competition can result  
19    in coexisting species having constricted and diverged trophic niches, the  
20    competing species might instead increase their niche sizes to maintain energy  
21    intakes. Empirical experiments can test invasion theory on competitive  
22    interactions and niche sizes across different spatial scales and complexity.

23

24    2. The consequences of increased inter-specific competition from a model alien  
25    fish *Leuciscus idus* were tested on two taxonomically and trophically similar  
26    native fishes, *Squalius cephalus* and *Barbus barbus*. Competitive interactions  
27    were tested in tank aquaria using comparative functional responses (CFRs)  
28    and cohabitation trials. The consequences of these competitive interactions for  
29    the trophic niche sizes and positions of the fishes were tested in pond  
30    mesocosms.

31

32    3. CFRs revealed that compared to *B. barbus*, *L. idus* had significantly higher  
33    attack and consumption rates; cohabitation trials revealed *B. barbus* growth  
34    rates were depressed in sympatry with *L. idus*. For *L. idus* and *S. cephalus*,  
35    differences in their functional response parameters and growth rates were not  
36    significant.

37

38    4. Pond mesocosms used stable isotope metrics to quantify shifts in the trophic  
39    niche sizes of the fishes between allopatry and sympatry using a substitutive

40 experimental design. Isotopic niches were smaller and more divergent in  
41 sympatric paired species than predicted by their allopatric treatments,  
42 suggesting trophic impacts from inter-specific competition. However, an all-  
43 species sympatric treatment revealed similar niche sizes with allopatry. This  
44 maintenance of niche sizes in the presence of all species potentially resulted  
45 from the buffering of direct competitive effects of the species-pairs by indirect  
46 effects.

47

48 5. Experimental predictions from tank aquaria assisted the interpretation of the  
49 constricted and diverged trophic niches detected in the paired-species  
50 sympatric treatments of the pond mesocosms. However, the all-species  
51 sympatric treatment of this experiment revealed greater complexity in the  
52 outcomes of the competitive interactions within and between the species.  
53 These results have important implications for understanding how alien species  
54 integrate into food webs and influence the trophic relationships between native  
55 species.

56

57 **Key words:** Comparative functional response; inter-specific competition, invasive  
58 species, non-native, predator-prey.

59    **Introduction**

60

61    The ecological impacts of biological invasions are wide ranging and include habitat  
62    disruption and genetic introgression with native species (Gozlan et al. 2010).  
63    Ecological impacts can also develop through the trophic interactions of the invader  
64    with native species, including via predator-prey relationships (Dick et al. 2013;  
65    Alexander et al. 2014) and competitive interactions with other consumers (Britton et  
66    al. 2018). The intensity of competitive interactions and so the severity of their impacts  
67    are predicted to be stronger and more intense when the invader and native species are  
68    taxonomically and/ or trophically similar due to their likelihood of exploiting similar  
69    prey resources (Dick et al. 2017).

70

71    Ecological theory can help predict the trophic consequences of biological invasions  
72    (Britton et al. 2018). Hypotheses on trophic niche theory suggest how alien and native  
73    species can coexist in food webs (Catford, Jansson & Nilsson 2009). If the alien  
74    species utilises resources that are unlimited or unexploited by native species, there  
75    will be little change in the competitive pressures of the invaded system, enabling the  
76    co-existence of species (Mason et al. 2008; Juncos et al. 2015). Should competitive  
77    interactions be more intense due to the alien species exploiting similar and limited  
78    prey resources to native species, their niches could constrict in size as the diets of  
79    each species becomes more specialized (e.g. Tran et al. 2015; Jackson et al. 2016).  
80    These smaller niches might also be divergent if the species exploit alternative  
81    resources to minimize their competitive interactions (Busst & Britton 2017; Britton et  
82    al. 2018). Competitive exclusion of native species from their original niche could  
83    occur if the inter-specific competitive interactions are particularly intense and

84 asymmetric (Bøhn, Amundsen & Sparrow 2008). Conversely, if species diversify  
85 their diet in response to increased competition then their niches might increase in size  
86 (Britton et al. 2018). The intensity of intra-specific competition can also have  
87 considerable influences on trophic niche sizes, with optimal foraging theory  
88 predicting that as it intensifies, niche breadths will increase as individuals diversify  
89 their diet in response to resource depletion (Svanbäck & Bolnick 2006). Moreover, as  
90 competitive interactions are important for structuring the populations of many taxa  
91 then understanding how alien species compete with native biota and integrate into  
92 native food webs is integral to understanding their ecological impacts (Riccardi et al.  
93 2013; Gallardo et al. 2016).

94

95 Across taxa, it remains equivocal as to how these potential shifts in the trophic niches  
96 of native species manifest following an invasion (Britton et al. 2018) and so can be  
97 investigated further using empirical experiments. Manipulating the abundances of  
98 alien and native species enables the outcomes of the altered strength of their  
99 competitive interactions to be measured (Britton 2018). For example, cohabitation  
100 pond mesocosm experiments can compare the results of inter-specific competition  
101 between sympatric alien and native fishes versus allopatric treatments involving only  
102 intra-specific competition (Britton 2018). Alterations in niche sizes and trophic  
103 positions between allopatry and sympathy can be quantified by stable isotope metrics  
104 (Tran et al. 2015; Britton et al. 2018). The competitive relationships between the  
105 species can then be informed by aquaria experiments (Britton 2018). Cohabitation  
106 aquaria experiments can utilise the same species as pond experiments, but under  
107 controlled conditions (Busst & Britton 2016), where higher growth rates within  
108 species indicates higher resource acquisition and greater competitive ability (Ward,

109 Webster & Hart 2006). Comparative functional response experiments (CFRs)  
110 compare consumption rates as a function of prey density between the alien and native  
111 species (Dick et al. 2013, 2014, 2017). A species with a significantly higher  
112 consumption rate than a comparator species has the ability to acquire more resources,  
113 i.e. their inter-specific interactions will be asymmetric.

114

115 The aim here was to use these experimental approaches to empirically predict the  
116 trophic impacts of an invasion by a model alien freshwater fish on two trophically and  
117 taxonomically similar native fishes. The model area was Great Britain, a temperate  
118 region where the model alien fish, ide *Leuciscus idus*, is non-native. The species is,  
119 however, present in many lentic environments due to introductions of hatchery reared  
120 fish for angling, despite risk assessment suggesting their invasion risk is high in  
121 Britain (Britton et al. 2010). It has yet to disperse widely in lotic environments. The  
122 species is also taxonomically similar to chub *Squalius cephalus* (synonym: *Leuciscus*  
123 *cephalus*), a native riverine species that tends to coexist with the trophically similar  
124 European barbel *Barbus barbus* (Gutmann Roberts & Britton 2018). Consequently, *S.*  
125 *cephalus* and *B. barbus* were the model native fishes. As CFRs tend to predict that  
126 high-risk alien species have significantly higher consumption rates than native  
127 analogues (Dick et al. 2013), it was predicted that: (i) inter-specific competition  
128 between the alien and native fishes would be asymmetric, with *L. idus* the superior  
129 competitor; and (ii) this asymmetric competition would result in the native fishes  
130 having reduced niche sizes and growth rates when in sympatry compared to allopatry,  
131 but with *L. idus* having niche sizes and growth rates similar between allopatry and  
132 sympatry.

133   **Materials and Methods**

134

135   ***Model fishes***

136   The three model fishes are all species in the Cyprinidae family that are either benthic  
137   or benthopelagic foragers. Although primarily lotic fishes, they are all also present in  
138   a range of lentic habitats (e.g. Jurajda, Ondračková & Reichard 2004; Taylor et al.  
139   2004). Whilst their diets typically comprise of macroinvertebrates, plant material can  
140   also be an important food source (Brabrand 1985; Balestrieri et al. 2006; Caffrey et al.  
141   2008). In all experiments, *L. idus*, *S. cephalus* and *B. barbus* were sourced from an  
142   aquaculture site in Southern England, with all fish of age 1+ years and 65 to 80 mm  
143   starting length (individuals of different lengths were randomly distributed across the  
144   experiments). All fish were tagged with 7 mm passive integrated transponder tags  
145   (approximate weight: 0.03 g) to enable individual identification. Fish were weighed  
146   post-tagging (to 0.1 g). These fish had been pond-reared on a diet of natural and  
147   formulated feeds. For aquaria-based experiments, the fish were allowed to acclimate  
148   to the aquaria conditions for 28 days at 20 °C before use. In the aquaria, fish were  
149   held in 45 L tanks where water filtration was provided via flow-through systems.  
150   When not being used experimentally, the fish were fed a formulated feed based on  
151   plant material to standardize prior experience. As different batches of fish were used  
152   in each experiment, the fish used in the experimental treatments and replicates were  
153   all of similar length and mass to eliminate experimental confounds based on  
154   differences in body sizes.

155

156

157    ***Comparative functional responses (CFRs)***

158    The prey species used in the CFRs were *Gammarus pulex* and Chironomid larvae. In  
159    the experiments, individual fish were randomly selected 24 h prior to use and  
160    allocated to 10 L experimental tanks at 20 °C supplied with oxygen to provide  
161    constant conditions. They were without food in this period to standardize hunger  
162    levels. Individual fish were then presented with a prey species at one of six densities  
163    (2, 4, 8, 16, 32 and 64), with a minimum of three replicates generated per density and  
164    prey species. Prey exposure was for one hour. The fish were then removed from the  
165    tank, the number of prey remaining counted, and the number of prey consumed  
166    determined by subtracting this number from the original prey density.

167

168    In the CFRs, the comparisons were between the non-native *L. idus* versus the two  
169    native fishes. For *B. barbus* and *S. cephalus*, consumption rate data were as per Guo  
170    et al. (2017). The *L. idus* consumption rate data were generated at the same time as *B.*  
171    *barbus* and *S. cephalus*, but these data have not been used previously. Analyses of  
172    CFRs of all fishes were assessed using the integrated package for functional response  
173    analysis in R ('Frair') (Pritchard et al. 2017). Logistic regressions of prey density  
174    versus the proportion of prey consumed were performed per fish species, with type II  
175    functional responses indicated by significant negative first-order terms (Pritchard et  
176    al. 2017). Values of the attack rate (*a*) and handling time (*h*) were then obtained using  
177    maximum likelihood estimation (MLE) in the Random Predator Equation (Rogers  
178    1972), which assumes a Type II response and non-replacement of prey:

179     $N_e = N_0 (1 - \exp(a(N_e h - T)))$    (Equation 1)

180    where *N<sub>e</sub>* is the number of prey eaten, *N<sub>0</sub>* is the initial density of prey, *a* is the attack  
181    rate, *h* is the handling time and *T* is the total time available. Finally, to visualise the

182 uncertainty around the fitted functional responses, bootstrapping ( $n = 1500$ ) was used  
183 to construct empirical 95% confidence intervals of the fitted functional responses  
184 (Paterson et al. 2015). These bootstrapped data provided the CFR plots between the  
185 species; where there was overlap in their 95 % confidence limits, differences in the  
186 functional response curves were considered as not significant (Paterson et al. 2015).

187

188 ***Co-habitation aquaria experiments***

189 The cohabitation experiments in tank aquaria were completed in 45 L tanks arranged  
190 on shelving with three tiers (top, middle and bottom shelves) and completed at 18 °C  
191 on 16:8 h light:dark regime. Each species was used in allopatry ( $N = 10$ ) and then in  
192 each two-species sympatric combination ( $n = 5+5$ ), with three replicates per  
193 treatment. Feeding was once per day using a sinking, fishmeal based pellet (1.0 mm  
194 diameter; 45 % protein, 20 % oil) at a fixed ration of 2 % mean starting body mass per  
195 day. Prior to their release into the tanks, the starting weight of each species per  
196 treatment was measured. The experiment ran for 30 days.

197

198 At the end of the experimental period, the fish were removed from the tanks and re-  
199 weighed. The increase in mass per species and treatment during the experimental  
200 period was determined by the ‘specific growth rate’ (SGR):

201  $([(\ln W_{t+1}) - \ln W_t]/t)/n \times 100$  (Equation 2)

202 where  $W_t$  = total starting weight of the species in the tank,  $W_{t+1}$  = total finishing  
203 weight,  $n$  = number of fish, and  $t$  = number of days between  $W_t$  and  $W_{t+1}$ . Differences  
204 in SGR between treatments and species were tested in a linear mixed effects model.  
205 This tested the effect of the interaction of species x treatment on SGR, where tank  
206 position (i.e. whether it was on the top, middle or bottom shelf) was used as the

207 random variable and fish starting weight was used initially as a covariate. However,  
208 starting weight per species was removed from the final model as its effect was not  
209 significant ( $P > 0.05$ ). Model outputs were the overall significance of the model and  
210 the mean SGR values ( $\pm 95\%$  confidence intervals) according to species and  
211 treatment.

212

### 213 ***Co-habitation pond mesocosms***

214 The experimental design was based on substitutive treatments using allopatric and  
215 sympatric contexts. There were three allopatric treatments, where each species was  
216 used individually ( $N = 12$ ) and three sympatric treatments using paired species (*L.*  
217 *idus/ B. barbus*; *L. idus/ S. cephalus*; *B. barbus/ S. cephalus*;  $n = 6+6$ ). A final  
218 sympatric treatment then used the three fishes together ( $n = 4+4+4$ ). All treatments  
219 were replicated three times.

220

221 The experiment was completed using the treatments within enclosures as per Britton  
222 et al. (2018), with the enclosures sitting within a larger, man-made pond (30 x 30 m; 1  
223 m consistent depth) that was located in Southern England. The enclosures comprised  
224 of an aluminium frame (length 1.7 m; width: 1.1 m; height: 1.2 m) within a net (mesh:  
225 7 x 7 mm) that prevented fish ingress and egress, but allowed both movements of  
226 invertebrates and the growth of macrophytes into the enclosure. The enclosures were  
227 placed randomly across the pond, other than in shallow, littoral areas, with  
228 approximately 0.5 m between each enclosure. They remained *in-situ* throughout the  
229 experimental period. Their placement on top of the substrate enabled macrophytes to  
230 grow through them (*Elodea* spp.); all enclosures had similar areal macrophyte  
231 coverage during the experiment. Netting (15 x 15 mm mesh) over the enclosures

232 prevented bird predation. The experiment ran for 150 days from April 2017. This  
233 provided time for approximately four stable isotope half-lives in the fish dorsal  
234 muscle (i.e. at least 94 % isotopic turnover) (Thomas & Crowther 2015). Temperature  
235 loggers (TinyTag TGP-4017) in the larger pond revealed the mean water temperature  
236 was  $17.3 \pm 0.8$  °C during the experiment.

237

238 On day 150, all the fish were recovered from the enclosures, euthanized (anaesthetic  
239 overdose, MS-222) and taken to the laboratory. Samples of putative food resources  
240 were taken from the larger pond for stable isotope analysis (SIA) using a sweep net.  
241 These focused on the two major macroinvertebrate putative prey species sampled,  
242 *Gammarus pulex* and Chironomid larvae (that also ensured consistency with the  
243 CFRs). The presence of these macro-invertebrates was checked in each enclosure at  
244 the conclusion of the experiment, although their abundances were not accurately  
245 quantified. No other macro-invertebrates were detected in sufficient abundances  
246 inside or outside of enclosures to warrant their analysis; as their abundances were low  
247 outside of enclosures then their low abundance inside enclosures was not considered  
248 to be due to fish predation pressure. The other major food resource was plant material  
249 ('macrophyte') that was highly abundant in all enclosures, and was also sampled for  
250 SIA. All putative food resources were sorted into samples (one sample = 3 to 9  
251 individuals per species for macroinvertebrates), with triplicate samples analysed for  
252 each group.

253

254 In the laboratory, individuals were identified by their PIT tag and re-weighed,  
255 enabling calculation of their SGR (Equation 2). A dorsal muscle sample was taken for  
256 SIA. SI sample sizes were a minimum of 12 fish per species per treatment, with a

257 minimum of four fish taken randomly per replicate (Britton et al. 2018). All samples  
258 were dried at 60 °C to constant mass before SIA ( $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$ ) at the Cornell  
259 University Stable Isotope Laboratory, New York, USA. Prior to analysis, samples  
260 were ground to powder and weighed (approximately 1000 µg, but with precise  
261 measures taken) in tin capsules. They were then analysed on a Thermo Delta V  
262 isotope ratio mass spectrometer (Thermo Scientific, USA) interfaced to a NC2500  
263 elemental analyser (CE Elantach Inc., USA). Analytical precision associated with the  
264  $\delta^{15}\text{N}$  and  $\delta^{13}\text{C}$  sample runs was estimated at 0.42 and 0.15 ‰ respectively. Data  
265 outputs were in delta ( $\delta$ ) isotope ratios (‰). The C:N ratios of the analysed samples  
266 were between 3.15 and 3.61, indicating relatively low lipid levels (Post et al. 2007).  
267 These ratios did not differ significantly between experimental treatments  
268 (Supplementary material; Fig. S1). Comparison of original versus lipid-normalised  
269 data (Kiljunen et al. 2006) revealed a very strong and significant relationship,  
270 indicating that the variability in the original  $\delta^{13}\text{C}$  data was not an artefact of  
271 differences in lipid levels (Fig. S2). The shift between the mean original and mean  
272 normalised  $\delta^{13}\text{C}$  data per species and treatment was 0.61 to 0.69 ‰ (Table S1), thus  
273 had a negligible effect on the relative positions in isotopic space of the species per  
274 treatment. In addition, the lipid concentrations of the analysed fish tissues were not a  
275 significant predictor of their growth rates, i.e. faster growing fish did not have higher  
276 lipid concentrations (Fig. S3). Thus, the original  $\delta^{13}\text{C}$  data were used throughout all  
277 analyses, as lipid levels were not a confound in the experiment.

278  
279 The SI data were used to calculate the trophic niche size of each fish species per  
280 treatment using the isotopic niche (Jackson et al. 2011). Whilst closely related to the  
281 trophic niche, the isotopic niche is also influenced by factors including growth rate

282 and metabolism, and thus represents a close approximation of the trophic niche  
283 (Jackson et al. 2011). The isotopic niche was calculated using standard ellipse areas  
284 (SEA) in SIBER (Jackson et al. 2011; Jackson et al. 2012). This is a bivariate measure  
285 of the distribution of individuals in isotopic space, with the ellipses enclosing the core  
286 40 % of data, so indicates the typical resource use of the analysed population (Jackson  
287 et al. 2011). A Bayesian estimate of SEA ( $SEA_B$ ) tested differences in niche sizes  
288 between treatments per species, calculated using a Markov chain Monte Carlo  
289 simulation ( $10^4$  iterations per group) (Jackson et al. 2011; Jackson et al. 2012).  
290 Differences in the size of isotopic niches (as  $SEA_B$ ) were evaluated by calculating the  
291 probability that the relative posterior distributions of the niche size of the allopatric  
292 treatment were significantly smaller or larger than those of each of their sympatric  
293 niches ( $\alpha = 0.05$ ) in SIBER. The SI data were then used to calculate isotopic niche  
294 overlap (%) between the species using  $SEA_C$  also calculated in SIBER, where  
295 subscript ‘c’ indicates a small sample size correction was used (Jackson et al. 2012).  
296 Use of  $SEA_C$  was mainly to get a representation of the extent of niche overlap  
297 between species, as it is more strongly affected by small sample sizes (< 30) than  
298  $SEA_B$  (Jackson et al. 2012).

299  
300 The SI data were then applied to a Bayesian mixing model to predict the relative  
301 proportions of the three putative food resources to fish diet per treatment within the  
302 package ‘Mixing Models for Stable Isotope Analysis in R’ (MixSIAR; Stock et al.  
303 2018) Stock & Semmens 2016). The model ran using ‘short’ run length (chain length:  
304 50,000 iterations with burn-in of 25,000, with posterior thinning (thin: 25) and 3  
305 chains). Model diagnostics were based on Gelman-Rubin and Geweke, with sufficient  
306 convergence to accept the results (Stock & Semmens 2013). The isotopic

307 fractionation values between the prey resources and fish were  $\delta^{15}\text{N}$ :  $5.10 \pm 0.25\text{‰}$ ;  
308  $\delta^{13}\text{C}$ :  $3.8 \pm 0.25\text{‰}$ , based on the fractionation factors derived for *B. barbus* and *S.*  
309 *cephalus* values on controlled diets based on plant and invertebrate protein sources  
310 (Busst & Britton 2016). Mixing model results were reported as means of all feasible  
311 solutions, with 5 to 95<sup>th</sup> percentiles of the distribution ranges.

312

313 To assist evaluation of the competition strength within and between species in the  
314 treatments, the mean intra- and inter-specific isotopic dissimilarities were calculated  
315 (Calizza et al. 2017). For the mean intra-specific isotopic dissimilarity ( $\text{MND}_{ii}$ ), the  
316 first step was to calculate intraspecific isotopic dissimilarity ( $\text{ND}_{ii}$ ) for each individual  
317 fish per species and treatment, determined as the mean isotopic (Euclidean) distance  
318 between each individual and their conspecifics in the treatment. The mean  
319 intraspecific isotopic dissimilarity for each species per treatment was then taken as the  
320 mean  $\text{ND}_{ii}$  value of all specimens in that treatment; higher values indicate increased  
321 dissimilarity. The same process was followed to determine the mean inter-specific  
322 isotopic dissimilarity ( $\text{MND}_{ij}$ ) per species and treatment, except the first step was to  
323 calculate the mean isotopic distance of each individual fish from their sympatric  
324 species ( $\text{ND}_{ij}$ ) (Calizza et al. 2017).

325

326 The SI, predicted diet, isotopic dissimilarity and SGR data were then tested for  
327 differences between treatments. Differences in  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  and SGR were tested in  
328 linear mixed effects models (LMEM). Enclosure was used as a random effect on the  
329 intercept to avoid inflating the degrees of freedom that would occur if individual fish  
330 were used as true replicates (Tran et al. 2015). Total starting mass of fish in each  
331 enclosure was initially used as a covariate, but was removed from final models as it

332 was not significant ( $P > 0.05$ ). Outputs from the models were the mean  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  and  
333 SGR per species and treatment. The mean  $\delta^{13}\text{C}$ ,  $\delta^{15}\text{N}$  and SGR data from the models  
334 were then used to determine the extent of the change in each species between their  
335 allopatric treatment and each sympatric treatment. The extent of the change between  
336 allopatry and sympatry was then also determined for isotopic niche size (as SEAc)  
337 and the relative assimilation of each food resource from the mixing model outputs.  
338 These data were then tested for the significance of their relationships using linear  
339 regression. The relationships of  $\text{MND}_{ii}$  and  $\text{MND}_{ij}$  with SGR were also tested using  
340 linear regression to determine if changes in intra- and/ or inter-specific isotopic  
341 dissimilarity were significantly related to growth rates. Initially, multiple regression  
342 was used, where the mean isotopic dissimilarity that explained most of the SGR  
343 variability was indicated by the highest standardised  $\beta$  coefficient value; univariate  
344 linear regression was then used on both dissimilarity indices. Note that in these tests,  
345 only data from sympatric treatments were used, as  $\text{MND}_{ij}$  could only be determined  
346 for treatments involving at least two fish species.

347

348 Statistical analyses were performed in R (Version 3.5.2; R Development Core Team  
349 2018). In all results, error around the mean represents 95 % confidence limits. All  
350 experiments were completed following ethical review and under the UK Home Office  
351 project licence 70/8063.

352

353    **Results**

354

355    *Comparative functional responses*

356    In the functional response experiments, the first order linear coefficient from logistic  
357    regressions revealed the functional responses of all species were Type II and  
358    significant (first order linear coefficients from logistic regressions: *G. pulex*: -0.02, -  
359    0.04, and -0.06, Chironomid larvae: -0.02, -0.01 and -0.06, for *B. barbus*, *S. cephalus*  
360    and *L. idus* respectively;  $P < 0.01$  in all cases). For *B. barbus* versus *L. idus* using *G.*  
361    *pulex* as prey, *B. barbus* had a significantly lower attack rate ( $a$ ) and higher handling  
362    time ( $h$ ) than *L. idus* ( $a$ : 1.18 vs. 3.23,  $z = -2.64$ ,  $P < 0.01$ ;  $h$ : 0.12 vs. 0.06,  $z = 2.58$ ,  $P$   
363     $< 0.01$ ). On Chironomid larvae,  $h$  was also significantly higher for *B. barbus* (0.03 vs.  
364    0.04,  $z = 3.93$ ,  $P < 0.01$ ), but the difference in  $a$  was not significant (3.38 vs. 4.79,  $z =$   
365    -1.42,  $P = 0.15$ ). In the functional response curves, *L. idus* had higher consumption  
366    rates compared with *B. barbus*, with their 95 % confidence limits having minimal  
367    overlap (Fig. S4, S5).

368

369    For *S. cephalus* versus *L. idus*, differences in  $a$  were not significant for *G. pulex* (2.09  
370    vs. 3.23,  $z = -1.65$ ,  $P = 0.10$ ), but were significantly higher for *L. idus* on Chironomid  
371    larvae (1.37 vs. 4.79,  $z = -4.18$ ,  $P < 0.01$ ). Handling times were significantly lower in  
372    *S. cephalus* on both *G. pulex* (0.03 vs. 0.06,  $z = -3.84$ ,  $P < 0.01$ ) and Chironomid  
373    larvae (0.01 vs. 0.03,  $z = -4.16$ ,  $P < 0.01$ ). For both prey species, the functional  
374    response curves revealed high overlap in the 95 % confidence limits of their  
375    consumption rates (Fig. S4, S5).

376

377

378 *Co-habitation aquaria experiment*

379 Across the three species, there was considerable variation in their specific growth  
380 rates, varying between  $0.39 \pm 0.21$  (*B. barbus* in sympatry with *L. idus*) and  $1.07 \pm$   
381  $0.21$  (*S. cephalus* in sympatry with *B. barbus*). The LMEM testing differences across  
382 the treatments was significant ( $P < 0.01$ ). For *S. cephalus* and *L. idus*, differences in  
383 SGR between treatments were low, with substantial overlaps in their 95 % confidence  
384 limits (Fig. 1A). However, for *B. barbus*, there was a substantial reduction in SGR in  
385 sympatry with *L. idus* compared with their SGR in allopatry (Fig 1A).

386

387 *Cohabitation pond mesocosms*

388 The largest ranges of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  across the experiment were measured in the  
389 allopatric treatments and the sympatric treatment where all the species were together  
390 (Table 1; Fig. S6). When two fishes were sympatric, the SI ranges reduced, especially  
391 in the *B. barbus/ L. idus* treatment (Table 1; Fig. S6). These reduced SI ranges were  
392 concomitant with changes in the positions of the isotopic niches between allopatry  
393 and sympatry (Fig. 2). The predicted isotopic niche overlap between the species in  
394 allopatry was 31 to 39 % (Fig. 2A). When all the fish were in sympatry, these  
395 overlaps were reduced to 3 % for *L. idus* versus *B. barbus*, 11 % for *S. cephalus*  
396 versus *L. idus*, and 12 % for *S. cephalus* versus *L. idus* (Fig. 2B). This reduction in  
397 niche overlap when in sympatry was also apparent in treatments involving two  
398 sympatric fishes, where the extent of overlap varied from 5 % for *S. cephalus* versus  
399 *B. barbus* (Fig. 2D) to 15 % for *S. cephalus* versus *L. idus* (Fig. 2E). Concomitantly,  
400 isotopic niche sizes (as SEAc) reduced, with the posterior distributions of  $\text{SEA}_B$   
401 revealing these reductions were significant for both native species in sympatry with *L.*  
402 *idus* (Table 2).

403 The LMEM testing differences in SGR between treatments was significant ( $P < 0.01$ ).  
404 Compared to allopatry, *B. barbus* and *L. idus* growth rates were significantly reduced  
405 in their sympatric treatments involving paired species. This was, however, not  
406 apparent in *S. cephalus* (Fig. 1B), where differences in  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  between the  
407 species per treatment were also significant ( $P < 0.01$ ). Differences in metrics between  
408 allopatry and sympathy per species and treatment revealed that as niche size reduced,  
409  $\delta^{13}\text{C}$  was significantly enriched ( $R^2 = 0.55$ ,  $F_{1,7} = 8.39$ ,  $P = 0.02$ ; Fig. 3A). This was  
410 not apparent for  $\delta^{15}\text{N}$  ( $R^2 = 0.01$ ,  $F_{1,7} = 0.74$ ,  $P = 0.79$ ). The stable isotope mixing  
411 model predicted this shift to enriched  $\delta^{13}\text{C}$  was through a significant dietary shift  
412 away from Chironomid larvae and towards macrophyte and *G. pulex* (Chironomid:  $R^2$   
413 = 0.92,  $F_{1,7} = 65.54$ ,  $P < 0.01$ ; *G. pulex*:  $R^2 = 0.93$ ,  $F_{1,7} = 79.99$ ,  $P < 0.01$ ; macrophyte:  
414  $R^2 = 0.59$ ,  $F_{1,7} = 8.79$ ,  $P = 0.03$ ; Fig. 3B). The 5 - 95 % percentiles of the mixing  
415 model dietary predictions suggested, however, that these dietary shifts were only  
416 significant in sympatric treatments involving *B. barbus* and *L. idus*, but not *S.*  
417 *cephalus* (Table 3).

418 The multiple regression testing the influence of  $\text{MND}_{ij}$  and  $\text{MND}_{ii}$  on SGR was not  
419 significant ( $R^2 = 0.52$ ;  $F_{2,6} = 3.22$ ,  $P = 0.11$ ), but with  $\text{MND}_{ii}$  explaining more of the  
420 variability in SGR (standardised  $\beta = 0.69$ ,  $P = 0.09$ ) than  $\text{MND}_{ij}$  (standardised  $\beta =$   
421 0.04,  $P = 0.93$ ). Univariate linear regression revealed the relationship between  $\text{MND}_{ii}$   
422 and SGR was significant ( $R^2 = 0.47$ ;  $F_{1,7} = 6.32$ ,  $P = 0.04$ ; Fig. 4A), but was not  
423 significant for  $\text{MND}_{ij}$  ( $R^2 = 0.28$ ;  $F_{1,7} = 2.65$ ,  $P = 0.14$ ; Fig. 4B).

424

425

426

427

428 **Discussion**

429

430 In general, CFRs predict that ecologically damaging invaders have higher  
431 consumption rates than native species (e.g. Dick et al. 2013; Alexander et al. 2014).  
432 Here, they predicted that alien *L. idus* had higher attack rates and lower handling  
433 times than native *B. barbus*, resulting in significantly higher consumption rates in *L.*  
434 *idus*. In the cohabitation experiments in aquaria, the growth rates of *B. barbus* were  
435 significantly depressed in the presence of *L. idus* compared to allopatry. In contrast,  
436 the consumption rates of the taxonomically similar *S. cephalus* and *L. idus* were not  
437 significantly different and their growth rates did not differ significantly between  
438 treatments in the cohabitation experiment. In combination, these results suggest that  
439 competitive interactions between *L. idus* and *B. barbus* were asymmetric, as per the  
440 prediction. The superior competitor was *L. idus* due to their greater ability to access  
441 prey. This asymmetry in inter-specific competition was not, however, apparent  
442 between *L. idus* and *S. cephalus*, contrary to the prediction.

443

444 A criticism of CFRs for assessing the ecological impacts of alien species is that they  
445 do not adequately represent the ecological complexity inherent within more natural  
446 systems, where species can utilise multiple prey resources and are competing within a  
447 community of species of varying population abundances (e.g. Vonesh et al. 2017).  
448 They also cannot easily measure the competitive interactions within and between  
449 species directly (Guo et al. 2017). This is despite the potential importance of intra-  
450 and inter-specific competition in driving invasion-mediated changes in food web  
451 structure (David et al. 2017). Notwithstanding, the CFRs here did provide information  
452 on the comparative consumption rates of the fishes on the two major

453 macroinvertebrate prey species used in the SIA of the pond experiment.  
454 Correspondingly, their predictions provided a basis for evaluating the competitive  
455 interactions of the fish in pond mesocosms.

456

457 In the pond mesocosms, there were some significant shifts in the size and position of  
458 the isotopic niches of the fishes across the treatments. Comparison of the niche sizes  
459 of the species in allopatry versus their paired sympatric treatments revealed some  
460 important differences. For *L. idus* and *S. cephalus*, the aquaria experiments predicted  
461 their competitive interactions would be symmetric and in the pond experiment, their  
462 isotopic niche sizes were both reduced compared to allopatry (significantly so for *S.*  
463 *cephalus*). Whilst both species increased their dietary proportions of *G. pulex* and  
464 reduced their proportion of Chironomid larvae, there were sufficient dietary  
465 differences to result in their increased niche divergence in sympathy versus allopatry.  
466 This result was consistent with other studies that suggest trophic niche constriction  
467 and divergence occurs when an invader and competing native species exploit similar  
468 food resources (Tran et al. 2015; Jackson et al. 2016). The growth rates of both  
469 species in sympathy were, however, similar to allopatry. For *L. idus* and *B. barbus*,  
470 the aquaria experimental predictions of asymmetric competition favouring *L. idus*  
471 were not evident in the pond mesocosms. When paired, there were significant  
472 reductions in niche sizes in both species, with increased niche divergence, when  
473 compared to allopatry. These changes were accompanied by significantly reduced  
474 growth rates. These results were, however, also consistent with other studies  
475 suggesting increased inter-specific competition is an important determinant of  
476 invasion-mediated trophic impacts (e.g. Bøhn et al. 2008; Tran et al. 2015).

477

478 The results of the sympatric treatment involving all species in the pond mesocosm  
479 experiment revealed that compared with allopatry, there were no significant changes  
480 in isotopic niche sizes or growth rates of any species. Also, across the entire  
481 experiment, there was a significant relationship between reduced growth rates and  
482 reduced mean intra-specific isotopic dissimilarity, but not between growth and mean  
483 inter-specific trophic dissimilarity. In combination, these results suggest that inter-  
484 specific competition was not the only mechanism responsible for the measured  
485 changes in isotopic niche sizes and position across the experiment, with differences in  
486 the intensity of intra-specific competition also potentially important. Theory predicts  
487 that as intra-specific competition intensifies, individuals should become increasingly  
488 opportunistic and thus have greater niche variation (Svanbäck & Bolnick 2006; Rossi  
489 et al. 2015). The relatively large niches apparent in all allopatric treatments were  
490 consistent with this, where the intensity of intra-specific competitive interactions was  
491 assumed to be highest. In the sympatric treatments, however, the smallest isotopic  
492 niche sizes occurred when conspecifics were at  $n = 6$ , not at  $n = 4$ , contrary to theory  
493 (Svanbäck & Bolnick 2006). Correspondingly, the interaction of reduced intra- and  
494 inter-specific competition in the all-species treatment might have been positively  
495 interacting to facilitate the niche expansions (Nelson et al. 2017). Alternatively, in the  
496 all-species treatment, the species-pair direct effects that were apparent in the species-  
497 pair sympatric treatments might have been buffered by indirect effects (Calizza et al.  
498 2017; David et al. 2017). However, further work is needed to decouple these  
499 competition processes to more fully understand why the species-pair direct effects did  
500 not scale up and influence niche sizes in the all-species treatment.

501

502 The changes in the fish isotopic niche sizes and positions in the pond mesocosms  
503 highlight how aquatic invasive species can influence food web structure. In a meta-  
504 analysis on the impacts of aquatic invaders, Gallardo et al. (2016) revealed that  
505 competition and predation are the key processes driving ecological impacts in aquatic  
506 ecosystems, with indirect competitive effects from alien consumers often adversely  
507 affecting native species, leading to substantial modifications in food web structure  
508 (David et al. 2017). Invasions of alien fishes including *Carassius auratus*, *Cyprinus*  
509 *carpio*, *Pseudorasbora parva* and *Lepomis gibbosus* have all been shown to result in  
510 major re-organisations of the isotopic structure of the food web (e.g. Jackson &  
511 Britton 2014; Tran et al. 2015; Copp et al. 2017; Britton et al. 2018). Here, the alien  
512 *L. idus* also resulted in some food web re-structuring, with the effects involving both  
513 direct and indirect competitive effects depending on the number of fishes in the  
514 treatments.

515

516 Predicting the trophic consequences of invasive species remains an important  
517 theoretical and applied research area. Predictions from CFRs are that high-risk alien  
518 species tend to have significantly higher consumption rates than native analogues  
519 (Dick et al. 2013), with this consistent across fish (Alexander et al. 2014), amphipods  
520 (Laverty et al. 2015), snails (Xu et al. 2016) and decapods (Howard et al. 2018). Here,  
521 CFRs were used to predict the symmetry of inter-specific competition between  
522 species according to comparisons of their consumer-resource dynamics under  
523 standardised conditions. The results of the pond mesocosms between allopatry and  
524 species-pair treatments then revealed some consistency with the CFR results,  
525 especially *S. cephalus* versus *L. idus*. In the all-species treatment, however, there was  
526 greater complexity apparent in the results, and this complexity was beyond what the

527 CFRs could measure and predict. Thus, whilst CFRs have substantially increased  
528 understandings of the trophic impacts of invasive species (e.g. Alexander et al. 2014;  
529 Howard et al. 2018), their utility for predicting impacts is more limited in complex  
530 environments that involve a number of competing consumers. This is important, as  
531 competitive processes are important for structuring populations over a wide range of  
532 taxa, including snakes (e.g. Luiselli 2006), lizards (e.g. Mitchell 1979) and birds (e.g.  
533 Shochat et al. 2004). Moreover, studies across taxa suggest that the outcomes of  
534 competitive interactions are also influenced by a range of traits (e.g. body size and  
535 foraging behaviours) that then determine the diet of individuals, with food web  
536 structure being the sum of these individual diets (Petchey et al. 2008). The experiment  
537 here thus makes an important contribution to understanding how alterations in  
538 competition strength within and between species can impact the trophic niche sizes  
539 and positions of populations, and thus food web structure, whilst controlling for the  
540 effects of body size. The results also highlight how alien species integrate into food  
541 webs and alter the trophic relationships between native species.

542

543 A potential confound within the experiments was the use of hatchery-reared fishes,  
544 rather than fish collected from the wild. Hatchery-reared fishes were used due to the  
545 difficulty of obtaining sufficient numbers of wild fish to satisfy the experimental  
546 designs whilst controlling for size. There were also no wild *L. idus* British populations  
547 of sufficient abundance to provide the sample sizes. Literature suggests that there can  
548 be differences in the behaviours of hatchery-reared versus wild fish. For example, the  
549 movement behaviour and habitat use differed between wild and hatchery reared *S.*  
550 *cephalus* (Bolland et al. 2008), although the hatchery fish could cope with elevated  
551 flows and remained close to their stocking locations, as per wild fish (Bolland et al.

552 2009). Moreover, hatchery-reared fishes that are conditioned with natural stimuli and  
553 exposed to natural foods tend to have elevated post-release survival and more natural  
554 behaviours (e.g. Brown et al. 2003). The hatchery-reared fishes used in the  
555 experiments were all pond-reared, feeding on a mix of natural and supplementary  
556 foods. Consequently, as their husbandry used similar conditions to those in the  
557 enclosure experiment, and involved pond habitats and natural foods, the fish were  
558 considered a strong proxy for testing the interactions of wild fishes.

559

560 In summary, three experimental approaches tested the trophic consequences of an  
561 alien fish on two native fishes. Aspects of the shifts in isotopic niches and growth  
562 rates of fish in relatively complex environments were interpreted using the results of  
563 two relatively simple experiments completed in controlled conditions. However, the  
564 greater complexity of the pond systems when all the species were present resulted in  
565 more complex interactions and less predictable outcomes, and highlighted the direct  
566 and indirect interactions that enable alien species to integrate into native food webs.

567

#### 568 **Authors' contributions**

569

570 JRB conceived the ideas, designed the methodology and collected data; JRB, FAT,  
571 CGR, EN and VDS analysed data; JRB led manuscript writing. All authors  
572 contributed critically to drafts and approved submission.

573

#### 574 **Data accessibility**

575

576 Data will be provided in Dryad on acceptance.

577    **References**

578

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Table 1. Minimum, maximum and ranges of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$  per treatment in the pond mesocosm experiment. Note data are combined for all species.

| Treatment                               | $\delta^{13}\text{C} (\text{\textperthousand})$ |         |       | $\delta^{15}\text{N} (\text{\textperthousand})$ |         |       |
|---|---|---------|-------|---|---------|-------|
|   | Minimum   | Maximum | Range | Minimum   | Maximum | Range |
| Allopatric <i>B. barbus</i>             | -26.29  | -23.18  | 3.11  | 9.06  | 9.77    | 0.71  |
| Allopatric <i>S. cephalus</i>           | -26.13  | -23.40  | 2.73  | 8.96  | 9.65    | 0.69  |
| Allopatric <i>L. idus</i>               | -26.12  | -23.35  | 2.77  | 8.96  | 9.87    | 0.91  |
| Sympatric <i>B. barbus/ S. cephalus</i> | -25.37  | -22.84  | 2.53  | 9.12  | 10.23   | 1.11  |
| Sympatric <i>S. cephalus/ L. idus</i>   | -25.48  | -23.18  | 2.30  | 9.22  | 10.16   | 0.94  |
| Sympatric <i>B. barbus/ L. idus</i>     | -24.42  | -22.84  | 1.58  | 9.23  | 9.80    | 0.57  |
| All species in sympatry                 | -26.08  | -23.24  | 2.83  | 8.88  | 9.86    | 0.98  |

Table 2. Mean stable isotope values, isotopic niche size (as standard ellipse areas, SEAc (c = correction for small sample size) and SEA<sub>B</sub> (Bayesian estimate of SEA) of the macroinvertebrate and macrophytes food resources, and for each fish species by treatment in pond mesocosms. For SEA<sub>B</sub>, the mean and standard error at a credible interval of 95% (in parentheses) are presented. \*Difference in niche size as SEA<sub>B</sub> between the treatment and allopatry is significantly different ( $P < 0.05$ ).

| Spp.                  | Treatment          | N  | Mean $\delta^{13}\text{C}$ (‰) | Mean $\delta^{15}\text{N}$ (‰) | SEA <sub>c</sub> (‰ <sup>2</sup> ) | SEA <sub>B</sub> ‰ <sup>2</sup> (CI 95%) |
|-----------------------|--------------------|----|--------------------------------|--------------------------------|------------------------------------|--|
| <i>Gammarus pulex</i> |                    | 3  | -26.22 ± 0.68                  | 7.38 ± 0.40                    |                                    |  |
| Chironomid larvae     |                    | 3  | -31.37 ± 1.47                  | 5.35 ± 1.47                    |                                    |  |
| Macrophyte            |                    | 3  | -27.82 ± 0.69                  | 1.54 ± 0.56                    |                                    |  |
| <i>L. idus</i>        | Allopatry          | 15 | -24.94 ± 0.21                  | 9.41 ± 0.06                    | 0.61                               | 0.51 (0.31-0.93)                         |
|                       | <i>B. barbus</i>   | 12 | -23.51 ± 0.12                  | 9.55 ± 0.04                    | 0.19                               | 0.19 (0.10-0.34)*                        |
|                       | <i>S. cephalus</i> | 12 | -23.69 ± 0.14                  | 9.49 ± 0.09                    | 0.27                               | 0.32 (0.15-0.51)                         |
|                       | All species        | 12 | -23.93 ± 0.12                  | 9.51 ± 0.07                    | 0.33                               | 0.33 (0.14-0.53)                         |
| <i>B. barbus</i>      | Allopatry          | 15 | -24.85 ± 0.23                  | 9.34 ± 0.05                    | 0.51                               | 0.64 (0.26-0.81)                         |
|                       | <i>L. idus</i>     | 12 | -23.87 ± 0.11                  | 9.49 ± 0.05                    | 0.21                               | 0.22 (0.08-0.27)*                        |
|                       | <i>S. cephalus</i> | 12 | -23.70 ± 0.15                  | 9.60 ± 0.05                    | 0.24                               | 0.26 (0.12-0.41)*                        |
|                       | All species        | 12 | -24.15 ± 0.19                  | 9.18 ± 0.06                    | 0.49                               | 0.35 (0.22-0.71)                         |
| <i>S. cephalus</i>    | Allopatry          | 15 | -24.68 ± 0.20                  | 9.46 ± 0.05                    | 0.52                               | 0.50 (0.27-0.80)                         |
|                       | <i>L. idus</i>     | 13 | -24.29 ± 0.14                  | 9.66 ± 0.04                    | 0.26                               | 0.26 (0.13-0.42)*                        |
|                       | <i>B. barbus</i>   | 12 | -24.47 ± 0.20                  | 9.94 ± 0.10                    | 0.70                               | 0.73 (0.33-1.16)                         |
|                       | All species        | 12 | -24.69 ± 0.26                  | 9.46 ± 0.06                    | 0.50                               | 0.65 (0.25-0.85)                         |

Table 3. Predicted dietary proportions of the three putative food resources for the three fishes by treatment in the pond mesocosms.

|                    |                    | Mean predicted dietary proportion (5-95 <sup>th</sup> percentile of distribution range) |                       |                  |
|--------------------|--------------------|---|-----------------------|------------------|
| Spp.               | Treatment          | Chironomidae  | <i>Gammarus pulex</i> | Macrophyte       |
| <i>B. barbus</i>   | Allopatry          | 0.33 (0.22-0.44)  | 0.25 (0.18-0.33)      | 0.42 (0.35-0.48) |
|                    | All species        | 0.18 (0.09-0.27)  | 0.32 (0.25-0.39)      | 0.50 (0.44-0.56) |
|                    | <i>S. cephalus</i> | 0.10 (0.03-0.19)  | 0.44 (0.38-0.50)      | 0.46 (0.40-0.51) |
|                    | <i>L. idus</i>     | 0.12 (0.05-0.21)  | 0.41 (0.35-0.47)      | 0.47 (0.41-0.52) |
| <i>S. cephalus</i> | Allopatry          | 0.31 (0.21-0.42)  | 0.28 (0.21-0.36)      | 0.41 (0.34-0.47) |
|                    | All species        | 0.32 (0.21-0.46)  | 0.28 (0.19-0.36)      | 0.40 (0.33-0.47) |
|                    | <i>L. idus</i>     | 0.22 (0.13-0.32)  | 0.37 (0.31-0.44)      | 0.40 (0.34-0.46) |
|                    | <i>B. barbus</i>   | 0.29 (0.18-0.42)  | 0.38 (0.29-0.46)      | 0.33 (0.26-0.39) |
| <i>L. idus</i>     | Allopatry          | 0.36 (0.24-0.49)  | 0.24 (0.16-0.33)      | 0.40 (0.32-0.47) |
|                    | All species        | 0.15 (0.07-0.23)  | 0.40 (0.34-0.46)      | 0.46 (0.40-0.51) |
|                    | <i>S. cephalus</i> | 0.09 (0.03-0.18)  | 0.43 (0.37-0.48)      | 0.48 (0.42-0.53) |
|                    | <i>B. barbus</i>   | 0.07 (0.01-0.14)  | 0.46 (0.40-0.51)      | 0.47 (0.43-0.53) |

## Figure captions

Figure 1. Mean specific growth rates of cohabitation experiments completed in (A) tank aquaria, and (B) pond enclosures, where C = control (i.e. each species in allopatry), Ch = sympatry with chub *Squalius cephalus*, Id = sympatry with ide *Leuciscus idus*, Ba = sympatry with barbel *Barbus barbus*, and All = all species in sympathy. Clear circles: barbel, black circles: chub, grey circles: ide. Note differences in axes values between (A) and (B).

Figure 2. Stable isotope bi-plots comparing the standard ellipse area ( $SEA_c$ ) the fishes in allopatry and sympathy, where A)  $SEA_c$  of each species in allopatry, B) the species all in sympathy, C) sympatric *L. idus* and *B. barbus*, D) sympatric *B. barbus* and *S. cephalus*, and E) sympatric *L. idus* and *S. cephalus*. Filled circles/ black dashed line: *L. idus*, filled triangles and black solid lines: *B. barbus*; clear squares, and grey solid lines: *S. cephalus*. The mean SI data for the fish putative food resources were Chironomid larvae:  $\delta^{13}\text{C}$ :  $-31.37 \pm 1.47\text{‰}$ ,  $\delta^{15}\text{N}$ :  $5.35 \pm 1.47\text{‰}$ ; *G. pulex*:  $\delta^{13}\text{C}$ :  $-26.22 \pm 0.68\text{‰}$ ,  $\delta^{15}\text{N}$ :  $7.38 \pm 0.40\text{‰}$ ; macrophyte:  $\delta^{13}\text{C}$ :  $-27.82 \pm 0.69\text{‰}$ ,  $\delta^{15}\text{N}$ :  $1.54 \pm 0.56\text{‰}$ .

Figure 3. (A) Relationships of differences in isotopic niche size (as  $SEA_c$ ) between allopatric and sympatric treatments versus their differences in  $\delta^{13}\text{C}$ ; and (B) Relationships of differences in mean  $\delta^{13}\text{C}$  between allopatric and sympatric treatments per species versus differences in their predicted dietary proportions per food resource (Chironomid larvae: clear circles, dashed line; *Gammarus pulex*: filled circles, small dashed line; macrophytes: grey circles, solid line). All straight lines

represent the significant linear relationship between the variables (linear regression: P < 0.03).

Figure 4. Relationships of the mean intra-specific (A) and intra-specific (B) trophic dissimilarity versus specific growth rate for fishes in sympatric treatments in the pond mesocosm experiment. The solid line represents the significant relationship between the variables according to linear regression.

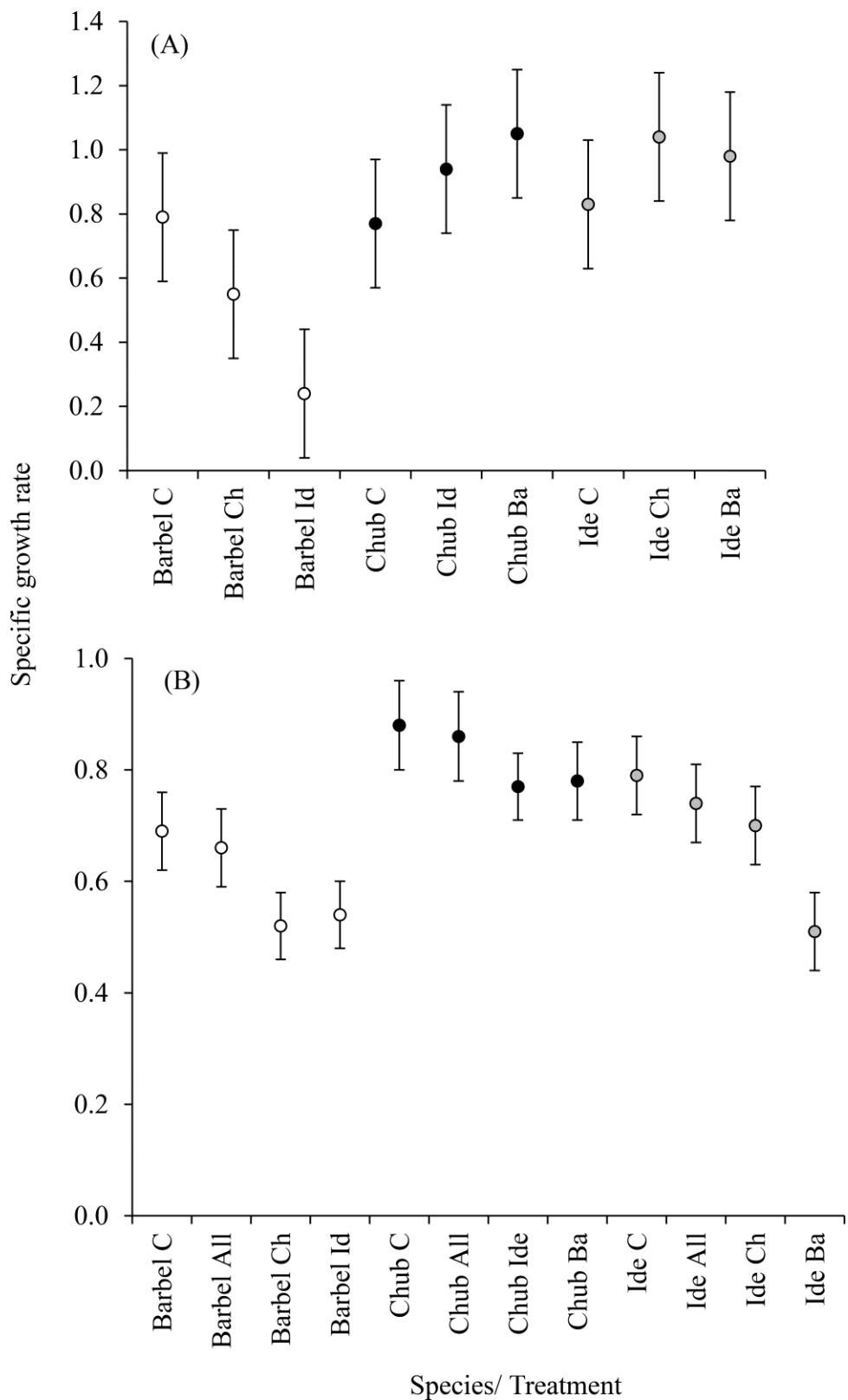


Figure 1.

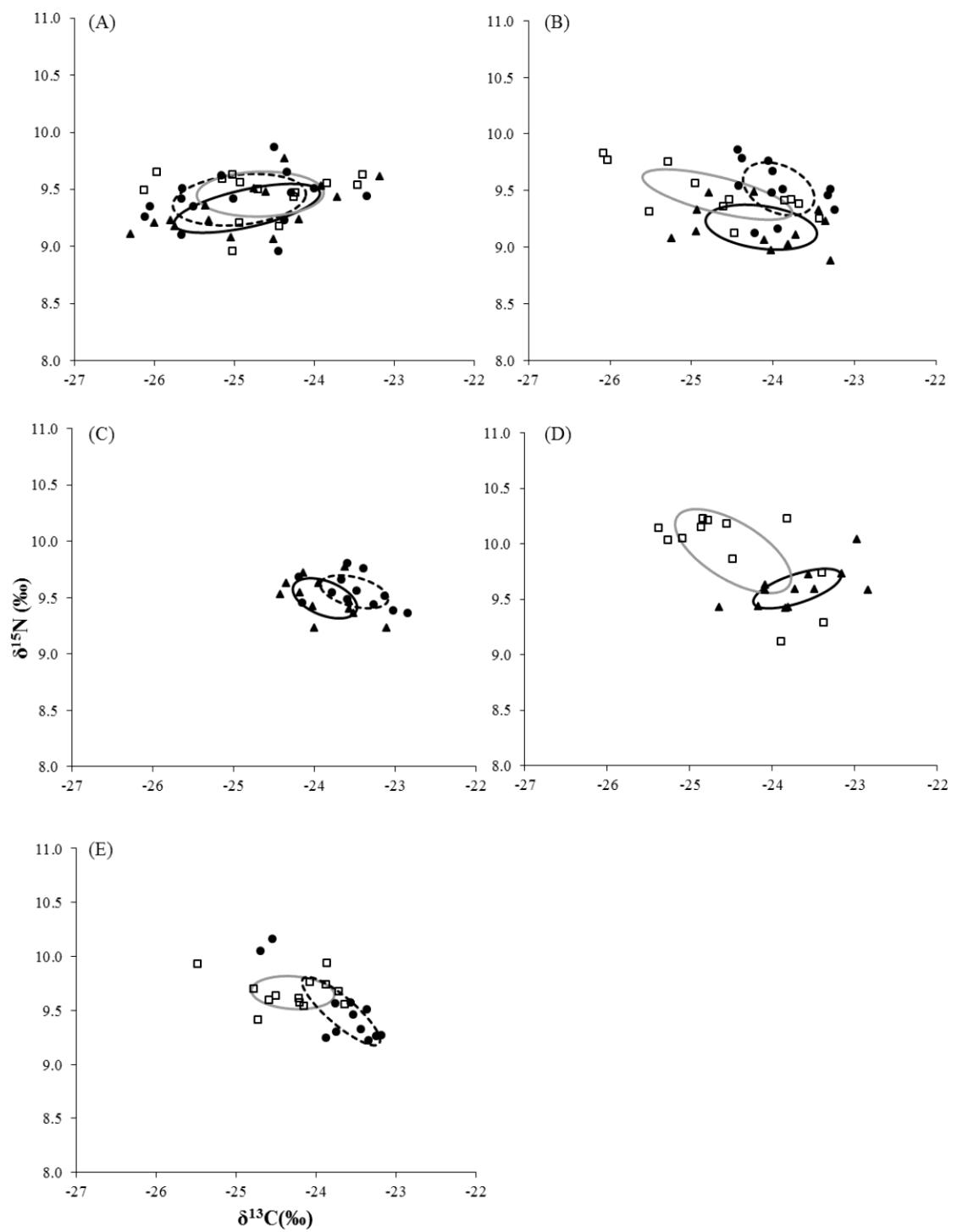


Figure 2.

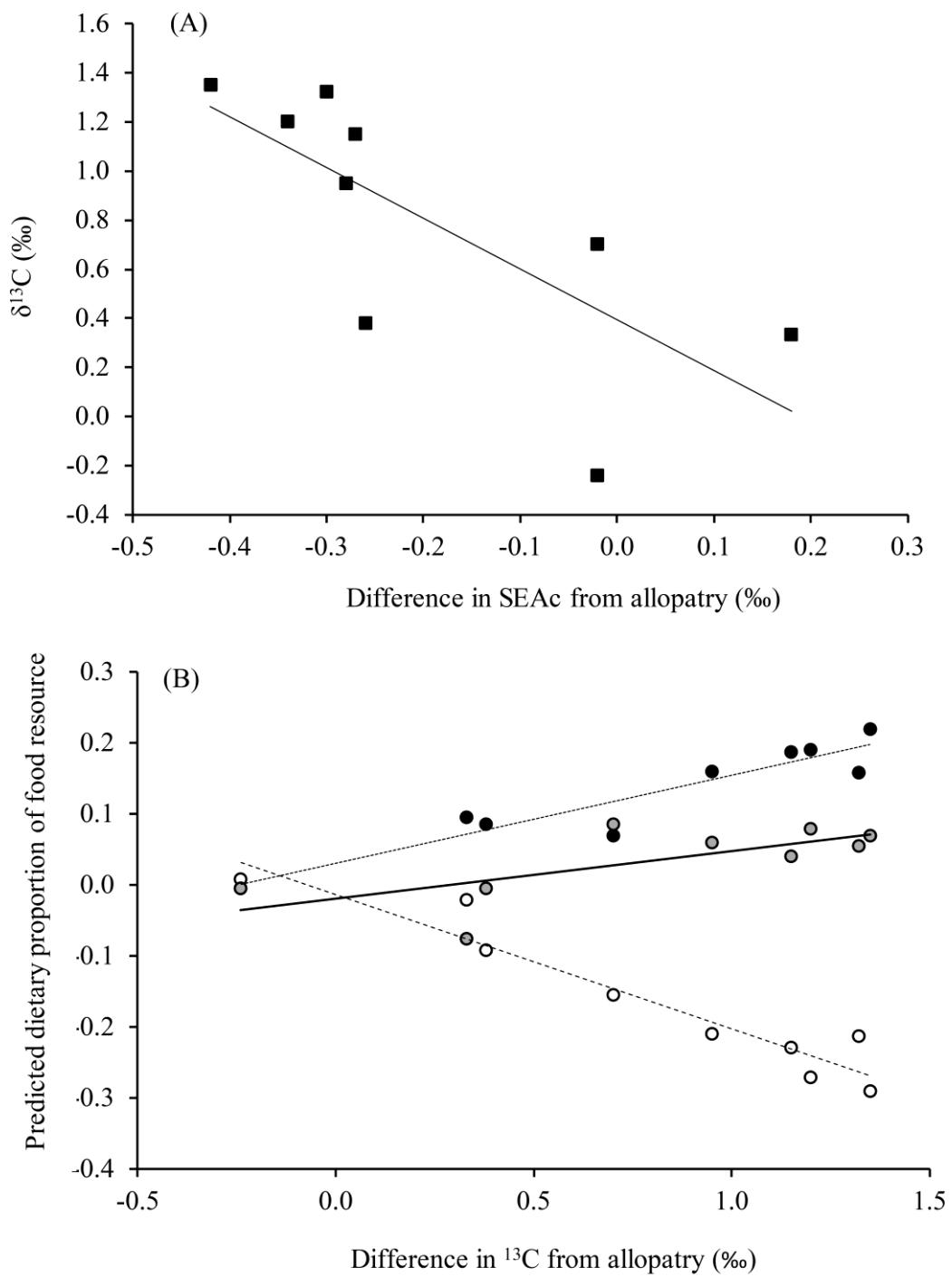
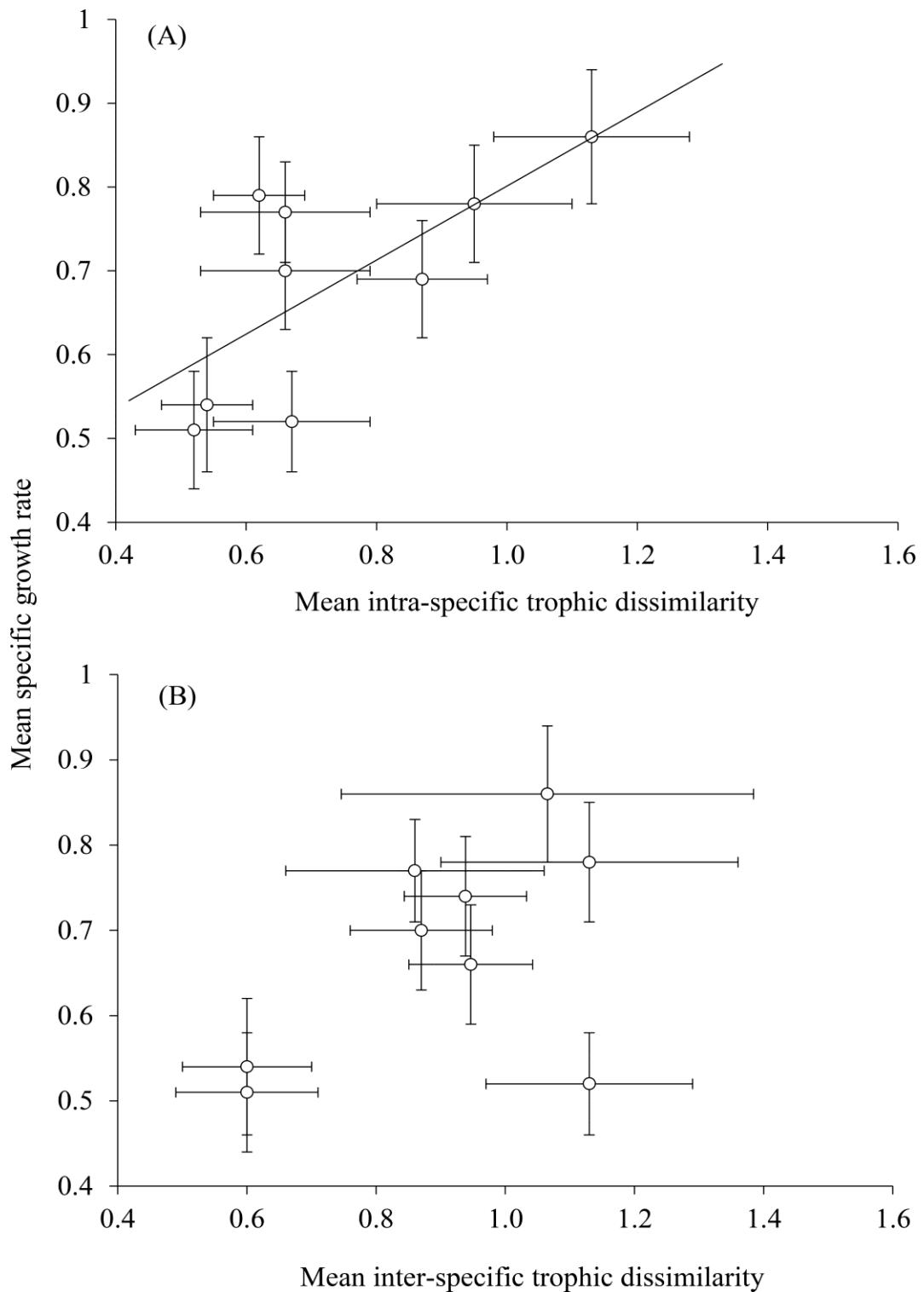


Figure 3.



Figure

4.

## **Supplementary material**

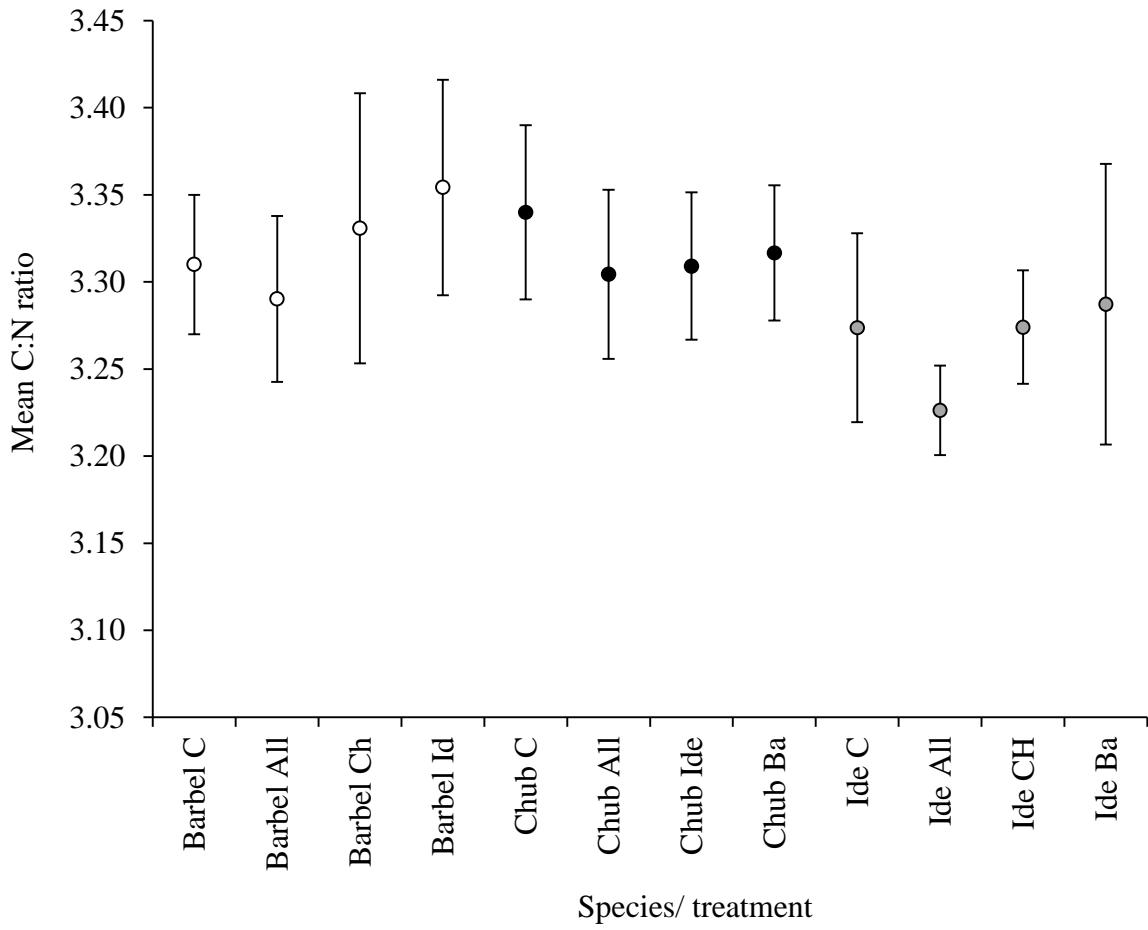


Figure S1. Mean C:N per species and treatment in the pond enclosures, where C = control, Ch = sympatry with chub *Squalius cephalus*, Id = sympatry with ide *Leuciscus idus*, Ba = sympatry with barbel *Barbus barbus*, and All = all species in sympatry. Clear circles: barbel, black circles: chub, grey circles: ide. Note differences in axes values between (A) and (B). Differences in C:N ratios between the species per treatment were not significant ( $F_{1,152} = 1.74$ ,  $P = 0.10$ ).

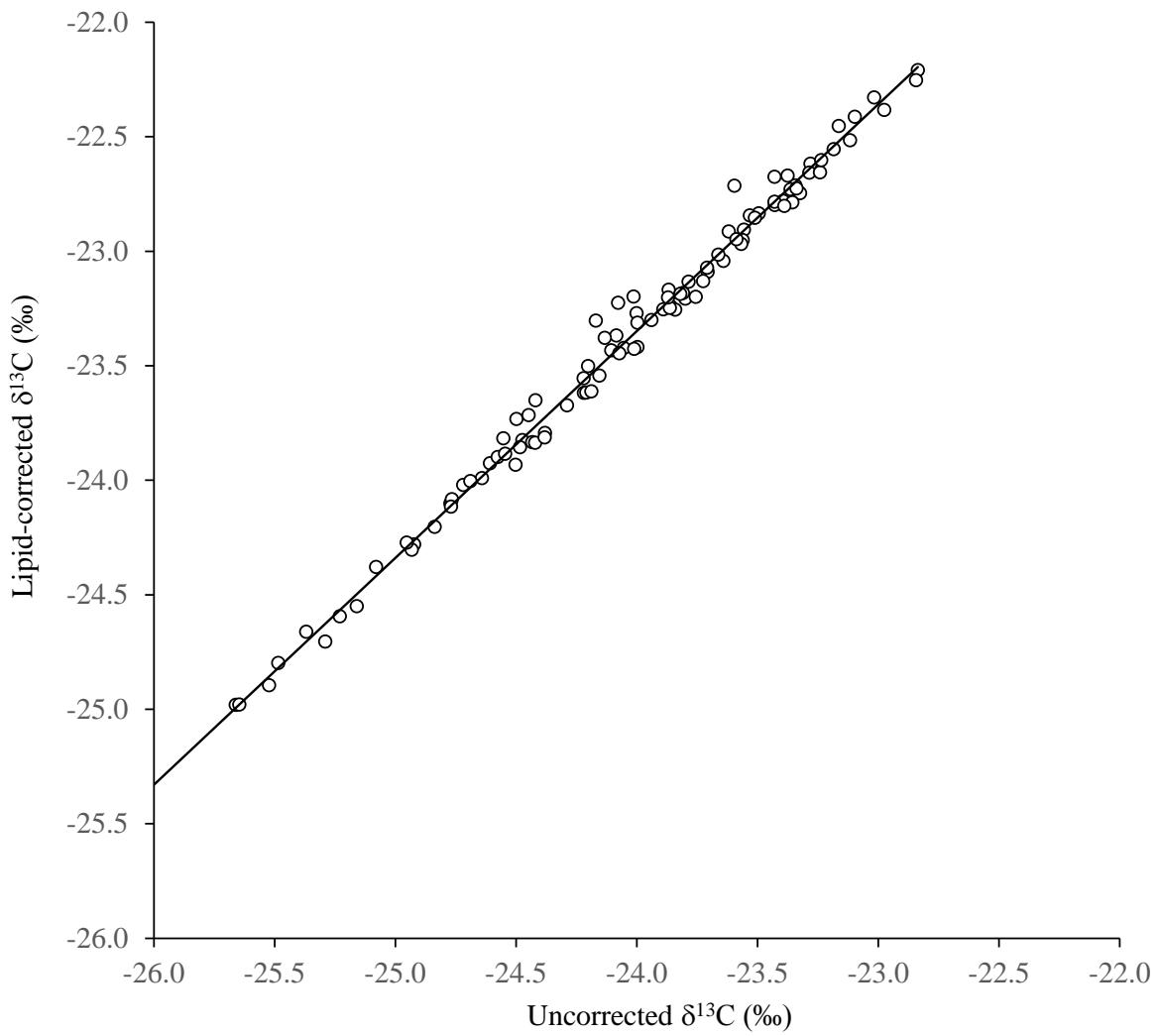


Figure S2. Relationship of uncorrected versus lipid corrected  $\delta^{13}\text{C}$  for all fish samples (Kiljunen et al. 2006), where the solid line is the significant relationship according to linear regression ( $R^2 > 0.99$ ,  $F_{1,152} = 15066.9$ ,  $P < 0.001$ ).

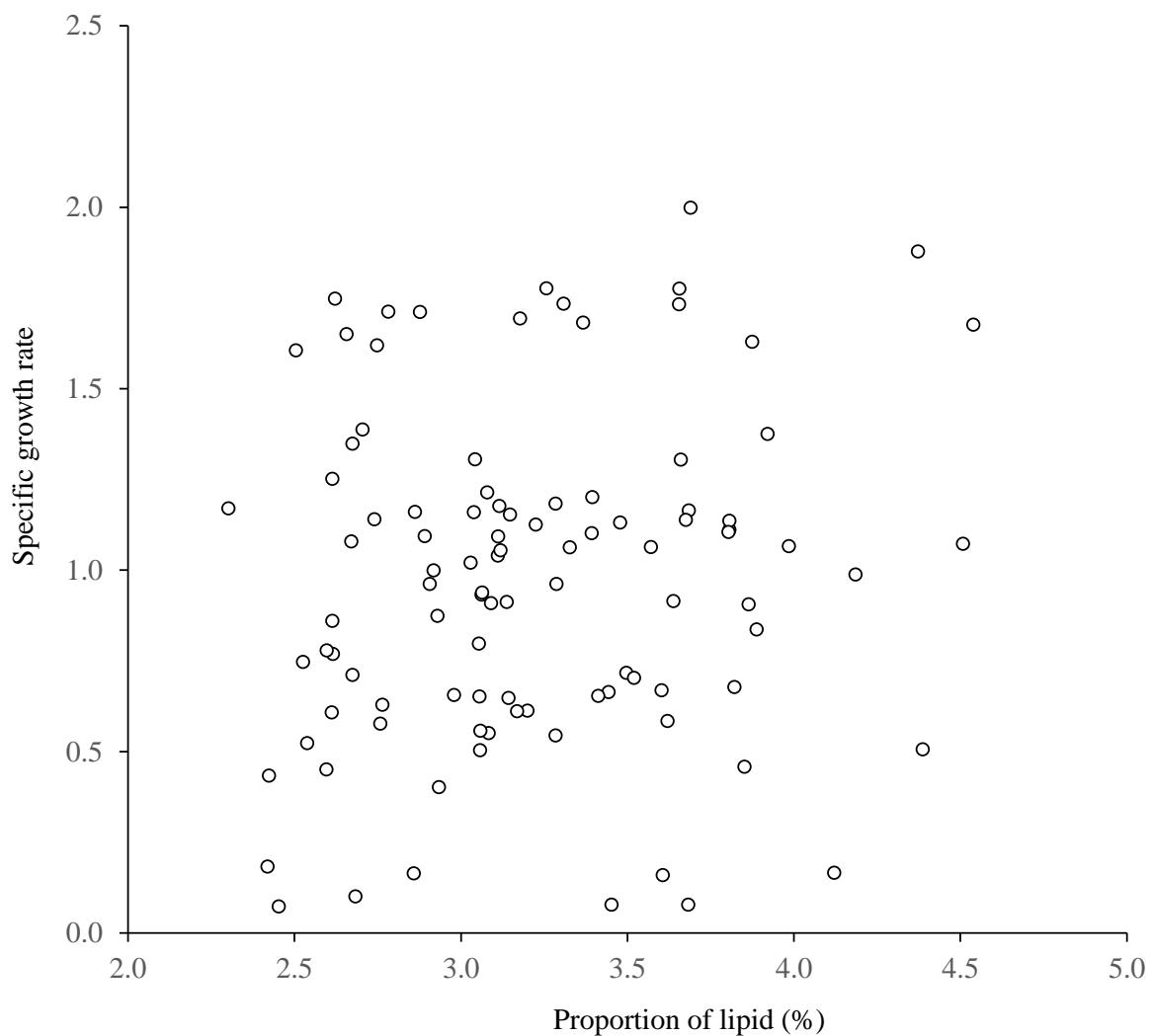


Figure S3. Relationship of proportion of lipid in the analysed dorsal muscle samples of each individual fish, as calculated  $\delta^{13}\text{C}$  and C:N ratios (Post et al. 2007), versus their specific growth rates. The relationship was not significant according to linear regression ( $R^2 = 0.02$ ,  $F_{1,152} = 2.18$ ,  $P = 0.14$ ).

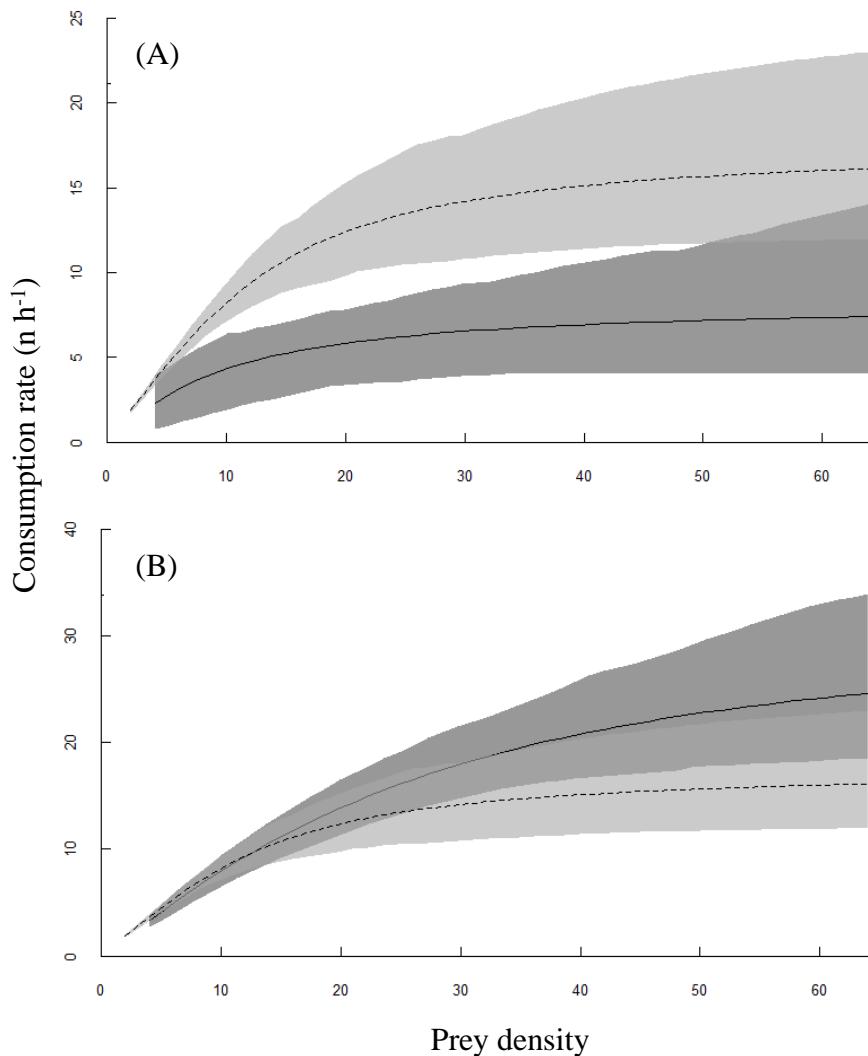


Figure S4. Comparative functional response curves for *Gammarus pulex* as prey, comparing *Leuciscus idus* (dashed line) versus (A) *Barbus barbus* (solid line) and (B) *Squalius cephalus* (solid line). Shaded areas around the curves represent 95 % confidence intervals generated by boot-strapping. Note differences in values on the Y axis.

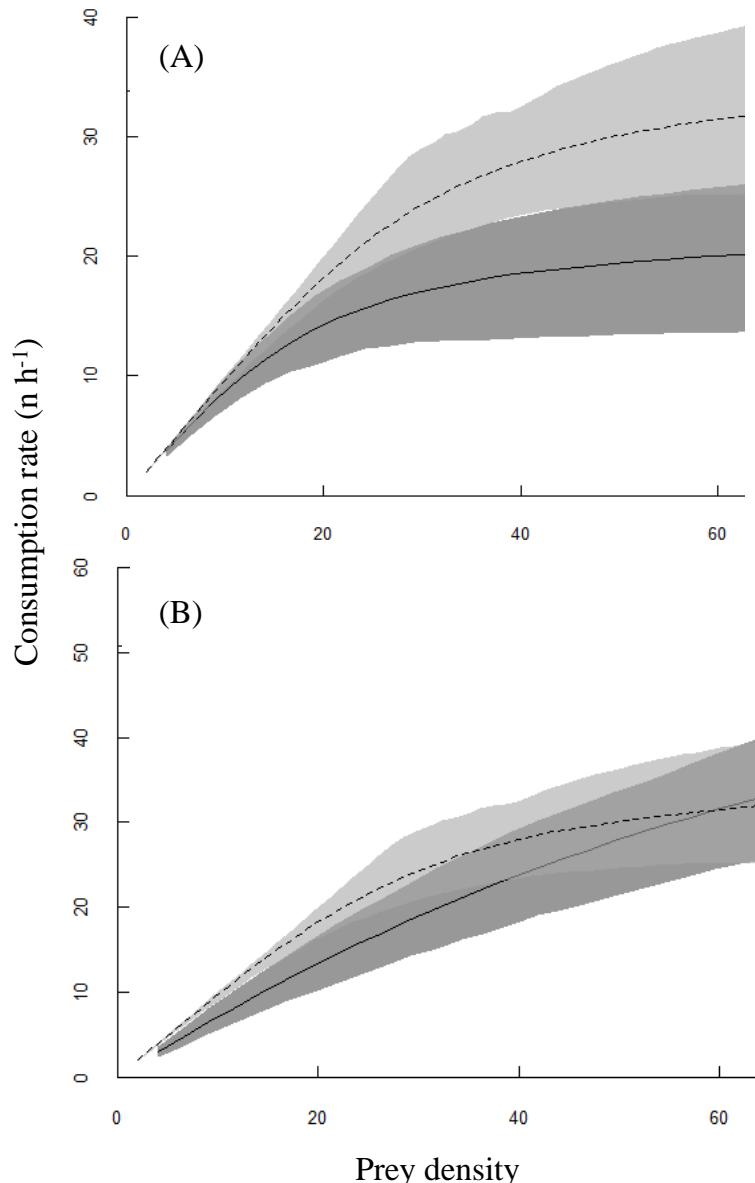


Figure S5. Comparative functional response curves for Chironomid larvae as prey, comparing *Leuciscus idus* (dashed line) versus *Barbus barbus* (solid line) (A) and (B) *Squalius cephalus* (solid line). Shaded areas around the curves represent 95 % confidence intervals generated by boot-strapping. Note differences in values on the Y axis.

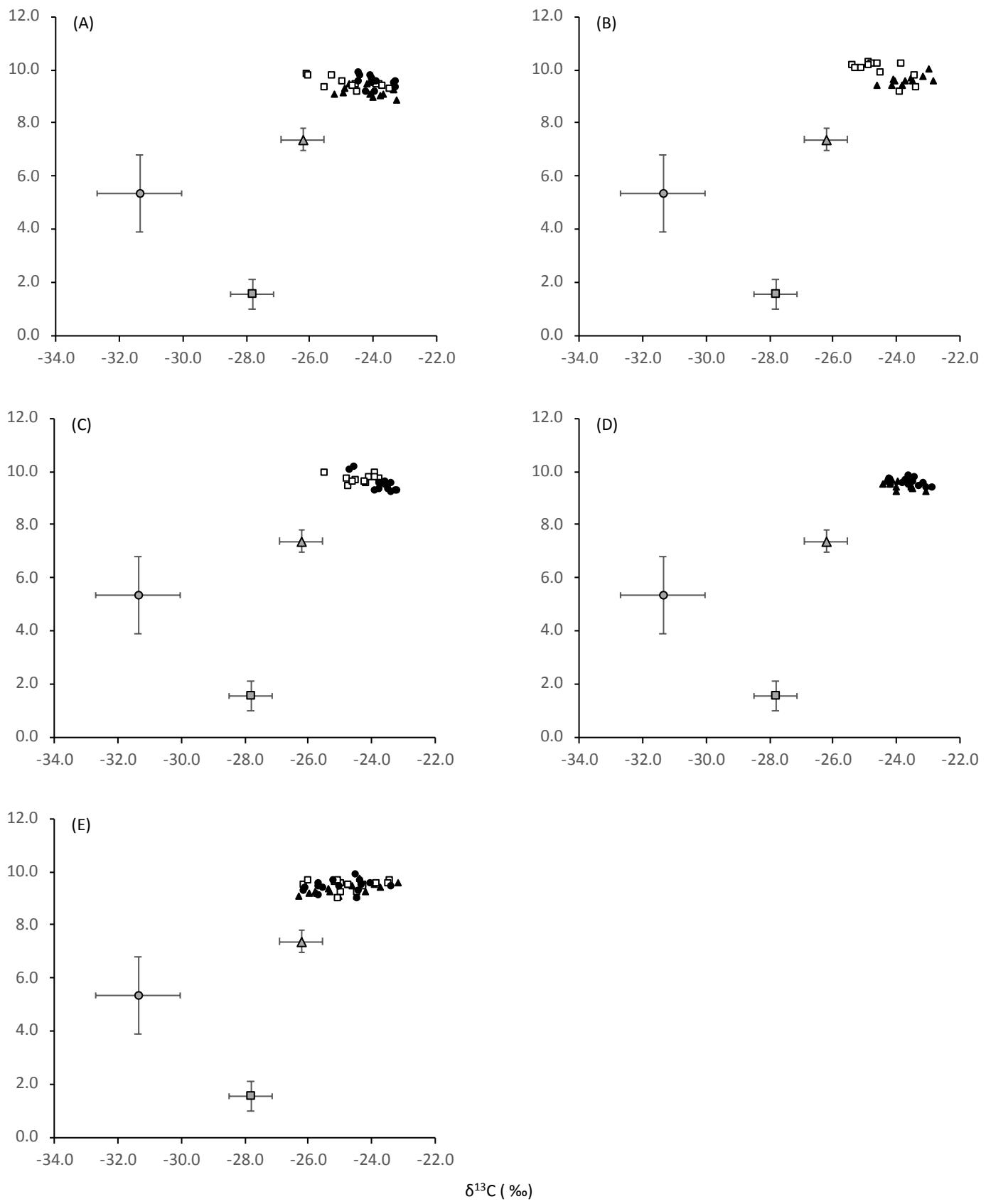


Figure S6. Stable isotope biplots for (A) All species sympatric treatment; (B) *Barbus barbus*/ *Squalius cephalus* species pair treatment; (C) *S. cephalus*/ *Leuciscus idus* species pair treatment; (D) *B. barbus*/ *L. idus* species pair treatment; and (E) All species in allopatry. For fish, filled circles: *L. idus*; filled triangles: *B. barbus*; clear squares: *S. cephalus*. For putative prey used in the stable isotope mixing models to predict fish diet, grey circle = Chironomid larvae; grey triangle = *Gammarus pulex*; grey square = macrophyte. Error bars represent 95 % confidence limits.

## **References**

- Kiljunen, M., Grey, J., Sinisalo, T., Harrod, C., Immonen, H. & Jones, R.I. 2006. A revised model for lipid-normalizing  $\delta^{13}\text{C}$  values from aquatic organisms, with implications for isotope mixing models. *Journal of Applied Ecology* 43, 1213-1222.
- Post, D.M., Layman, C.A., Arrington, D.A., Takimoto, G., Quattrochi, J. & Montana, C.G. 2007. Getting to the fat of the matter: models, methods and assumptions for dealing with lipids in stable isotope analyses. *Oecologia*, 152, 179-189.