

**African savanna elephant (*Loxodonta africana*) impacts on  
vegetation in a fenced area and the broader implications for  
elephant conservation**



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## **Abstract**

The first chapter assessed whether a monitored population of African savanna elephants (*Loxodonta africana*) impacted tree species in the small, fenced Karongwe Private Game Reserve (KPGR), South Africa and to determine levels of tree recovery. Trees in high-use areas were significantly less likely to show signs of debarking and push over. Tall trees were significantly more likely to be impacted by elephants, being associated with heightened risks of debarking and branches being broken but lower risks of being pushed over. Trees close to the fence line were not more impacted than trees near the centre of the reserve. The level of use, distance to the fence and tree height were not significant predictors of tree recovery indicators. Future mitigation efforts should focus on trees with high levels of impact and low levels of recovery. The second chapter considered how elephant impact influenced secondary damages to trees and how this effected tree recovery levels. Secondary damage was identified as insects, where wood borers and termites were considered. Irrespective of tree height, termites were found to be more likely to colonise damaged trees without signs of recovery and wood borers were more likely to colonise damaged trees showing signs of recovery. Therefore, carefully considering management approaches for elephant-induced termite and wood borer impact on trees should be applied in this fenced reserve. Following this, the third chapter considered an alternative method for identifying tree height in the KPGR, using aerial photographs with 3-D photogrammetry. Initial findings here indicate that this method was successful at identifying tree height within this fenced environment. This method should be repeated in such environments in conjunction with other mapping agencies. Lastly, the fourth chapter investigated school children's perceptions of elephants and elephant conservation at different geographical locations. Different levels of exposure to elephants were significant in influencing school children's perceptions. More research is required to enhance environmental education practises to support elephant conservation across different locations.

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## **Author's declaration**

I (KT) confirm that the research presented within this thesis is my own.

The following research papers were submitted or prepared for publication in collaboration with Genoveva Esteban (GE), Andrew Ford (AF), Nathalie Pettorelli (NP) as my core supervisory team. External collaborators Kayla Zoon (KZ) and Angelo Poupard (AP) contributed to studies within this thesis. For Chapter 2 and 3, KZ aided with data collection. AP and AF provided some analyses in Chapter 2 and 4, respectively. KT led the writing of the manuscript:

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Thompson, K., Esteban, G., Pettorelli, N., Ford, A., The potential of photogrammetric point clouds derived from conventional aerial survey for estimating tree heights. *Photogrammetric records*. (prepared for publication; Chapter 4)

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## **1 Introduction**

### **1.1 Organisation of the Thesis**

The thesis is presented in an integrated format, whereby material is incorporated in a style suitable for publication in a peer review journal. The first chapter outlines the main themes of the thesis: African savanna elephant (*Loxodonta africana*) impacts on vegetation and the broader implications for elephant conservation. The study areas, aims and objectives of the research project are then introduced. Data chapters 2, 3 and 4 are each presented as original and complete pieces of research, i.e., manuscripts. The final data chapter (chapter 5) is presented as a chapter that will be later condensed into a manuscript for publication. Chapter 6 discusses the implications of this research, future research directions and provides concluding remarks. The complete list of references is provided after the final chapter, to avoid replication and improve readability. Appendices containing data collected are included at the end of the thesis (Appendix 4).

### **1.2 African elephant conservation status**

African elephants (*Loxodonta africana* and *Loxodonta cyclotis*) are found across the African continent at different population densities (Chase et al. 2016; Blanc et al., 2007, 2008; Thouless et al., 2016). After decades of deliberating whether African elephants should in fact be classified as two species, the International Union for Conservation of Nature (IUCN) has now listed African savanna elephants (*L. africana*) as ‘Endangered’ and the African forest elephant (*L. cyclotis*) as ‘Critically Endangered’ (Gobush et al., 2021). This is significant as before the latest status announced this year (2021), the IUCN listed all African elephants as ‘Vulnerable’. The decision was made by the IUCN to treat savanna elephants and forest elephants as separate

species, based on genetic evidence on elephant populations (Roca et al., 2015; Mondol et al., 2015). Forest elephants are found in tropical forests in Central Africa and West Africa, and rarely overlap with the range of savanna elephants (largely East and Southern Africa) (Roca et al., 2007; Ishida et al., 2011; Mondol et al., 2015; Palkopoulou et al., 2018; Kim and Wasser, 2019). Additionally, forest elephants are smaller, gestate longer, have oval ears and smaller tusks. Savanna elephants live in large family units (known as breeding herds) have larger ears and different shaped skulls to forest elephants (Figure 1.1).



**Figure 1.1** African savanna elephant (*L. africana*) (top image) and African forest elephant (*L. cyclotis*) (bottom image). Top image taken by the author (2018) and bottom image taken by Frank af Petersons, Save the elephants (Gobush et al., 2021).

This scientific development is extremely important mainly for the conservation implications it will have. The new IUCN assessment (2021) has found that in the last 31 years, the population of forest elephants has decreased by more than 86%, while the population of savanna elephants has decreased by at least 60% in the last 50 years (Gobush et al., 2021). The causes of these declines have been attributed to ivory poaching, habitat loss, and land fragmentation (Lemieux and Clarke, 2009; Thouless et al., 2016; Wittemyer et al., 2014). These alarming new statistics only increase the need for continued research on these sentient animals (Reinwald et al., 2021). It is important to acknowledge here that decreasing populations of savanna elephants is not spatially uniform across the continent, and great variability in elephant populations is evident across Africa. Therefore, conservation of both savanna and forest elephants needs to be carefully tailored to location. Considerable caution and local knowledge are required when applying this information into policy and management, and further actions are needed to conserve both species and reduce such unsustainable rapid declines.

### ***1.2.1 Milestones in elephant conservation***

Acknowledging the decline in elephant populations has historically given rise to many conservation projects, driven by international policies, as a result of poaching and conflict (Thouless et al., 2016; Chase et al., 2016). African elephants are part of the ‘Big Five’: the most dangerous and toughest animals to hunt in Africa (Capstick, 1983; Du Toit, 2001). Management of elephant culling was undertaken up until the early ‘90s, but this method has largely ceased today (Carruthers et al., 2008). However, in some countries game hunting of African savanna elephants is still legally permitted, and it is important source of income for those countries (Naidoo et al., 2016).

In 1989, when both African elephants were considered a single species, the African savanna elephant was listed in the Convention on International Trade in Endangered Species of Wild

Flora and Fauna (CITES). CITES-approved sales of national ivory stockpiles took place in 2002 and 2008, for Botswana, Zimbabwe and Namibia and later South Africa to two buyers: Japan and China. At this time, the demand for ivory in Japan decreased whilst demand in China grew, causing ivory prices to increase dramatically in China and Africa (Wittemyer et al., 2011, 2014) and debates around the benefits and consequences of these sales are highly divided (Stiles, 2004; Bennett, 2015; Biggs et al., 2017). This opening of the legal ivory market in China raised concerns by conservationists that this would lead to a rise in poaching. Added to this, illegally trafficking of ivory occurs today, and there has been a significant increase since 2006 (Underwood et al., 2013; Milliken, 2016).

Empirical research conducted by Monitoring the Illegal Killing of Elephants (MIKE) and the Elephant Trade Information System (ETIS) of ivory seizures between 2009-2013, affirmed that poaching levels of elephants and ivory trafficking occurred at alarming rates, and elephant populations across the continent went into net decline (CITES, 2018, 2019). At this time, growth of the consumer class in China increased the demand for ivory, driving poachers to increase harvesting ivory for money (Vigne and Martin, 2017; Meijer et al., 2018) and CITES recognised that elephant poaching has, again, reached an unsustainable level.

Following this, the Great Elephant Census was conducted in 2014, spanning 18 countries and 295,000 miles (Chase et al., 2016). Results released in 2016 revealed that there had been a 30% drop of savanna elephants in seven years. At a national level, National Ivory Action Plans (NIAPs) have been developed to combat illegal ivory trade, where the African savanna elephant is protected at different levels. As half of the species range is outside of protected areas (Taylor, 2009) and the level of protection is an important predictor of the species presence and density (de Boer et al., 2013), providing protection is critical but fundamentally challenging because of these factors. However, conservation measures are vital, as Chase et

al., (2016) demonstrated in areas where protection measures fail, elephant populations reduced by 70% within a 10-year period.

Savanna elephant reintroductions within fenced reserves across South Africa has proved successful in maintaining and increasing elephant population numbers over the last 25 years (Blanc and Barnes, 2007; Chase et al., 2016). Given the recent change in status of savanna elephants, this population increase is crucial for conserving elephants that are generally in decline. However, this population increase has raised concerns with regards to the suitability of these constrained areas for increasing elephant numbers. This thesis will consider a small, fenced reserve with high elephant density (Karongwe Private Game Reserve (KPGR), South Africa), to gain insights into the impacts of elephants on vegetation in response to their habitat use.

### **1.3 Importance of elephants**

Elephants are among the most intelligent animals, with complex consciousness and strong emotions (Bates, Pool and Byrne, 2008). Their societal structures of large networks with hierarchical organisation may be one of the most elaborate of any vertebrate (Irie and Hasegawa, 2009). They have a flexible, fission-fusion society which means that they can differentiate between hundreds of individuals (Byrne and Bates, 2011). Elephant herds are led by matriarchs, which communicate with each other using low-frequency ultrasound calls (Payne et al., 1986) over distances of more than 1 km (Garstang, 2004). There is also increased evidence demonstrating sophisticated cognitive abilities in elephants (Irie and Hasegawa, 2009). Studies have shown that they can recognise their own reflection in a mirror (Plotnik et al., 2006), and are able to learn various commands by jungle dwellers (Rensch, 1957). Furthermore, they are unique in terms of having a prehensile trunk, which is used (amongst

other purposes) for feeding, modifying objects, greeting other elephants and in conflict with other elephants (Hart and Hart, 1994).

African and Asian elephants are important ecologically as they are ecosystem engineers, promoting diversity of both habitats and species of flora and fauna (Bond, 1993; Jones et al., 1994). They play a key, ecological role in maintaining the linkages in the food web and their extinction would cause changes in both forest and savanna ecosystems (Western, 1989). Elephants are also important agents of seed dispersal, increasing habitat mosaics and diversifying mammalian communities (Bunney et al., 2017; Cochrane, 2003). Studies have shown that with few elephants present, vegetation can become homogenous, and woodland can become dense and encroaching which indirectly shapes the remaining wildlife community (Fritz et al., 2002). Elephants play an essential role in creating forest gaps, which helps diversify forests enabling vegetation to grow in otherwise canopy shadowed areas, in turn supporting other vertebrate feeding (Western, 1989; Valeix et al., 2011). Their distinct feeding and movement patterns shape the faunal and floral diversity of savanna and forest ecosystems (Ben-Shahar, 1993; de Beer et al., 2006). Elephant presence within savanna ecosystems is therefore essential for many species, and their removal will lead to cascading effects on many ecological processes (MacGregor et al., 2004).

Elephants are not only ecologically important; they are economically and culturally significant. As noted earlier, their behaviour and cognitive abilities make them a valuable species, and popular to many people globally. In areas across Africa, elephants are economically important as they are a species of interest for tourists, where ecotourism benefits local communities (Jones and Barnes, 2007; Libanda and Blignaut, 2007). Novelli et al. (2006) argued that this knowledge may contribute to shaping societies attitudes towards elephant conservation. The economic value of wildlife tourism in Africa, including elephants as a key species, is a significant proportion of their Gross Domestic Product (GDP) (Naidoo et al., 2016). The

COVID-19 pandemic has shown just how essential tourists are for wildlife conservation and the people who are reliant on that income across Africa (Lindsey et al., 2020). Newsome (2020) suggested that longer-term recession of wildlife tourism will lead to increased poaching and losses of wildlife. This only exacerbates the need for sustainable elephant conservation, for tourist dependent industries, local communities, and sustainable development across many African countries.

#### **1.4 Savanna ecosystems**

African savanna elephants (*Loxodonta africana*) are keystone species for savanna ecosystems (Western, 1989), helping promote diversity of both habitats and species (Brooks and MacDonald, 1983; Owen-Smith, 2006; Campos-Arceiz and Blake, 2011). African savanna ecosystems are comprised of both woody (trees and shrubs) and herbaceous (grasses) vegetation that determine the functioning of these ecosystems (Bond, 2008; Shannon et al., 2011). Abiotic drivers such rainfall and soil type influence the composition and structure of such savannas (Levick et al., 2009). Direct disturbance agents such as fire and herbivory can also contribute to changes in savanna structure (Levick et al., 2009) and ultimately ecosystem change (Shannon et al., 2008; Midgley, Lawes and Chamaillé-jammes, 2010).

Abiotic drivers such rainfall and soil type influence the composition and structure of such savannas (Levick et al., 2009). Direct disturbance agents such as fire and herbivory can also contribute to changes in savanna structure (Levick et al., 2009; Straver et al., 2009) and ultimately ecosystem change (Shannon et al., 2008; Midgley, Lawes and Chamaillé-jammes, 2010). Fire can be damaging to an ecosystem, but it can also benefit the regeneration of certain flora species that have fire resilience adaptations such as post-fire seeding and resprouting (Govender et al., 2006). Species that are not resilient are therefore particularly vulnerable to any fire events (Bond et al., 2001). Fire is a non-selective ecological driver, which can cause a

reduction in woody biomass by removing small seedlings, and larger well established woody tree species (Govender et al., 2006; Mapaure, 2013; Aleper et al., 2008). However, if the scale and frequency of fires are managed effectively then an ecosystem will recover. The influence of fire and herbivory on vegetation change is difficult to interpret, as complex feedback relationships exist (Holdo et al., 2009; Midgley et al., 2010).

Large herbivores that are found within savanna ecosystems include species such as white rhinoceros (*Ceratotherium simum*), African savanna elephant and giraffe species (*Giraffa* spp.). Of these, the African savanna elephant has been suggested to be the most prominent herbivore that can influence savanna vegetation structure (Owen-Smith, 1992; Kerley et al., 2006). Their feeding and movement behaviours can affect woody vegetation structure composition (Shannon et al., 2008) and resultantly cause an increase in shrubland. Therefore, the rate and extent at which savannas are being altered from woodland to shrubland because of large herbivores is of concern to specific countries, especially within the southern Africa states. Added to this, large herbivores can prove difficult to manage at various spatial extents due to their size, and their long-term impact on vegetation is not fully understood (Scholtz et al., 2014).

#### ***1.4.1 Elephant vegetation selectivity***

Elephants can have a significant influence on ecological processes (Oliveras and Malhi, 2016), as they can alter the structure and composition of landscapes (Dublin et al., 1995; Cumming et al., 1997; Laws 1970). Elephant impact on woody vegetation can lead to a reduction in the number of trees within savannas, which can also lead to the extirpation of some woody plant species (O'Connor et al., 2007). Their feeding and movement activities influence vegetation structure, as they debark trees, break large branches, and often uproot trees (Coetzee et al., 1979; Mosugelo et al., 2002; O'Connor et al., 2007; Owen-Smith and Chafota, 2012). African elephants select tree species with high concentrations of nutrients (Holdo, 2003), and large

volumes of foliage (Boundja and Midgley, 2010; Levick and Asner, 2013). Their feeding activity may not directly lead to tree mortality, but if the main stem is broken, the tree has been uprooted or more than 50% of the stem has been debarked, then these susceptible trees are more likely to die (O'Connor, Goodman and Clegg, 2007).

In addition, certain tree species are resilient to elephant impacts and respond with coppicing to hedge level growth (such as mopane, *Colophospermum mopane*) benefitting many browsing species such as mesoherbivores (Lewis 1991; Styles and Skinner, 2000). Depending on the severity of the impact some tree species can persist through time, but impact may increase vulnerability to other indirect disturbance agents such as fire (Croze, 1974; Leuthold, 1977; Pellew, 1983; Lock, 1993; Dublin, 1995; Western and Maitumo, 2004).

The cognitive behaviour of elephants means that they are highly selective in their foraging behaviours, where plant species selectivity and size are both considered (Vesey-Fitzgerald, 1973, Landman et al., 2008). Their selective behaviour means that only specific targeted

species are impacted, compared to that of other disturbance agents (Dohn et al., 2017). They select trees with large volumes of foliage to gain maximum energy output, and the level of impact has been suggested to depend on tree characteristics such as height and canopy width (Boundja and Midgley, 2010; Levick and Asner, 2013, Howes et al., 2020). Additionally, elephants have shown preference for marula (*Sclerocarya birrea*) trees greater than 6 m in height, where they can feed on the canopy foliage (Cook et al., 2017). This selective feeding behaviour has been shown to be the cause of large tree ( $\geq 5$  m) mortality across savanna biomes (Vanak et al., 2012; Helm and Witkowski, 2013). Research around this topic has been at the forefront of savanna ecology as concerns have been raised with the level of tree replacement of large woody tree species (Laws, 1970; Shannon et al., 2008).

### **1.5 Vegetation patterns within fenced reserves**

Elephant reintroductions to fenced reserves are possible due to their suitable conditions for translocated elephants from larger reserves (Gaugris and Van Rooyen, 2010; Mackey et al., 2006; Thouless et al., 2016). Fenced reserves play an integral role in conservation as they aid in the decrease of human-wildlife conflict, poaching and the spread of diseases from livestock (Boone and Hobbs, 2004; Pirie et al., 2017). However, within fenced reserves, suitability of land, elephant densities and vegetation sustainment are often met with apprehensions (Lombard et al., 2001; Stretch et al., 2002; Caughley, 1976). Fences can influence the movement patterns of wildlife, as species that are constrained cannot move to areas with more abundant resources (Hopcraft et al., 2010). This can be especially important during the dry season when there is lower levels of forage available and water availability can become reduced (Oates and Rees, 2013; Skarpe, 1992; Tefempa et al., 2008). This can in turn effect large herbivore distributions within these confined spaces (Chamaille-Jammes et al., 2007; Redfern et al., 2003).

Hence, understanding how large herbivores utilise landscapes is important for reserve management. Concerns on the loss of trees  $\geq 5$  m tall across Africa's protected areas has been partly attributed to impacts by savanna elephants, which in turn is likely to be influenced by the proximity of fences and artificial water points (AWP). Large savanna trees ( $\geq 5$  m) are considered important in savanna structures, as they provide resources (Dean et al., 1999) and enhance spatial heterogeneity (Manning et al., 2006). The future of the large trees persisting within fenced reserves is often thought as unsustainable due to elephants browsing behaviours within these confined spaces (Slotow et al., 2005). For example, Cook et al. (2017) found that within the Jejane Private Nature Reserve (JPNR), South Africa, elephant reintroductions lead to an increase of the annual mortality rates of marula trees. Cook et al. (2017) stressed the lack of seedling regeneration as a result of elephant impact, which needs to be further quantified.

### ***1.5.1 Elephant impact to vegetation in South Africa***

An increase in elephant population density results in an increased demand on resources, where under natural conditions (free roaming and no land use restrictions), this increase would cause a range expansion or distribution shift. However, this is not the case throughout South Africa where there are high elephant population densities in confined spaces (Landman et al., 2008). This is due to suitable areas contracting because of human encroachment, and protected areas and fenced game reserves have been designated by management approaches (Lehmann et al., 2008; Lombard et al., 2001). There is increasing pressure on managed fenced reserves to support elephants sustainably, as there is little known on the long-term effects of maintaining elephants on vegetation within these fenced areas at different spatial and temporal extents.

Historically, studies on elephant impact in South Africa have been based around the Kruger National Park (KNP) to determine the decline in woody species and reduce the impacts of elephants (Eckhardt, Wilgen and Biggs, 2000; Levick et al., 2009). KNP is a renowned

stronghold for African savanna elephants, and it is the largest protected area in South Africa (~20 000 km<sup>2</sup>). Elephant populations have significantly increased over the last 20 years, where 17 086 elephants were resident in 2015, growing at 4.2% per annum (Ferreira and Simms, 2017). Van Wyk and Fairall (1969) stated that the most important tree species in KNP were red bush willow (*Combretum apiculatum*), clusterleaf (*Terminalia sericea*), knob thorn (*Sengalia nigrescens*), marula and mopane (*Colophospermum mopane*). These five tree species constituted about 80% of the tree species population within this area. The potential impact of elephants on marula trees within the KNP, due to their ecological significance, resulted in a research project in 1979 to determine the extent of this (Coetzee, et al., 1979). This study indicated that the impact at the time of study did not threaten the marula population. Contrary to these outcomes, a later study by Trollope et al. (1998) recorded declines in the woody vegetation of between 1960 and 1989, suggesting that this could be due to the extreme increase in elephants during this time as well as other environmental factors.

There have been extensive studies on the impact of elephants to tree species in open (unfenced) reserves (Holdo, 2003; Morrison, Holdo and Anderson, 2016). However, there are relatively few including Gaugris and Van Rooyen (2010), Kerley and Landman (2006) and Howes et al. (2020) that have focused on the impact that elephants have on tree species within small (< 200 km<sup>2</sup>) fenced reserves. Additionally, these studies have not considered the recovery levels of trees at different heights, after impact from elephants occurs. The recovery levels on tree species after elephant impacts occurs has also not been addressed in other studies within fenced areas, that focus on a single species (Gadd, 1997; Jacobs and Biggs, 2002). Thus, it is essential that we can determine recovery levels, such as coppicing and regrowth of trees after elephant impact occurs, so that reserve managers can apply appropriate mitigation strategies to sustain tree species to support elephant survival. This thesis aims to contribute to this gap in knowledge.

At a small reserve extent (< 200 km<sup>2</sup>), the impact that elephants have on marula tree populations has been recorded in South Africa (Gadd, 1997; Weaver, 1995; Cook et al., 2017). Gadd (1997) found that marula trees were significantly impacted by elephant browsing, and recruitment and regeneration of these trees were weak. Weaver (1995) found that elephant impact was significant in both marula and knob thorn trees. Various studies on the effect of elephants within enclosed reserves have been conducted, but the density of elephants relative to the reserve size differs greatly, and one single tree species is often considered (Ruess and Halter, 1983; Jacobs and Biggs, 2000; Coetzee et al., 1979).

### ***1.5.2 Secondary impacts to vegetation***

Elephant impact to vegetation has to date, been focussed on the abundance of large trees due to their environmental and economic significance (Shannon et al., 2008). Elephants can cause branch breaking and debarking, which has been used to determine levels of non-severe to severe impact (Boundja and Midgley, 2010; MacGregor and Connor, 2004; Seloana et al., 2017), often derived from the Walker eight-point scale (Walker, 1976). Walker's (1976) method is used to record vegetation composition and utilisation. The types of elephant impact can enhance tree susceptibility to secondary impacts such as termites (*Coptotermes* species), woodborers (*Cerambycidae* species) or other insects, which may shorten trees' life span (Hatcher, 1995).

Termites can penetrate trees through fractures in the bark (Gould et al., 1993) and establish secondary nests inside the tree's cavities (Harris, 1968). This can occur more readily once the tree bark has been stripped by elephants (Helm et al., 2011) (Figure 1.2). Additionally, wood borer damage to trees is characterised by the woodboring activity of larvae and adult beetles in the stems and branches of damaged or stressed hosts (Halperin and Geis, 1999; Peters et al., 2002; Nair, 2007). Adult mortality of marula trees has been suggested to be attributed to rapid

invasion by wood borers after bark stripping occurs and the sapwood is exposed (Helm et al., 2011; Coetzee et al., 1979; Guy, 1989; Jacobs and Biggs, 2002). This secondary impact may therefore hinder the trees' ability to recover, effecting the survival of trees within savanna ecosystems (Holdo, 2007). Therefore, it is important that we understand how tree species recover after secondary damage occurs, which has not yet been identified.

To date, there are relatively few studies that have considered the effects of elephants on invertebrates. Disturbance from elephants has been shown to influence dung beetle species diversity and biomass (Botes et al., 2006). Trees that are toppled by elephants may also provide refugia for species such as ground dwelling invertebrates (Govender, 2005). Musgrave and Compton (1997) considered the composition of phytophagous insect communities within Addo Elephant National Park (AENP), finding on average greater levels of impact on plants which were browsed by elephants. This was suggested to be caused by the change in nutrients as a result of severe pruning, causing rapid regrowth which is more palatable than unbrowsed material. However, the Kaffrarian succulent thicket habitat in AENP is unique in terms of plant cover, consisting of mainly evergreens (Stuart-Hill, 1992). Therefore, there is still a need to examine the effects on other different types of woody vegetation. The direct relationship between elephant impact and the subsequent secondary impact and tree recovery levels have yet to be fully understood. This thesis will aim to identify this gap in knowledge, to contribute to sustainable management approaches of elephants and vegetation within fenced reserves.



**Figure 1.2** Field recordings of elephant impact and secondary damage to tree species within the KPGR (2019)

### ***1.5.3 Methods to determine elephant impact***

To date studies on elephant impact have used field-based methods for data collection on trees biophysical and vertical structural parameters (Nkosi et al., 2019) (Figure 1.3). Historically, field-based methods have been used in ecological studies to provide detailed ecological data at small extents (Simms, 2009; Buchanan et al., 2013) which are often focused on specific tree species. For example, Cook et al. (2019) considered elephant impact over time on marula trees, using field-based methods in an area where elephants were excluded, and later reintroduced to a site within the KNP. This attempt to address the vegetation change over a fixed time, was dependent on accurate, replicable data acquisition, which is often not the case on large spatial areas, and in turn can hinder research attempting to address temporal changes to vegetation. Added to this, field-based methods are impractical and costly to implement for large data sets that are required on a regular basis to develop effective assessments for conservation management outcomes.



**Figure 1.3** Field recordings of elephant impact to *Vachellia robusta* tree within KPGR, South Africa (2018). A staff with known height is used to determine tree metrics via *VolCalc* (Barrett and Brown, 2012)

Remote sensing techniques offer an alternative approach, that are not only cost-effective for application in large areas (Duro et al., 2007), but can also enable detailed ecological insights depending on the technique that is applied. Remote sensing methods at a 2-D scale using aerial photography, have been used to determine South African vegetation imaging landscape for many years. Mutanga and Ahmed (2016) identified that studies have focused on long term vegetation cover changes (Eckhardt, Van Wilgen, and Biggs, 2000; Russell and Ward, 2014), bush encroachment (Hudak and Wessman, 1998; Sirami, Seymour, Midgley, and Barnard, 2009; Wigley, Bond, and Hoffman, 2010), and biodiversity (Cumming et al., 1997).

Monitoring long-term savanna woody cover dynamics is possible using remotely sensed imagery, which could offer opportunities for conservation authorities.

To date, texture analysis has been applied to analyse woody vegetation using aerial photographs and satellite images (Asner et al., 2003; Hudak and Wessman, 1998). In many cases, optical imagery is used which can have limitations if imagery has clouds, shadow, low spatial variability and being 2-D in nature which may be insufficient for providing the necessary information to determine tree metrics (Kachamba et al., 2016). To derive tree metrics at a 3-D scale in savanna ecosystems, airborne laser scanning (ALS) methods have been used with light detection and ranging (LiDAR) scanners within the Kruger National Park (KNP) (Asner et al., 2009; Levick et al., 2009). ALS techniques are increasingly being used for the measuring and monitoring of 3-D vegetation structure (Lefksy et al., 2002), within these heterogeneous savanna ecosystems (Asner et al., 2009; Johansen et al., 2010; Levick et al., 2009; Levick and Rogers, 2008; Levick and Asner, 2013; Davies, Gaylard and Asner, 2018). However, ALS applications are extremely expensive to disseminate on large temporal and spatial extents, so alternative methods are required to identify and analyse 3-D tree structures.

The performance of 3-D photogrammetry applied to aerial photographs for the retrieval of savanna tree attributes, has not been studied extensively. There is potential for using aerial photography to derive 3-D vegetation dynamics to quantify tree heights at a landscape scale which could provide an economically viable approach. The use of aerial photographs within savanna ecosystems could potentially offer an opportunity to utilise an accessible resource to inform best practises for management approaches. Therefore, this thesis will address this gap in knowledge.

## **1.6 Approaches to elephant management**

The optimal size of nature reserves has been the subject of debate for many years (Soule and Simberloff, 1986; Noss and Cooperrider, 1994; Shafer, 1995; Soule and Terborgh 1999; Margules and Pressey, 2000). Small game reserves (< 200 km<sup>2</sup>) can help to protect and conserve ecological spaces and the species within them. Evidence suggests that privately owned conservation areas can protect biodiversity and are profitable (Mitchell et al., 2006; Kramer et al., 2002).

The need for private game reserves has increased over recent decades due to land becoming increasingly fragmented as a result of human development (van Hoven, 2015). This has meant that there is an increased need for small (< 200 km<sup>2</sup>) enclosed reserves. As these areas are enclosed, reserve managers need to effectively monitor fauna populations. This is especially important for elephants, who are unable to immigrate and emigrate to other areas in response to resource distribution within these fenced areas. Added to this, the movement patterns of elephants have been shown to be influenced by the proximity of the fence line, suggesting an increase in habitat utilisation within the centre of a reserve (Vanak et al., 2010). The direct effect of fences in relation to herbivore behaviour within small, fenced reserves has yet to be identified and needs further investigation (Hayward, 2009) which we will consider in this study. A greater understanding of elephant impact to vegetation within enclosed, small reserves is needed, to support to the conservation of elephant populations within South Africa.

Management approaches for reserves varies spatially as there are many factors that can influence the utilisation of trees and the level of elephant impact (Ntumi et al., 2005; Henley and Cook, 2019). Current methods include monitoring elephant's spatial movement activities, for both high- and low- level impacts (Loarie et al., 2009). Within South Africa, African savanna elephants have co- existed on managed fenced reserves (Asner et al., 2016). However, concerns over their impact on large trees, has seen a significant rise over recent years, and management has focussed on protecting endemic species and maintaining the natural system to

an ‘ideal’ state (Kerley et al., 2010; Cumming et al., 1997). The ideal state has been based on historic records as well as tourist perceptions (Lomard, 1995).

Factors that impact management decisions include the presence of fences and AWP (Hisocks, 1999). Elephants have sophisticated cognitive abilities and a significant memory capacity, where they follow a natural migratory pattern passed down from generations of matriarchs when they can roam freely. Fences often prevent this behaviour, and roads can act as pathways which they use to move through an area (Cushman, Chase and Griffin, 2010). During the dry season, AWP serve as a permanent refuge within confined areas when natural resources dry up. These abiotic management methods can cause elephants to be confined within smaller areas, limiting resource distribution and overutilisation of certain plant species (Loarie et al., 2009).

Recent research has demonstrated how mitigation approaches to elephant management has changed from directly managing elephant numbers by invasive methods, to elephant distributions being managed in relation to gradients of their impact on vegetation (Cook and Henley, 2019; Henley and Cook, 2019). This method considers solutions to large-scale landscape manipulation, promoting interconnectivity and reducing fragmented landscapes. Protected areas such as the KNP is using this mitigation strategy across certain areas to protect large trees ( $\geq 5$  m) from elephant damage, but this landscape scale approach is a viable option for this site due to its size (~20 000 km<sup>2</sup>) and is not viable for smaller reserve extents. Mitigation measures for small reserves need to consider a combination of options such as translocations and contraceptive programmes, but there is a significant lack of knowledge of the implications that these measures have long term on a small-scale reserve (Duffy, 2002). Added to this, the long-term implications of these mitigation methods on elephant’s societal change have not yet been evaluated (Nyakaana et al., 2001; Wittemyer et al., 2009).

Even though many of the challenges faced by elephants are similar, the approaches to their conservation and management vary greatly. Within South Africa, savanna elephants are numerous, and populations are growing. Their populations are managed within Private Nature Reserves and National Parks, where natural mortality may not suffice to control elephant numbers and meet conservation goals (Woolley et al., 2008). Therefore, management is required to regionally monitor elephant populations (Owen-Smith et al., 2006). Translocating elephants is an option, but this is only possible with areas of suitable habitat and elephant numbers in the area is appropriate. Management methods in areas where high elephant densities are found include culling, contraception (Fayrer-Hosken et al., 2000; Delsink et al., 2006), or creating habitat corridors and increasing available areas within proximity is also effective (Osborn and Parker, 2003).

### **1.7 People and elephant conservation**

African elephant population declines have historically given rise to many conservation organisations, both *in-situ* and *ex-situ*. In-situ research-based organisations such as non-governmental organisation (NGOs), are often reliant on funding from international funders and donors to support research and conservation practises. Long standing elephant conservation organisations such as ‘Save the Elephants’ have been at the forefront of elephant conservation. Partnering both research on African elephants and frontline monitoring has enabled this organisation and many organisations alike to work with and directly conserve elephant populations. Rangers, conservationists, and anti-poaching organisations, are at the frontline of elephant conservation, tirelessly protecting and monitoring these animals. This includes anti-poaching patrols, monitoring fence lines and anti-snare procedures.

Approaches to elephant conservation differ between cultures, where community involvement has been applied across savanna environments (Sitati, 2012; Sitati and Wapole, 2006; Parker

and Osborn, 2006). Community involvement views local participation as a prerequisite to sustainable conservation (Ashrnafi and Leader-Williams, 2005). Local knowledge holds intrinsic values and importance to science and conservation. With community knowledge associated with their immediate environment, this can prove vital to improve ongoing ecological research and conservation programmes. Therefore, community involvement is essential for passing on valuable knowledge to future generations effectively to people both *in-situ* and *ex-situ* for effective elephant conservation.

### ***1.7.1 Environmental education***

The ability to conduct relevant scientific research on conservation issues is important to determine the extent of how ecosystems will be affected. This information is key to enable effective communication with decision makers and stakeholders, but this alone is not sufficient to solve complex conservation issues (Kansky and Knight, 2014). Environmental education (EE) is a key conservation strategy that can be applied to important conservation topics (Kapur, 2017). This method provides environmental awareness information that can influence individuals and communities, to students of all levels. EE can be carried out at all ages, however EE practises targeted at school children is important, as they are not likely to have fully formed opinions or attitudes about the environment (Pelletier et al., 2004). Studies have highlighted the importance of targeting children, for nurturing positive environmental behaviours and help spread positive effects of EE programmes to a wider demographic (Rakotomamonjy et al., 2014; Damerell et al., 2013).

Environmental education is engrained in conservation programmes such as protecting nature and wildlife (Athman and Monroe, 2001; Fraser et al., 2015). This is especially important when considering endangered species that need effective conservation, as these species would likely not survive without intervention strategies. Elephants are one such species, which have been described

earlier in this chapter. They are faced with challenges such as land fragmentation, poaching and habitat destruction, causing population numbers to decline (Leimgruber et al., 2003; Nelson, Bidwell, and Sillero-Zubiri, 2003). Understanding children's perception of elephant conservation could improve conservation education delivery, enhancing children's connection to elephants which is crucial for their long-term conservation (Ardoin and Bowers, 2020).

Environmental education programmes based on environmental issues need to be carefully applied depending on how school children perceive concerns around elephant conservation. This can only be achieved if we have a clear understanding of school children's perception, *in* and *ex situ* of elephants which will likely be influenced by the level of exposure to elephants. This has yet to be identified, where studies have often focussed on adults within local communities and their response to Human Elephant Conflict (HEC) (Hart and O'Connell, 1998; Bandara and Tisdell, 2003; Raihan Sarker and Røskaft, 2010; Western et al., 2015). This is important to establish, however determining how school children perceive elephants is an essential method in supporting long-term management of elephants. By understanding school children's perceptions, we can better understand their attitudes towards elephant conservation and provide support and guidance to critical areas.

## **1.8 Project Rationale**

A greater understanding of elephant impact to vegetation within enclosed, small reserves is needed, to determine the long-term sustainability of vegetation and ultimately, how this understanding can contribute to the conservation of elephant populations within South Africa. There have been various studies on the impacts that elephants have on vegetation at this scale, but these have been limited to single species and lack important information on the recovery of tree species that exhibit elephant impact. Determining the level of tree recovery after elephant impact occurs is essential to be able to maintain vegetation with elephant presence in

enclosed areas. Additionally, the fenced nature of enclosed reserves has also been overlooked as a potential determinant of impact levels in many studies which requires further investigation. Therefore, the levels of impact and recovery in response to elephant habitat use and fence line effect is the focus of Chapter 2.

Added to this, the increasing elephant densities in fenced reserves (Skarpe et al., 2004; Shannon et al., 2008) and the decreasing numbers of large trees (Ben-Shahar, 1993; Eckhardt et al., 2000; Moncrieff et al., 2008), has resulted in a need for improved knowledge of how elephants and other subsequent disturbances can result in tree death (Holdo, 2005). Tree species are faced with several biotic constraints such as insects (Wargo, 1996; Hakeem et al., 2012), and this secondary impact partnered with the lack of tree recovery, has been overlooked in many studies focussed on elephant impact within fenced reserves. Therefore, there is need to determine how insect presence relates to tree impact and how this affects tree recovery. This is the focus of Chapter 3.

To be able to effectively predict the effects that elephants have on vegetation, field data collection and statistical analysis techniques have been used in this PhD research to provide quality data visualisation and management. Mixed-effect models using statistical analyses programmes have been used effectively to predict tree species response in previous studies considering for elephant impact. However, multi-model analyses on multiple tree species need to be explored further to determine the effects of elephants and vegetation recovery (Chapter 2), considering for the secondary effects of elephant impact (Chapter 3).

Field based methods used to monitor impact are valuable, however distance-sampling techniques including remote sensing can be applied to determine tree metrics. To date, most remote sensing studies concerning elephant impact have utilised coarse scale remote sensing methods including multispectral images such as Landsat and "Satellite pour l'Observation de

la Terre" (SPOT) images (Munyati and Sinthumele, 2016). However fine scale, remote sensing techniques are limited and determining the 3-D structure of trees has relied on ALS, and a combination of aerial imagery and high-resolution satellite imagery (Asner et al., 2009; Levick et al., 2009). The performance of 3-D photogrammetry applied to aerial photographs for the retrieval of savanna tree attributes and vegetation height at a reserve extent has not been studied extensively. This thesis will apply some of these methods and explore the efficacy of using 3-D photogrammetry from aerial photographs to determine vegetation height at a reserve extent. This is the focus of Chapter 4.

Determining how elephants impact vegetation on a temporal and spatial extent is important in achieving direct elephant conservation by maintaining areas with suitable elephant numbers and adequate vegetation. However, elephants are threatened by habitat fragmentation because of human encroachment. This in turn increases human elephant conflict (HEC), where people living near elephants can come into conflict with elephants, and therefore attitudes towards elephants can often be negative. Communities living near elephants and wildlife conservation areas need improved education to become engaged with conservation practices and value viable careers in conservation. Additionally, participation in *ex situ* education programmes is important for increasing awareness and encourage funding for elephant conservation (Makecha and Ghosal, 2017). This will improve understanding of conservation, methods of mitigating HEC, and will also encourage opportunities within the wildlife tourism industry, ultimately supporting elephant conservation.

There has been little focus on determining how school children perceive elephants and elephant conservation, both *in situ* and *ex situ* (Makecha and Ghosal, 2017) School children are a key demographic, as they are not likely to have fully formed opinions or attitudes about the environment (Pelletier et al., 2004). Additionally, understanding how school children perceive elephant conservation could improve conservation education delivery, enhancing children's

connection to elephants which is crucial for their long-term conservation (Ardoin and Bowers, 2020). Chapter 5 of this thesis will therefore also consider how school children perceive elephant conservation at different geographic locations (U.K., Kenya and South Africa) to determine how elephants and elephant conservation are perceived.

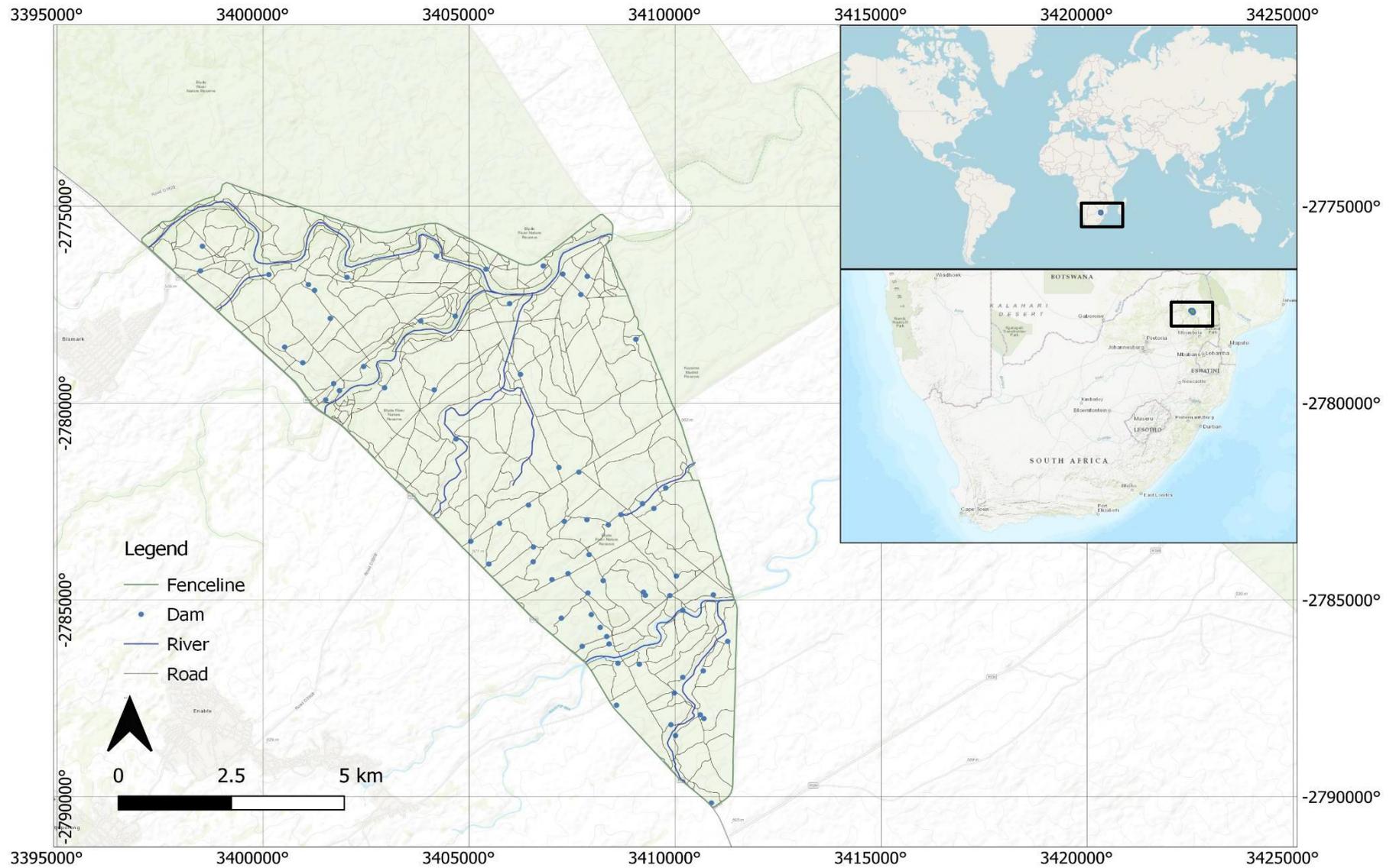
### **1.8.1 Study areas**

Both fieldwork data collection and distance-sampling techniques were conducted on Karongwe Private Game Reserve (KPGR) in Limpopo, (South Africa) for Chapters 2, 3 and 4 (Figure 1.4). KPGR is a small, fully fenced 7,960-hectare reserve within the savanna biome in the Limpopo Province of South Africa (S24.227061, E30.603302) and has a mean annual rainfall of 790 mm (Low and Rebelo, 1998). Temperatures range from 0 °C to 45 °C, with an average of 22 °C. KPGR falls within a summer-rainfall area where rainfalls mainly between October and April.

The reserve originally consisted of 10 individual private farmlands, but the fences were removed in 1999 and various game species were introduced, including African savanna elephant, lion (*Panthera leo*), leopard (*Panthera pardus*), cheetah (*Acinoyx jubatus*), and white rhinoceros (*Ceratotherium simum*), to establish a game reserve. The reserve is bordered on all sides by public roads, all of which are within 50 m of the fence line. The western fence line (19.1 km) runs along the paved R36 route, while gravel roads run parallel along the eastern (14.5 km) and northern (11.9 km) fence lines. The R36 acts as the main passage between Hoedspruit and Tzaneen, the two larger cities in the vicinity of the reserve.

Within the KPGR the altitude above mean sea level (AMSL) varies from 489 m in the east to 520 m in the west to (Lehmann et al., 2008) and therefore drainage predominantly flows from west to east. Makhutswi River in the north is the only perennial river. The Kuvyenami and Mafunyane Rivers drain the centre, and the Korongwe and Mathumi Rivers drain the south of

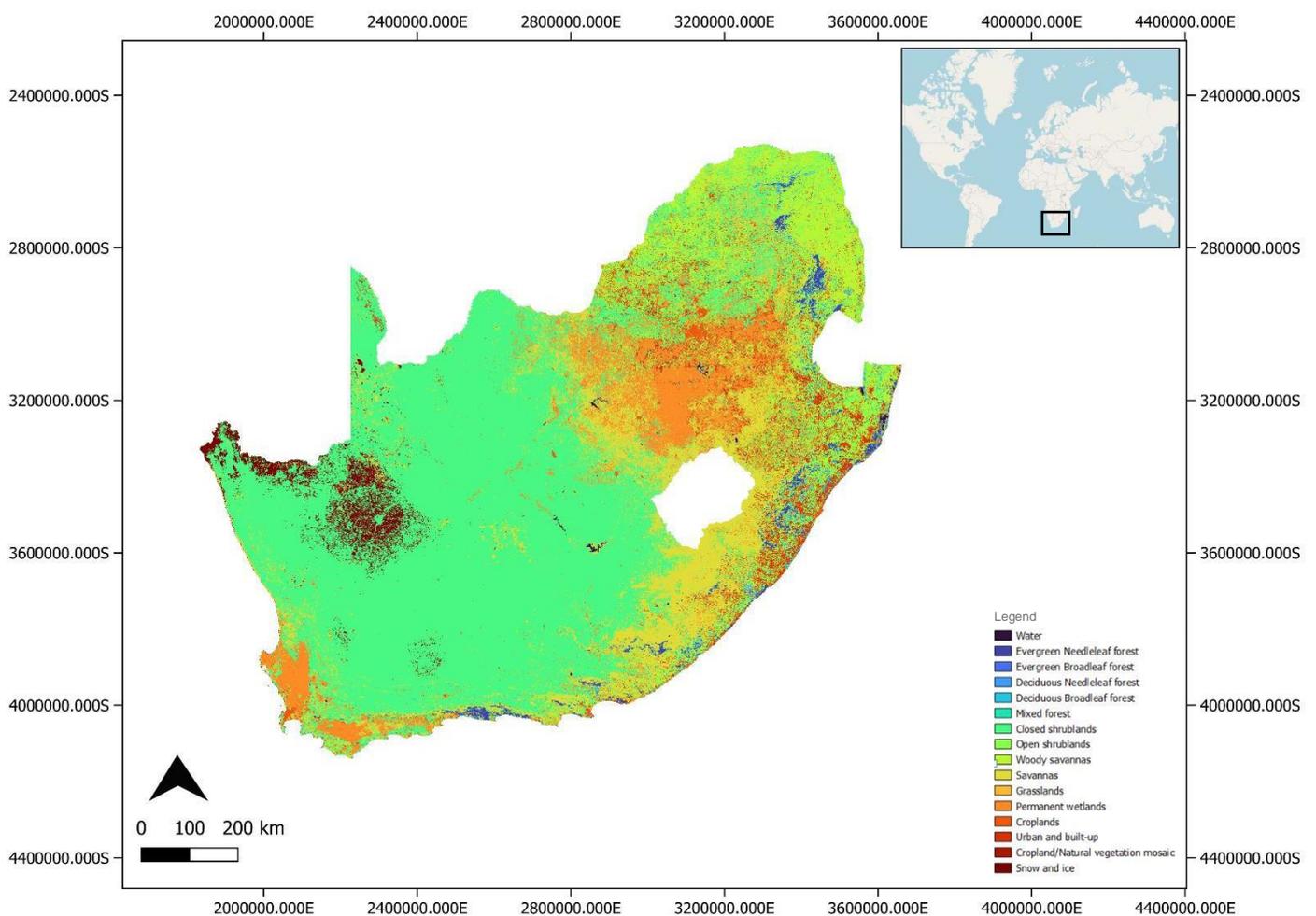
the reserve. All these rivers join up with the Olifants River. There are also large and small koppies (rocky outcrops) are distributed around the reserve.



**Figure 1.4** Location of Karongwe Private Game Reserve (KPGR) in Limpopo, South Africa. (Map compilation by the author using data via ESRI. Projection: Geographic. Datum: WGS84)

### 1.8.2 Vegetation type

KPGR is within the savanna biome (Rutherford and Westfall, 1994) within the Mixed Lowveld Bushveld (Low and Rebelo, 1998) (Figure 1.5) The vegetation patterns across the reserve are bush on the uplands, open tree savanna in the bottomlands, and there is dense riverine woodland on the riverbanks (Lehmann et al., 2008). The shrub layer and grass layer are moderately developed (Low and Rebelo, 1998). Across the reserve, there are many dolerite intrusions and areas covered by gabbro. Basement gneisses and granite underlie this region (Low and Rebelo, 1998). Knob thorn and marula trees dominate this savanna biome, and *Combretum* spp. are also prevalent.



**Figure 1.5** Land cover classification of South Africa (Map compilation by the author using data via MODIS and IPUMS. Projection: Geographic. Datum: WGS84)

### ***1.8.3 Elephant population on KPGR***

The elephant population on KPGR has increased since their reintroduction in 1999. They were first introduced in 1999 when management released one adult male and four adult females, all of which still live on KPGR. Four more elephants have since been introduced to the reserve and 19 elephants were born, of which five have died and two have been relocated. From 2011, the elephant population has consisted of one stable family unit ( $n = 18$ ), defined as all adult females, all subadults and juveniles of both genders (Poole, 1989) plus three adult bulls until 2015 (one adult bull was translocated) (Table 1.1). Two male elephants were administered with the gonadotrophin releasing hormone (GnRH) vaccination in 2012. GnRH vaccinations is a non-invasive contraceptive measure for managing wildlife populations, where fertility control is warranted in smaller, fenced game reserves with populations in high densities (Lueders et al., 2017). In 2016, both males were no longer administered with contraception, and an elephant birth in 2019 brings the total population to 21 elephants (20 elephants at the time of field study for Chapter 2, 3 and 4). Like other elephant populations, the family unit (from now on to be referred to as the breeding herd), remained together, while the three adult bulls occasionally associated with each other and the herd since their reintroduction (Poole, 1989; Western and Lindsay, 1984). More recently, the adult bulls have been seen to associate with the breeding herd, and field observations now rarely observe the breeding herd and bull elephants separately (Appendix 3: Figure A.3; see also Chapter 6). Owen-Smith et al., (2006) suggests that an effective elephant population density is  $0.28 \text{ km}^2$ , and therefore, KPGR should support 22.28 individuals.

**Table 1.1** Elephant populations on Karongwe Private Game Reserve (KPGR, South Africa)

since their introduction in 1999. Since 2016, E1 and E3 were no longer administered with the GnRH vaccination.

<b>Animal Name</b>	<b>Sex</b>	<b>D.O.B</b>	<b>Translocated from</b>	<b>Reintroduction date</b>	<b>Removed/translocated date</b>
<b>Gattis</b>	M	1990	Kapama	24-Jul-99	13-Apr-04
<b>Madala</b>	M	1978	Kapama	24-Jul-99	13-Apr-04
<b>E2 - Mr M</b>	M	1982	Kapama	24-Jul-99	Oct-15
<b>E1 - Fumbe</b>	<b>M</b>	<b>1980</b>	<b>Kapama</b>	<b>24-Jul-99</b>	
<b>E3 - Flippy</b>	<b>M</b>	<b>1986</b>	<b>Magudu, KZN</b>	<b>Jul-99</b>	
<b>E4 - Tsuku</b>	F	1987	Magudu, KZN	Jul-99	
<b>E5 - Nick</b>	F	1985	Magudu, KZN	Jul-99	
<b>E6 - Telipha</b>	F	1987	Magudu, KZN	Jul-99	
<b>E7 - Duma</b>	F	1985	Magudu, KZN	Jul-99	
<b>Pula</b>	M	05-Oct-00			Aug-08
<b>Mafunyane</b>	M	23-Oct-00			Aug-08
<b>E7.1 - Lavara</b>	F	13-Jul-00			
<b>UK</b>	M	05-Jul-02			Feb-10
<b>UK</b>	M	29-May-03			Feb-10
<b>E5.1</b>	F	29-Jun-03			
<b>E4.1</b>	M	28-Oct-03			
<b>E6.1</b>	F	14-Aug-04			
<b>E7.2</b>	UK	12-Aug-05			
<b>E4.2</b>	F	08-Apr-06			
<b>E5.2</b>	F	26-Jun-06			
<b>E7.3</b>	F	Nov-07			
<b>UK</b>	UK	Nov-07			
<b>E4.3</b>	F	Dec-08			
<b>E5.3</b>	UK	Jan-09			
<b>E6.2</b>	F	Feb-09			
<b>E7.4</b>	UK	May-09			
<b>E7.1.1</b>	UK	Dec-11			
<b>E5.1.1</b>	UK	Dec-11			
<b>E5.4</b>	UK	25-Oct-19			

#### **1.8.4 Environmental education**

We indicated earlier to the importance of school children’s perceptions of elephants and how that can influence the likelihood of them wanting to protect elephants. This thesis will also aim to contribute to the understanding and perception that school children have with elephants at different geographic locations. We selected school children as our research demographic, as understanding children’s perception of elephant conservation may facilitate improved methods of conservation education delivery (Ardoin and Bowers, 2020). Schools were selected based on their location with regards to elephant presence or absence and in different countries (Table 1.2; see chapter 5 for full details).

**Table 1.2** Selected study site locations

<b>Study location</b>	<b>Elephant exposure</b>	<b>Land use</b>
<b>England: Wrenn School, Wellingborough</b>	In captivity (zoos)	Urban environment
<b>Kenya: Brainhouse School, Mathare North, Nairobi</b>	Predominantly open reserves	Rural and urban environment
<b>South Africa: Diputhi School, Limpopo</b>	Fenced reserves only	Mostly rural environment

## 1.9 Research aims and objectives

Using African savanna elephants (*Loxodonta africana*) as the study species, the main aim of this thesis is to:

- *Quantify the patterns of the direct and indirect impacts elephants have on vegetation and determine the levels of tree recovery using field data collection.*

Additional aims are to:

- *Explore distance sampling methods for detecting tree attributes within a fenced reserve.*
- *Quantify the extent at which school children perceive elephant conservation, considering for their importance at an economic and environmental level, and their perception of wildlife conservation careers.*

The research objectives (O) are:

**O1. Assess how field data collection and modelling approaches used for studying elephant impact on tall trees (> 5 m height) can be applied and expanded to additionally identify the recovery levels of multiple tree species within a fenced reserve;**

**O2. Identify how field data collection and modelling analyses can be used to determine the secondary effects of elephant impact on trees > 2 m, and how this influences tree species recovery level;**

**O3. Determine the ability of photogrammetric point clouds derived from conventional aerial survey for estimating tree heights;**

**O4. Identify whether school children in countries with and without elephant presence perceive elephants as a threat or as an animal that they value both economically and environmentally.**

The research objectives are met in the data chapters as follows:

**Chapter 2: Modelling the impact of elephants on vegetation within a fenced reserve, using impact and recovery levels to assess the influence of these variables on tree height, identified elephant range use and fence line (O1)**

**Chapter 3: Investigating how insect presence can affect the level of recovery on different tree species at various heights after elephant impact occurs, using modelling approaches (O2)**

**Chapter 4: Applying 3-D photogrammetry from historical aerial photographs to test how effective this method is in identifying tree height for elephant impact studies (O3)**

**Chapter 5: Quantifying how children perceive elephant conservation through questionnaires across locations with elephant presence and elephant absence (O4).**

## **2. Impacts of African savanna elephants (*Loxodonta africana*) on tall trees and their recovery within a small, fenced reserve in South Africa**

- *Objective 1: Assess how field data collection and modelling approaches used for studying elephant impact on tall trees (> 5 m height) can be applied and expanded to additionally identify the recovery levels of multiple tree species within a fenced reserve.*
- *Main findings: Trees in high-use areas were significantly less likely to show signs of debarking and push over. Tall trees were significantly more likely to be impacted by elephants, being associated with heightened risks of debarking and branches being broken but lower risks of being pushed over. Trees close to the fence line were not more impacted than trees near the centre of the reserve. The level of use, distance to the fence and tree height were not significant predictors of tree recovery indicators. Therefore, we suggest that future mitigation efforts should focus on trees with high levels of impact and low levels of recovery.*

### **2.1 Abstract**

African savanna elephants (*Loxodonta africana*) can have detrimental impacts on trees due to their feeding habits. To date, studies exploring elephant impact on trees and their recovery levels within small reserves restricted by fences, have lacked focus in determining elephant high-use areas, the direct effect of tree recovery. The aim of this study is to assess whether elephants cause significant impact on several tree species in the small fenced Karongwe Private Game Reserve (KPGR) and to determine levels of tree recovery. We analysed the level of impact using vegetation transects, where all trees  $\geq 5$  m in height were surveyed (n=634 trees). Elephant location data were used to identify high- and low-use areas. Thirty-two tree species were recorded, with 5 species accounting for 80%; these were used for further analysis. Trees

in high-use areas were significantly less likely to show signs of debarking and push over. Tall trees were significantly more likely to be impacted by elephants, being associated with heightened risks of debarking and branches being broken but lower risks of being pushed over. Trees close to the fence line were not more impacted than trees near the centre of the reserve. The level of use, distance to the fence and tree height were not significant predictors of tree recovery indicators. Therefore, we suggest that future mitigation efforts should focus on trees with high levels of impact and low levels of recovery.

### ***Keywords***

Karongwe Game Reserve, *Loxodonta africana*, Tree impact, Tree recovery, Space use, Elephant density, Fence

## **2.2 Introduction**

African savanna elephants are keystone species for savanna ecosystems (Western, 1989), helping promote diversity of both habitats and species (Brooks et al., 1983). Elephant populations have been declining in range and numbers for decades due to ivory poaching, habitat loss and land fragmentation (Lemieux and Clarke, 2009; Thouless et al., 2016). African savanna elephant populations have decreased by at least 60% over the last 50 years, according to the latest IUCN assessment (Gobush et al., 2021). To counteract these trends, countries such as South Africa have engaged in major elephant reintroductions to protected areas, many of which include conversion of agricultural land to fenced reserves (Lehmann et al., 2008; Lombard et al., 2001). This method has proved successful in maintaining and increasing elephant population numbers; however, suitability of land, elephant densities and vegetation sustainment is often met with concerns (Stretch et al., 2002; Caughley, 1976).

In an open ecosystem without land use restrictions, elephant herd size varies depending on resource availability (Young et al., 2009). The impact of elephants on tree species in open

reserves (unfenced) has been well studied (Holdo, 2003; Morrison, Holdo and Anderson, 2016). Tree species composition of an area has been shown to change when elephants are present in high enough densities to modify their habitat (Cummings et al., 1997). To determine the level of elephant impact, research has focussed on high profile species such as marula trees (*Sclerocarya birrea*) (Gadd, 1997; Weaver, 1995; Jacobs and Biggs, 2002; Wiseman et al., 2004). Trees with larger stems (> 10 cm in diameter) are more likely to be selected by elephants for debarking, whilst smaller trees are more likely to be toppled (Boundja and Midgley, 2010; Ihwagi et al., 2012; Ssali et al., 2013). Gaugris and Van Rooyen (2010), Kerley and Landman (2006), Landman et al. (2008) and more recently Howes et al. (2020), have documented the impact that elephants have on small reserves, where natural elephant feeding behaviours are restricted by fences.

Spatial restriction of elephants within fenced reserves can exacerbate their impacts on their habitat (Laws, 1970; Hoare, 1999; Baxter, 2005). Fencing may cause elephants to become sedentary, reduce seasonal movement, prolong, and concentrate feeding impacts (Cumming et al., 1997; Lombard et al., 2001; Guldmond et al., 2008). Fencing acts as a fixed boundary, where the confinements of elephants could deprive access to seasonal habitat, in turn increasing encounters of selected tree species (O'Connor et al., 2007). The potential for elephants to utilise the same patches of vegetation increases in small, fenced reserves, because of their inability to distribute themselves effectively in response to resource availability (Slotow et al., 2005; Mackey et al., 2006; de Boer et al., 2015). Additionally, the fence line itself can prove problematic in extensive areas, as they may cause elephants to bunch up against the fence line (Loarie et al., 2009). Movement patterns of elephants have moreover been shown to be influenced by the proximity of the fence line, with studies suggesting increased habitat use within the centre of the reserve (see e.g. Vanak et al., 2010).

Elephant impact on vegetation is known to be affected by their feeding behaviours. Potential impacts of elephants on vegetation include broken branches, the main stem being broken, debarking, the tree being pushed over, and the elephant causing the death of the tree (Table 1). Recovery levels are generally determined as the ability of individual trees to survive after elephant browsing occurs. Bark recovery (Wigley et al., 2019), coppicing (Jacobs and Biggs, 2002) and sprouting (Bond and Midgley, 2001) are generally used as indicators of tree recovery. Elephants consume both woody vegetation and grasses, and they characteristically select vegetation depending on seasonal availability (Buss, 1961; Laws, 1970; Owen-Smith, 1988; de Boer et al., 2000). They typically feed on tree species with high nutrients in their leaves (Holdo, 2003; Jachmann, 1989; Wiseman et al., 2004, Novellie et al., 1991) and select trees with large volumes of foliage to gain maximum energy output; the level of impact has been suggested to depend on tree characteristics such as the trees height and canopy width (Boundja and Midgley, 2010; Levick and Asner, 2013, Howes et al., 2020; Thornley et al., 2020). Private wildlife reserves are often set on degraded livestock areas, which can force elephants to utilise woody vegetation year-round due to poor grazing conditions (O'Connor, 2007; Smallie and O'Connor, 2000). It is however important to acknowledge that conflicting views remain on elephant vegetation preferences and their nutritional characteristics (Scholes and Mennell, 2008). An increase in bark and roots being consumed may indicate nutritional stress, which may in turn result in greater impact on woody vegetation (Guy, 1976; Barnes et al., 1994).

Therefore, the ability for individual trees to recover after being impacted by elephants, is essential to maintain species within the reserve (Moe et al., 2009; Kohi et al., 2011; Scogings et al., 2012). The resilience of tree species depends on whether that species possesses recruitment and regeneration rates that match the rate of mortality over time (O'Connor et al., 2007, Thomson, 1975; Lock, 1977; Cumming, 1981). Once impact has occurred, trees can

recover through coppicing, regrowth as well as seedling regeneration, but the success of this is dependent on the level of impact to the tree. Adults of some species such as mopane (*Colophospermum mopane*) have high coppicing ability (Lewis, 1991; Ben-Shahar, 1996; Styles and Skinner, 2000), while other species have weak regrowth ability (e.g., umbrella thorn (*Vachellia tortilis*)) (MacGregor and O'Connor, 2004). Most *Vachellia* and *Senegalia* populations heavily impacted by elephants have persisted through time on account of their regeneration ability (Croze, 1974; Leuthold, 1977; Pellew, 1983; Lock, 1993; Dublin, 1995; Western and Maitumo, 2004). Studies on marula trees determined that even though impacts from elephants were high, mortality rates were low as affected trees did show signs of recovery (Gadd, 2002). Small and medium-sized herbivore species including small browsers such as impala (*Aepyceros melampus*) and rodents have been suggested to prevent seedling recruitment in marula trees, due to high utilisation and seed predation (Haig 1999; Helm et al., 2009; Helm and Witkowski, 2012). Elephants may kill regenerating stems through over-browsing, which may be exacerbated in fenced reserves (Moseby et al., 2018). There is a need to determine the recovery levels on impacted individual trees across fenced reserves with elephant presence, so reserve managers can apply appropriate measures to maintain sustainable populations of tree species while securing elephant survival. In this study, we attempt to determine levels of tree recovery following elephant induced impact whilst considering habitat use and fence line effect. While doing so, we test the following hypotheses:

(H1) Impacts will be more prevalent on tall trees ( $\geq 5$  m) within high use areas, and low levels of recovery will be displayed on these trees

(H2) Trees close to the fence line will be less impacted than trees further from the fence line, which will display lower levels of recovery

## 2.3 Method

### 2.3.1 Study site

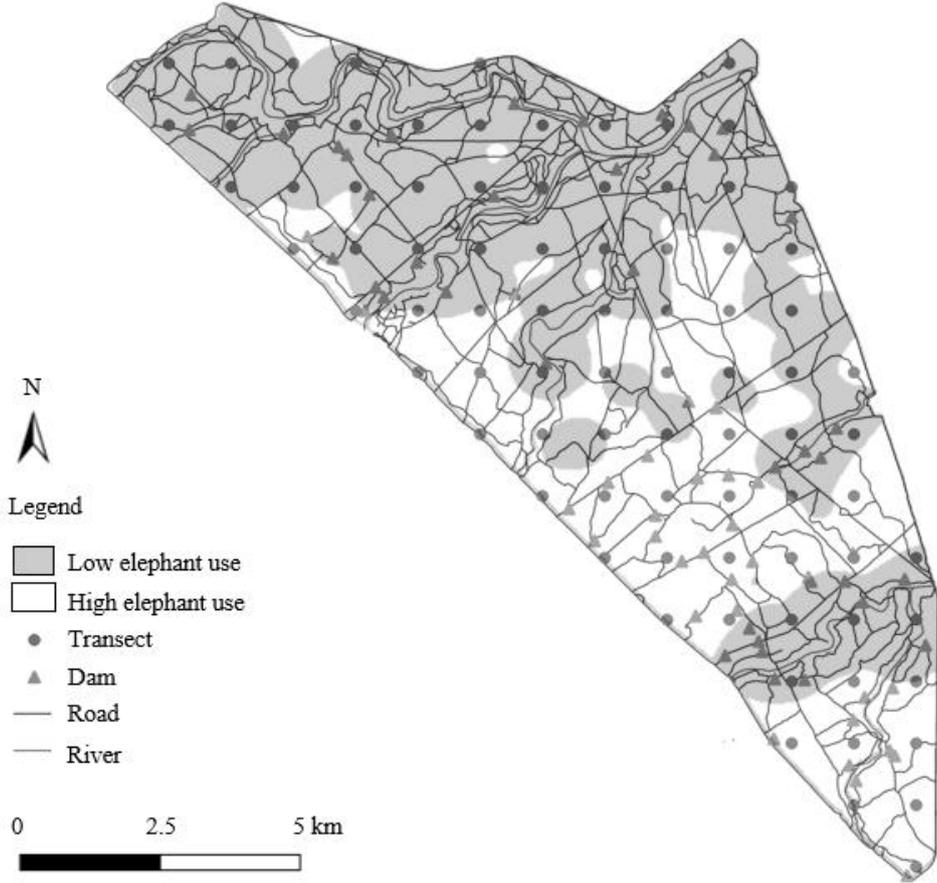
The study was carried out in Karongwe Private Game Reserve (KPGR), a 7,960-hectare fenced private reserve in the Limpopo province of South Africa (S24.227061, E30.603302). The reserve consists of two savanna vegetation types: Granite Lowveld and Tzaneen Sour Bushveld type (Mucina and Rutherford, 2006). Average daily temperatures range from 5-17 °C in winter (June to August) and 17-28 °C in summer (December to February). The altitude varies from 489 m to 520 m above sea level (Lehmann et al., 2008).

The reserve originally consisted of 10 individual private farmlands, but division fences were removed in 1998 and a Reserve was established. KPGR is bordered by public roads, which are 50 m from the fence line. The western fence line (19.1 km) runs along the paved R36 route, while gravel roads run parallel along the eastern (14.5 km) and northern (11.9 km) fence lines.

Elephants were translocated to KPGR in 1999 from Kapama Game Reserve and Magudu, Kwaza-Zulu Natal (7 individuals). Since 2011, the elephant population has consisted of one stable family unit of adult females, both male and female subadults and juveniles. There are also two bulls present on this reserve. Owen-Smith et al. (2006) suggests that an effective elephant population density is 0.28 km<sup>2</sup>, so based on this estimation, KPGR could support 22.28 elephants. KPGR currently supports 20 elephants.

Elephants within KPGR are never more than 3 km from an artificial water point: there are 70 dams across the reserve; some are pumped when the water level is low. Distance to the water was therefore not considered as a limiting factor of elephant distribution or an explanatory variable likely to shape tree damage; it was therefore not included in our models (Harris, Russell, van Aarde, and Pimm, 2008; Shannon et al., 2006) (Figure 2.1).

**Figure 2.1.** Spatial distribution of water points, vegetation transects and elephant utilisation distribution across Karongwe Private Game Reserve (KPGR)



### **2.3.2 Data collection**

Elephant locations were determined by sightings twice a day: AM drives (05:00) and PM drives (15:00), as part of a long-term study using visual recordings (data collection began in 1999). Sightings were recorded within 5 m of the observer, where the GPS recording was taken for the elephant. After locating the focal animal, the following parameters were recorded:

- Date
- Time
- Location
- Map coordinates (derived from Garmin™ GPSMap® 60CSX - GNSS)

Vegetation data were collected between July and September 2018 using 84, 10 x 100 m transects distributed across the reserve (Appendix 1: Table A.1; 4: Table A.4.1). Sampled trees near the fence line were those who were found being within 100m of the fence line. Out of the total 84 performed, 29 transects were carried out in areas considered to be near the fence line. Navigation-grade GNSS co-ordinates were acquired at the start of each transect. Every tree of height  $\geq 5$  m and diameter breast height (DBH) of  $\geq 15$  cm was sampled for elephant impact and recovery (Coetzee et al., 1979; Staub et al., 2013). Our study considered  $>5$  m trees as research to date has focussed on elephant impact on trees within this height class as they are often targeted by elephants, but these studies lacked insights on multiple tree species within a small, fenced reserves with high elephant density (Weaver, 1995; Biggs and Jacobs, 2002; Helm et al., 2009; Helm and Witkowski, 2012). When a tree met the necessary requirements, the following parameters were recorded:

- Species, height (m), DBH (cm)
- Elephant impact type (Table 2.1)

- Tree recovery type (Table 2.1)

Tree impact types were derived from the Walker damage scale (Walker, 1976) (Table 2.1). Elephant impact on trees was easily distinguished from that of other browsers due to their foraging behaviours. Elephants feed on woody vegetation by breaking off branches, toppling and bark stripping using their tusks (Coetzee, 1979; Boundja and Midgley, 2010), whereas smaller browsers are narrowly selective for new leaves, flowers and fruits at lower heights (Owen-Smith and Chafota, 2012). We also recorded levels of recovery for each tree.

**Table 2.1.** Scale used to record elephant browsing during field observations. Impacts and recovery levels on tree species were derived from Walker (1976).

<i>Variable</i>	<i>Observation</i>
<i>Impact</i>	Branches broken (A)
	Condition of the tree: Alive/Dead (B)
	Main stem broken (C)
	Main trunk debarked (D)
	Pushed over (E)
<i>Recovery</i>	Presence of coppicing (tree regeneration from stump) (F)
	Bark regrowth (G)
	Presence of sprouting (resprouting from stems) (H)

### 2.3.3 Data analysis

‘High-use’ and ‘low-use’ areas were determined using elephant location data collected in 2018. Habitat use is described as a categorical variable, with high-use areas including areas where elephants are within their home and core range and low-use areas including areas not within

their home and core range. A utilisation distribution (UD) was created to provide a measure of the probability an elephant to be found at a given location (Worton, 1989); the ‘heatmap’ tool in QGIS was then used to perform a quartic (biweighted) kernel density estimation (KDE) using a discrete data set to produce a continuous UD. To define the home range of the elephants, 95% of volume contours of the KDE was extracted to remove the outliers. 50% of the space use distribution, determined the elephants kernel core range (CR) and was extracted for this study. ‘High-use’ areas of elephants were defined as merged recordings of elephants within the home range and core range. ‘Low-use’ areas corresponded to areas where no elephant presence was recorded.

Data exploration determined that sample sizes were too small to effectively test our two hypotheses for the following impact and recovery variables: B, C, G and H (Table 2.2). Therefore, GLMMs (binomial distribution) were used to model the likelihood of a given impact type (A, D, E, Table 2.2) to be found on a particular tree as a function of the height of that tree, its distance to the fence, and whether or not the tree was located in an area highly used by elephants (all fixed effects). Transect identity was modelled as a random effect. We also used this approach to model the likelihood of coppicing (F; Table 2.2) to occur on a given tree as a function of the height of that tree, its distance to the fence, and whether or not the tree was located in an area highly used by elephants (Table 2.2). A baseline model was constructed with all the possible interactions and main effects. Akaike information criterion (AIC) (Burnham and Anderson, 2004) and model averaging was used to select a combination of the top models. We limited the calculation of the conditional averages to models within 2 delta AIC of the best model. The conditional average for each model was used for further inference. All models were built in R using the “lme4” package (Pinheiro and Bates, 2000; R Core Team, 2014).

Model assumptions were verified by plotting residuals for spatial dependency. We determined that the random effect approach is sufficient for spatial dependency by conducting a Moran’s

I test on all models (Getis, 2008). Results confirm that spatial autocorrelation is absent (p-value > 0.05) in the residuals of all models.

**Table 2.2** GLMM structure of dependent and independent variables (fixed effects) for all impact and recovery variables

<i>Dependent variable</i>	<i>Independent variables</i>
<i>Branches Broken (A)</i>	Habitat Use + Height + Habitat Use: Height + Fence line
<i>Debarking (D)</i>	Fence line + Habitat Use + Height + Fence line: Height + Fence line: Habitat Use
<i>Pushed over (E)</i>	Habitat use + Height + Fence line
<i>Coppicing (F)</i>	Habitat Use + Height + Habitat Use: Height + Fence line + Fence line: Height + Fence line: Habitat Use

## 2.4 Results

634 trees were considered for analysis; these data were gathered on the five most common species in the reserve: knobthorn (*Sengalia nigrescens*) 30%; marula 19%; velvet corkwood (*Commiphora mollis*) 13%; red bushwillow (*Combretum apiculatum*) 11%; leadwood (*Combretum imberbe*) 5%. 570 (90%) of these trees expressed visible signs of elephant impact. Overall, levels of impact across the reserve were thus high, but levels of recovery were low in both high-use and low-use habitats (Figure 2.2).

In total 10% of trees were found to be dead. 13% of all sampled trees were found to be pushed over. 58% of trees exhibited branches broken and only 2% of sampled trees had their main stem broken. 10% of trees displayed debarking. 5% of trees showed signs of coppicing, 2% sprouting and 2% regrowth.

Debarking was more likely to occur on trees found in low-use areas; similarly, trees were more likely to be pushed over in low-use areas ( $P=0.04$  and  $P=0.03$ , respectively). The likelihood of finding trees with branches broken was not influenced by elephant habitat use (all  $P > 0.05$ ; Table 2.3).

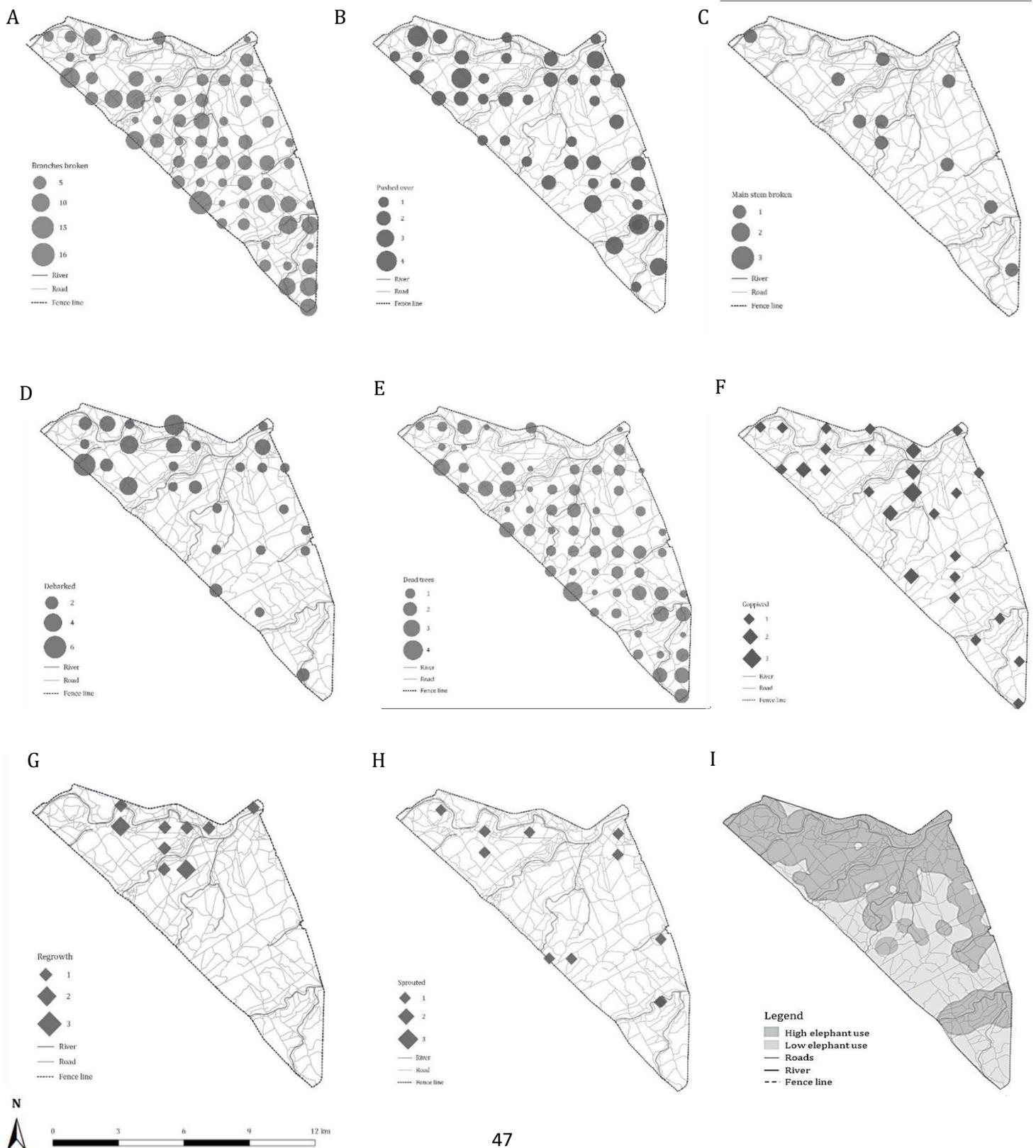
Taller trees were significantly more likely to show signs of debarking ( $P=0.01$ ) and branches being broken ( $P=0.01$ ). However, taller trees were less likely to be pushed over ( $P=0.01$ ).

Trees within proximity to the fence line did not significantly experience less impacts from elephants than trees further apart from the fence line (all  $P > 0.05$ ; Table 2.3). The level of use, distance to the fence and tree height were not significant predictors of tree recovery indicators.

**Table 2.3.** GLMM outputs of dependent and independent variables (fixed effects) for all impact and recovery variables on the best models.

<i>Variable</i>	<i>Fixed effects</i>	<i>GLMM coefficient (estimate)</i>	<i>SE</i>	<i>Z value</i>	<i>P value</i>
<i>Branches broken (A)</i>	Habitat use	0.63	0.96	0.66	0.51
	Height	0.19	0.07	2.80	<b>0.01</b>
	Habitat use: Height	0.18	0.14	1.25	0.21
	Fence line	-0.29	0.59	0.49	0.62
<i>Debarking (D)</i>	Fence line	3.17	2.46	1.29	0.19
	Habitat Use	-1.07	0.52	2.07	<b>0.04</b>
	Height	0.31	0.08	3.94	<b>0.01</b>
	Fence line: Height	-0.38	0.23	1.69	0.09
	Fence line: Habitat use	-1.54	1.49	1.03	0.30
<i>Pushed over (E)</i>	Habitat use	-0.74	0.35	2.12	<b>0.03</b>
	Height	-0.22	0.08	2.81	<b>0.01</b>
	Fence line	0.17	0.66	0.26	0.79
<i>Coppicing (F)</i>	Habitat use	2.61	2.89	0.89	0.37
	Height	-0.13	0.12	1.05	0.29
	Habitat use: Height	-0.70	0.42	1.68	0.09
	Fence line	4.80	4.59	1.05	0.29
	Fence line: Height	-0.75	0.64	1.18	0.24
	Fence line: Habitat use	-2.18	2.36	0.92	0.36

**Figure 2.2** Spatial distribution of total impacts (●) and recovery (◆) on trees: A= Branches broken; B = Pushed Over; C= Main Stem Broken; D= Debarked; E= Alive; F= Coppiced; G= Regrowth; H=Sprouted and I= Total elephant habitat use during 2018



## 2.5 Discussion

This study shows that African savanna elephant impact on trees does not occur randomly. Contrary to our expectations, (1) the level of tree impact was not determined by the proximity to the fence line; (2) tree height and habitat use impacted differently risks of debarking, branches being broken, and tree being pushed over; (3) tree recovery could not be predicted from tree height, the level of elephant use in the area occupied by the tree, or the distance from the tree to the fence line.

The results regarding the level of impact in response to tree height were consistent with impact from elephants that have been seen on other sites across African savannas, when considering for impacts related to branches being broken and debarking. Tree height has been shown to be a significant indicator of the presence of elephant impact (Makhabu et al., 2006; Mapaure and Mhlanga, 2000; Scogings et al., 2012), which has also been the focus of many studies (Biggs and Jacobs, 2002; Helm et al., 2009; Helm and Witkowski, 2012; Cook et al., 2017). Previous studies considering elephant impacts on marula trees, have shown that tall trees between 5-11 m high showed signs of impacts, and the greatest mortality was found in trees in the 5-8m height class (Biggs and Jacobs, 2002; Helm et al., 2009; Helm and Witkowski, 2012; Cook et al., 2017).

However, our findings do not match previous observations that the containment of elephants increases impacts on trees within the areas they mostly utilise (Cumming et al., 1997; Lombard et al., 2001). Additionally, we identified that the likelihood of a tree being debarked or being pushed over was reduced in high-use areas, which was surprising as we expected habitat use to correlate with impact. This mismatch between expectations and observations could be a result of how habitat use was determined in this study, as we only considered elephant sightings for one year (2018). Further work is required to establish whether the reported patterns remain consistent once more information on elephant distribution is taken into account.

We also considered the distance from the fence line in relation to tree impact, as this had not been previously studied in a small, fenced reserve. Recent research has focussed on efforts to deter elephants from fences to prevent elephants breaking out of reserves, reduce crop raiding and human-wildlife conflict (Pozo et al., 2019; Changa'a et al., 2016; Ngama et al., 2016; King et al., 2011) but little is known about the edge-effect of fences on tree damage and recovery (Vanak et al., 2010). We found no significant relationship between distance to the fence line and all impact and recovery variables. This suggests that management plans aimed at focusing efforts on reducing the impacts of elephants on trees based on their distance to the fence line may not be evidence-based.

Furthermore, we analysed the level of recovery on impacted trees, as this had been largely overlooked in previous studies which focussed on impact in small, fenced reserves considering multiple tree species. We considered tree recovery an important factor, as the ability for trees to recover after being impacted is essential for the long-term sustainability of tree populations (Leuthold, 1996). Our results showed a significant lack of recovery on most sampled trees, including large trees within high-use areas, which is a concern in an area of high elephant density and high levels of impact. Previous studies have emphasised that if tree species are unable to recover from impact, severely impacted trees will not persist through time (O'Connor et al., 2007; Moe et al., 2009; Kohi et al., 2011; Scogings et al., 2012). Even if tree species do exhibit signs of recovery, they can become sterilized if no seeds are produced (Midgley et al., 2020). Added to this, there are numerous factors that could compromise recruitment of large trees such as fire and excessive herbivory from smaller antelope whose densities have increased as a result of increased abundance of artificial water points (AWP) in private nature reserves. Therefore, including smaller trees, establishing seedling recruitment and tree survival rates is needed in future studies on elephant impact to determine if this impact to large trees unsustainable.

Current landscape conditions in parts of Africa, particularly in South Africa where fencing is more prevalent, limit space use and increase impacts in the savanna landscape (Loarie et al., 2009). We have attempted to address some of the explanatory factors likely to impact tree damage levels by elephants and tree recovery within this study, but there are limitations to our work. Our study is constrained both temporally and spatially, as we only collected data over one time period within one area. The study could be replicated in the wet and dry season as a comparison to determine how seasonality and water availability effects tree recovery. Tree impact could be recent, therefore there may not have been sufficient time to display signs of recovery that could be identified during the time of study. We only looked at trees >5 m in height and focussed on the five most common species. We then had to score impact binomially, which reduced our ability to explore how responses differ according to damage level. As demonstrated in Gadd (2002), tree species may be more vulnerable to mortality if a greater percentage of bark has been stripped. Therefore, future studies should consider scoring impact levels into quantitative formats, to discern from low level impacts and high levels impacts (Helm et al., 2009). This would aid in identifying areas where high levels of impact occur. Future research also needed to look at information over multiple years and other factors that might impact elephant movements, such as human- wildlife conflict areas and other anthropogenic disturbances that were beyond the scope of this study.

Management of elephants on small reserves such as the study site in question is challenging as there are multiple factors to consider. This is due to the size constraints of the reserve where expansion is not a viable option, which is the case in many reserves throughout South Africa with increasing elephant population numbers. Elephant bulls have been shown to cause greater impacts to vegetation compared to cows as they are larger bodied and have more destructive tendencies, especially when in musth (Greyling, 2004). The success of contraception as a management tool to control population numbers has been shown in several small reserves in South Africa, including the Greater Makalali Private Game Reserve and Tembe Elephant Park

(Bertschinger et al., 2018). Contraceptives were also administered to bull elephants in 2016 in the reserve considered in this study. Future studies should explore the impact of contraception on the behaviour of elephants in small reserves, considering any changes of behaviours and ultimately how these impacts on vegetation (Stretch et al., 2002).

Within the study site, the level of impact on trees was high but tree mortality was relatively low (10% of trees were dead). Trees exhibiting any signs of recovery was also very low (<3% for each recovery variable). If elephant impact continues at the rate that we have found on this reserve, then there is cause for concern as tree regeneration levels are low and ultimately mortality will increase. This suggests that elephant impact may be unsustainable in the long term, despite the elephant density falling within the carrying capacity estimate (Pienaar et al., 1966). Therefore, mitigation measures may be required to reduce elephant impact on tree species. Further research is needed to determine the severity of impact, regeneration and recruitment levels on individual tree species, so that mitigation measures can be put in place to reduce and manage levels of impact. Mitigation measures include wire netting, and beehive protection for individual trees as well as controlling elephant distribution in areas that are vulnerable to severe elephant impacts (Henley and Cook, 2019).

Our study has given an insight into impacts on trees by elephants and subsequent recovery, within a fenced reserve with high elephant density, but there is still a need to determine the long-term impacts of elephant on vegetation. We have stressed the need to focus mitigation efforts on trees with high levels of impact where recovery was not identified. A possible addition to this study would be to include trees of smaller height classes to see if impacts occur as readily to smaller trees and if so, how well they recover (<5 m in height). This has been suggested by anecdotal evidence, where elephants have shown an increase in preference for seedlings. We also suggest that future studies should consider the secondary effects of elephant impact, for example bark stripping makes trees more susceptible to further impacts (Campbell

et al., 1996; Wigley et al. 2019). As reserves vary in elephant density, methods of population control and types of vegetation vary from one reserve to another, it is imperative that we establish new holistic management methods for the sustainability of fenced reserves and to ultimately support long-term elephant conservation.

### **3. The effect of insects on elephant-induced tree impact within a small, fenced reserve in South Africa**

- *O2: Identify how field data collection and modelling analyses can be used to determine the secondary effects of elephant impact on trees > 2 m, and how this influences tree species recovery level*
- *Main findings: Secondary impact was identified as insects, where wood borers and termites were considered. Irrespective of tree height, termites were found to be more likely to colonise damaged trees without signs of recovery and wood borers were more likely to colonise impacted trees showing signs of recovery. Therefore, carefully considering management approaches for elephant-induced termite and wood borer impact on trees should be applied in this fenced reserve.*

#### **3.1 Abstract**

African savanna elephants (*Loxodonta africana*) have been recognised as ecosystem engineers, where their feeding habits have been shown to alter landscapes. Within small, fenced reserves, studies exploring elephant impact on trees and their recovery have overlooked secondary impacts that could be contributing to tree mortality. The aim of this study is to assess the significance of both elephant impact and secondary impact, and the subsequent tree recovery. We identified secondary impact as insects and considered wood borers and termites in this study. This was conducted in the small fenced Karongwe Private Game Reserve (KPGR), South Africa. We analysed the level of damage, recovery and insect presence using vegetation transects, where all trees  $\geq 2$  m in height were surveyed (n=1278 trees). Forty tree species were recorded, with 5 species accounting for 77% of the dataset and used for further analysis.

Termites were found to be more likely to colonise impacted trees without signs of recovery. However, wood borers were more likely to colonise impacted trees showing signs of recovery. Termites and wood borer presence on impacted trees was not dependent on tree height. We suggest carefully considering management approaches for elephant-induced termite and wood borer impact on trees.

### ***Keywords***

Karongwe Game Reserve, *Loxodonta africana*, Tree impact, Tree recovery, Termite impact, Wood borer impact, Elephant density

## **3.2 Introduction**

African savanna elephants (*Loxodonta africana*) are known to be capable of altering landscapes by changing species composition and reducing plant biomass (White and Goodman, 2009). Their ability to structurally modify vegetation has resulted in several studies on savanna trees, the impacts elephants can inflict to them (Ben-Shahar, 1998; Gandiwa et al., 2011; Asner et al., 2016), and the subsequent impact this may have on other tree species (Kerley and Landman, 2006; Hrabar and Du Toit, 2014; Joseph et al., 2018). The direct impact elephants have on vegetation is a particular concern in areas of high elephant density (Bounja and Midgley, 2009), such as fenced reserves, where increased impact can decrease savanna vegetation heterogeneity (Pringle, 2008). To date, literature has largely focussed on elephant impact with regards to the abundance of large trees, due to their significant environmental and economic value (Shannon et al., 2008). Types of impacts caused by elephants include breaking branches and stems (Nasseri, McBrayer and Schulte, 2011), as well as debarking (Calenge et al., 2002). Debarking has been shown to increase tree susceptibility to other associated impacts, such as disease infection (for example, heart rot of the stem and branches) (Shannon et al., 2011; White and Goodman, 2009; Helm et al., 2011). Tree impact can also enhance tree susceptibility to

termites (*Coptotermes* species), woodborers (*Cerambycidae* species) or other insects, which may shorten trees' life span (Hatcher, 1995).

Termites can indeed penetrate trees through fractures in the bark (Gould et al., 1993; N'Dri et al., 2011) and establish secondary nests inside the tree's cavities (Harris, 1968), increasing tree susceptibility to insect colonisations. This happens more frequently when bark is removed by animals such as elephants or porcupines (*Hystrix africaeaustralis*) (Helm et al., 2011). Termites seem to prefer older, stressed trees with low water content and gaps within the bark (Cowie et al., 1989; Gould et al., 1993; Werner et al., 2008). Within the Kruger National Park (South Africa), termites have been shown to prevent regrowth of marula trees (*Sclerocarya birrea*) (Coetzee et al., 1979) and affect tree survival (Cook and Henley, 2019). Additionally, termite presence can result in trees being hollowed out from the inside, which may exacerbate vulnerability to elephant's and other animals' impact (Werner et al., 2008).

Wood borer impact to trees is characterised by the boring activity of larvae and adult beetles in the stems and branches of impacted or stressed hosts (Halperin and Geis, 1999; Peters et al., 2002; Nair, 2007; Liu et al., 2008). Adult mortality of marula trees has been suggested to be attributed to rapid invasion by wood borers after bark stripping occurs and the sapwood is exposed (Helm et al., 2011; Coetzee et al. 1979; Guy 1989; Jacobs and Biggs, 2002). Structural impact caused by elephants and other herbivores, enables wood borers to substantially weaken the stem (Coetzee et al., 1979), exacerbating the original impact by reducing the trees' ability to recover (Guy, 1989). Recently, Vogel et al. (2014) found that the presence of insects on large, older trees with high elephant impact was negatively related to knob thorn trees' (*Sengalia nigrescens*) survival, indicating elephant impact could indeed be indirectly facilitating insect attack and shortening the trees lifespan.

The interactive effects of multiple disturbances on different woody savanna species, such as the subsequent impact of insects on trees impacted by megaherbivores, are currently not completely understood (Holdo, 2007; Migley et al., 2010). To date, studies have considered the importance of elephant impact on supporting arboreal species populations (Pringle et al., 2015) and the functional relationship between ant abundance and the benefits of protecting trees from elephant's herbivory (Palmer et al., 2013). Increasing elephant densities in fenced reserves (Skarpe et al, 2004; Shannon et al., 2008) and the decreasing numbers of large trees (Ben-Shahar 1993; Eckhardt et al., 2000; Moncrieff et al., 2008), have resulted in a need for improved knowledge of how elephants and other subsequent disturbances can result in tree death (Holdo, 2005). Tree species are faced with several biotic constraints such as insects (Wargo, 1996; Hakeem et al., 2012), and this secondary impact partnered with the lack of tree recovery, has been overlooked in many studies focussed on elephant damage. This paper aims to address this knowledge gap, by investigating how insect presence relates to tree impact and affects tree recovery. While doing so, this contribution will test the following hypotheses:

- 1) Insects are more likely to colonise trees damaged by elephants than trees not damaged by elephants.
- 2) Insects are more likely to colonise taller trees damaged by elephants than shorter trees.
- 3) Insects are less likely to colonise damaged trees that exhibit signs of recovery

### **3.3 Method**

#### ***3.3.1 Study site***

The study was carried out in Karongwe Private Game Reserve (KPGR), a 7,960-hectare fenced private reserve in the Limpopo province of South Africa (centred on 30.60° E 24.23° N). The reserve consists of two savanna vegetation types: Granite Lowveld and Tzaneen Sour Bushveld (Mucina and Rutherford, 2006). Daily mean ambient air temperatures range from 5-17° C in

winter (June to August) to 17-28° C in summer (December to February). The elevation range is 489-520 m above mean sea level (Lehmann et al., 2008).

The reserve originally consisted of 10 individual private farmlands, but division fences were removed in 1998 and a Reserve was established. KPGR is bordered by public roads, which are as close as 50m from the fence line. The western fence line (19.1 km) runs along the paved R36 route, while gravel roads run parallel along the eastern (14.5 km) and northern (11.9 km) fence lines.

Elephants (7 individuals) were translocated to KPGR in 1999 from Kapama Game Reserve and Maggudu, Kwaza-Zulu Natal. Since 2011 the elephant population has consisted of one stable family unit of adult females, both male and female subadults and juveniles. There are also two bulls present on this reserve. Owen-Smith et al. (2006) suggest that an effective elephant population density is 0.28/km<sup>2</sup>, based upon which KPGR could support 22.28 elephants. KPGR currently supports 20 elephants.

### ***3.3.2 Data collection***

Vegetation data were collected in June – October 2019, with 84, 10 x 100m transects to represent the vegetation type across the reserve (Appendix 1: Table A.1; 4: Table A.4.2). A navigation-grade GNSS (Garmin™ GPSMap® 60CSX) was used to acquire co-ordinate pairs at the start of each transect. Every tree of height  $\geq 2$  m and diameter breast height (DBH) of  $\geq 10$  cm was sampled for elephant damage and recovery (Coetzee et al., 1979; Staub et al., 2013).

When a tree met the necessary requirements, the following parameters were recorded:

- Species, height (m), DBH (cm)
- Elephant impact type (Table 3.1)
- Tree recovery type (Table 3.1)
- Insect presence: Termites, Wood borers

Tree impacts were derived from the Walker damage scale (Walker, 1976) (Table 3.1) to determine the types of elephant impacts on each tree during data collection. Elephant impacts on trees is easily distinguished from that of other browsers due to their foraging behaviours (Jachmann and Bell, 1985). Impacts were recorded according to the methods used by Jacobs and Biggs (2002). The types of recovery were also recorded for each tree based on the types of recovery identified by Gadd (2002) (Table 3.1). All trees sampled were recorded using a binary scoring system for each parameter.

**Table 3.1** Scale used to record elephant browsing impacts and recovery levels on tree species after Walker (1976).

<b>Variable</b>	<b>Observation</b>
<i>Damage</i>	Branches broken Condition of the tree: Alive/Dead Main stem broken Main trunk debarked Pushed over
<i>Recovery</i>	Presence of coppicing Bark regrowth Presence of sprouting

### 3.3.3 Data analysis

A total of 1278 trees were used in our analysis, on the five most abundant tree species: knobthorn (*Senegalia nigrescens*), marula (*Sclerocarya birrea*), velvet corkwood (*Commiphora mollis*), red bush willow (*Combretum apiculatum*), and leadwood (*Combretum imberbe*).

Data exploration was carried out following the protocol described in Zuur et al. (2010). Generalized linear mixed effect models (GLMM; binomial distribution) were used to model the likelihood of a given insect type to be found on any given tree, as a function of the height of the tree, the level of tree impacts, and the two-way interactions between covariates (all fixed effects). We also used GLMM to model the likelihood of a given insect type to be found on any impacted tree, as a function of the height of the tree, whether the impacted tree showed any signs of recovery, and the two-way interactions between covariates (all fixed effects). In all cases, transect identity was modelled as a random effect. Identified impact and recovery types were classified into ‘ImpactScore’ (0-5, depending on the number of types of impacts identified) and ‘RecoveryScore’ (0-3, depending on the number of types of recovery identified) for each observed tree (Table 3.1). We tested the model structure by adding the independent variables and then tested the main effects plus the interactions; the best models were used for further inference. All models were built in R using the “lme4” package (Pinheiro and Bates, 2000; R Core Team, 2014).

Model assumptions were verified by plotting residuals for spatial dependency. We determined that the random effect approach was sufficient for spatial dependency by conducting Moran’s I test on all models (Getis, 2008). Results confirm that spatial autocorrelation is absent in the residuals of all models, and inference was taken from the best performing models (Moran’s I for all models  $< 0.02$ ; p-value  $< 0.05$  for all models).

### **3.4 Results**

Termites were more likely to be found on more impacted trees, ( $P=0.02$ ). Tree height was less important as the impact score increased ( $P=0.01$ ).

The likelihood of finding wood borers on trees, on the other hand, was not impacted by tree height or impact score (all  $P > 0.05$ ).

We also found that recovery was significantly linked to wood borers presence: the higher the recovery score, the greater the chance of finding woodborers on impacted trees ( $P = 0.03$ ). However, the likelihood of finding termites on impacted trees was not related to the tree's recovery score ( $P > 0.05$ ). Both termite and wood borer presence were not affected by tree height of impacted trees ( $P > 0.05$ ) (Table 3.2).

36% of sampled trees had no termite or wood borer presence. Trees that had termite presence had the highest level of impacts (60%) compared to wood borers (50%) and trees without insect presence (37%). Trees that had termite presence also had the highest level of recovery (31%). Trees with wood borer presence had similar levels of recovery compared to trees without insect presence (19% and 17%, respectively). The majority of impacts that were found on trees with and without insect presence were branches broken. Trees with termite presence had the lowest number of trees pushed over compared to trees with wood borer presence and trees without insects (15, 36 and 31, respectively).

**Table 3.2** GLMM outputs of dependent (termites and wood borers) and independent variables (fixed effects) for both insects tested.

Variable	Fixed Effects	GLM coefficient (estimate)	SE	Z value	P value
Wood borer	Height +	-0.04	0.05	-0.66	0.49
	RecoveryScore	0.35	0.17	2.16	<b>0.03</b>
Termite	Height	-0.13	0.11	-1.21	0.23
	RecoveryScore	-0.68	0.76	-0.89	0.37
	Height:	0.09	0.10	0.86	0.39
	RecoveryScore				
Wood borer	Height	0.08	0.06	1.39	0.10
	ImpactScore	0.64	0.39	1.63	0.10
	Height:	-0.07	0.06	-1.25	0.21
	ImpactScore				
Termite	Height	0.09	0.07	1.29	0.19
	ImpactScore	1.14	0.47	2.41	<b>0.02</b>
	Height:	-0.17	0.07	-2.51	<b>0.01</b>
	ImpactScore				

### 3.5 Discussion

Our results show that (i) termites are more likely to colonise impacted trees; (ii) wood borers are more likely to colonise impacted trees that are showing signs of recovery, and that (iii)

insect presence on impacted trees did not depend on tree height.

Studies to date that have considered elephant impact on trees have overlooked the impact of insects. We show here that termites colonising damaged trees should be regarded as a significant contributor to secondary impacts. We also found that the effect of tree height was not related to insect presence, suggesting that insects do not colonise tall trees. This raises concerns, as management efforts to date on trees have focused on large trees as they are considered the most susceptible to elephant impact (Biggs and Jacobs, 2002; Helm et al., 2009; Helm and Witkowski, 2012; Cook et al., 2017). Howes et al. (2020) determined that tree impact by elephants in an enclosed reserve was non-lethal to trees, and that taller trees were less likely to suffer from elephant impact. However, once elephant impact on trees has occurred, it was not known how the level of insect damage impacts tree mortality. We have shown here that termite presence on impacted trees could be leading to increased mortality if the tree is unable to recover from elephant impact, as they were more likely to be found on impacted trees that did not show any signs of recovery.

It could also be conceivable that debarking may not be the main driver for subsequent insect impact. Debarking has been shown to increase tree susceptibility to other impacts including fires (Ihwagi et al., 2010) and diseases, and has shown to be attributed to cause direct mortality (White and Goodman, 2009). This method of elephant feeding has also been shown to make the tree more susceptible to termites, wood borers and other insect activity and ultimately, shortening the trees lifespan (Hatcher, 1995). However, our results show that termites are more likely to colonize trees that have a greater number of impacts, which could result in further impacts and increase likelihood of tree mortality. Added to this, our results showed that wood borer presence was likely to be found on trees that exhibited signs of recovery, suggesting that even trees showing signs of recovery may encounter further impacts from insects. We suggest mitigation methods should focus on the most susceptible trees that show high levels of impact,

that are likely to incur further impacts from termites and consider wood borers presence on trees with recovery.

We stress that our study does not indicate the removal and prevention of insects at this stage. We are suggesting that more focus should be directed towards secondary impacts, as opposed to just considering elephant impact on its own in relation to tree mortality. Additionally, elephant feeding habits result in greater availability of food for termites and woodborers (Holdo and McDowell, 2004). In fact, within fenced reserves that limit wildlife migration between reserves, high densities of termite mounds (*Macrotermes*) are important to conservation as they are able to sustain wildlife population by sustaining nutritious forage availability across seasons. Trees species within proximity to termite mounds within savanna environments benefit as termite mounds can aid tree growth due their increased soil fertility as well as water availability (Davies et al., 2016). This can be crucial during the dry season when forage is limited (Davies et al., 2016). Studies have also suggested that within fenced reserves, elephants show feeding preferences to foraging over grazing, which only enhances the need for termite mounds (Jouquet et al., 2011). There have been studies on the relationship between termite mounds and herbivore feeding patterns (Muvengwi et al., 2013; Okullo et al., 2012; Van der Plas et al., 2013). A recent study focussed on the influence of mound size on vegetation diversity, finding that elephant damage to mound associated vegetation reduced the microclimate effects provided by vegetation (Joseph et al., 2013). However, termite mounds have been shown to provide both refugia and high-quality forage for a range of herbivore species (Fleming and Loveridge, 2003; Grant and Scholes, 2006; Mobæk et al., 2005). Therefore, we acknowledge that termite mounds are essential for biodiversity and providing ecosystem services and further research on the relationship between elephant impact and *Macrotermes* (mound-forming termites) is needed.

Further studies are therefore needed to determine the level of mitigation required to suppress

elephant induced impacts and the subsequent impacts of insects on susceptible trees. This way, we will be able to determine the best approach to maintain sufficient vegetation for elephant feeding. Some tree species are not impacted highly by feeding and can persist through time, however further impacts by insects can cause an increased severity of impacts and limit the ability of a tree to recover. Conservation efforts should focus on trees that are unable to recover and are susceptible to elephant and insect impact.

We attempted to address some of the explanatory factors of elephant impact and the impact of insects, however there are limitations to this study. We only considered termite and wood borer presence in our analysis. Our study is constrained both temporally and spatially, as we only collected data over 3 months in one area. If tree impact was recent, then there may have not been enough time to show levels of recovery or subsequent insect activity. We recommend that studies using our approach consider a larger temporal range as well as considering other insect species. It may also be informative to further studies to identify the density and size of termite mounds and their spatial distribution, associated with impacted trees in a fenced reserve, as termite mounds themselves play a part in the spatial heterogeneity of the landscape.

Our study has provided fresh insight regarding insect presence on trees impacted by a high density of elephants within a fenced reserve, and their subsequent recovery. There remains a need to determine how insects directly impact further impacts to trees, and their subsequent recovery, in the longer-term. Therefore, we suggest carefully considering management approaches for elephant-induced insect impact on trees. We acknowledge the importance of insects for many ecological processes and other species that live within fenced reserves. We have shown that even impacted trees that exhibit recovery have wood borer presence, which is a concern when considering the level of damage in a fenced reserve. As reserves vary in elephant density and the spatial distribution of termite mounds and therefore termite induced impacts, it is essential that we determine how to manage the secondary effects of elephant

impact so that vegetation can be maintained.

#### **4. The potential of photogrammetric point clouds derived from conventional aerial survey for estimating tree heights**

This chapter has been redacted. The document and/or data contains information about research in progress where there is an intention to publish later.

See <https://eprints.bournemouth.ac.uk/37449/>

## **5. African and English school children's perception of elephants and elephant conservation**

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See <https://eprints.bournemouth.ac.uk/37451/>

## **6 General discussion and conclusions**

### **6.1 Overview of thesis**

Understanding the changes in vegetation structure within landscapes that elephants utilise is important for informing conservation and management practices, to support elephant conservation (Greyling, 2004). For conservationists, advances in field-based methods and remote sensing techniques have been applied to elephant impact studies in savanna ecosystems to increase understanding of such changes (Guldemon et al., 2017; Nkosi et al., 2019).

The focus of this research was:

- To determine elephant impact on tree species of > 5 m in height within a high elephant density fenced environment, through quantifying the level of impact and recovery on various tree species (Chapter 2)
- To then determine the secondary impacts on trees > 2 m in height, through quantifying the level of elephant impact, secondary impact and recovery on various tree species (Chapter 3)
- To understand the value of remote sensing techniques to complement field data collection by testing an alternative method estimating vegetation height at a reserve extent (Chapter 4).
- To explore school children's perceptions of elephant conservation in areas where elephants were both present and absent, to identify factors that would improve and foster perceptions of elephants and ultimately their conservation. (Chapter 5).

The African savanna elephant provided a strong model species for analysing impact to vegetation, given their capacity as ecosystem engineers (Western, 1989). In addition, the Karongwe Private Game Reserve (KPGR), South Africa, provided an appropriate fenced savanna ecosystem in which to examine elephant impact, given its highly monitored elephant population and high elephant density. The sites that were selected to determine school children's perceptions of elephants and elephant conservation, provided key comparisons of areas where elephants were native and non-native, in areas that were accessible.

## **6.2 Field data collection on elephant impact and recovery: limitations and implications**

The level of elephant impact to trees > 5 m was assessed using field transects for data collection and twice daily sightings of elephants to delineate habitat use within a fixed, fenced environment (Chapter 2). Even though detecting elephant impact with this method is temporally limited, metrics determined here gave insights into the level of impact on the following five of the most common tree species within the reserve: knobthorn [*Sengalia nigrescens*]; marula [*Sclerocarya birrea*]; velvet corkwood [*Commiphora mollis*]; red bushwillow [*Combretum apiculatum*]; leadwood [*Combretum imberbe*]). Determining the impact to various tree species was key, which complemented the growing body of literature quantifying the amount of elephant impacts across different environments (Boundja and Midgley, 2010; Holdo, 2003; Levick and Asner, 2013; Morrison, Holdo and Anderson, 2016) including studies that were specific in focus on single tree species (Gadd, 1997; Weaver, 1995; Jacobs and Biggs, 2002).

Additionally, determining the level of tree recovery, quantifying high-use and low-use of elephant areas and the effect of the fence line were also considered. Few studies have addressed the levels

of recovery across multiple tree species with elephant impacts, and those that have identified this have not considered habitat use and fence line as important factors (Croze, 1974; Leuthold, 1977; Pellew, 1983; Lock, 1993; Dublin, 1995; Western and Maitumo, 2004). Understanding the influence of these factors is integral, as determining areas that elephants utilise within closed ecosystems will determine suitable methods of management for vegetation sustainment. Our study highlights the lack of recovery of some species in high-use areas, but no significant effect on fence line.

Within savanna ecosystems, where seasonality affects growth rates of vegetation, and many other biotic and abiotic factors affect recovery rates of trees after elephant impact, the limited temporal capacity of this study may be insufficient to determine the severity of elephant impact. Repeating the methods explored in Chapter 2 could resolve this, over consecutive years where vegetation impact and recovery could be determined on a larger temporal scale. However, the efficacy of this method would diminish in situations considering for tree impact and recovery alone. This could be improved by including other environmental factors, such as rainfall and temperature, within the statistical models in addition to those that were determined in Chapters 2 and 3. Including additional fixed effects within these models will increase the complexities of these and the subsequent inference which will require model selection methods to be employed (AIC; see 6.6.2). This would ultimately give a more comprehensive approach.

Given that a large proportion of literature focusses on large trees due to their economic importance, cultural significance, and environmental benefits to biodiversity, we considered large tree height (> 5 m) as an essential factor of elephant impact. Large trees are vital for ecosystem functioning and are important for the cycling of nutrients (Ludwig, De Kroon and Prins, 2008), provide a

forage source for fauna (Kerley and Landman, 2006) as well as nesting sites for birds of prey (vultures and raptors; Vogel et al., 2014). However, it may be important to determine the effects on a range of tree heights, as Asner et al. (2015) stated that elephant density plays the greatest role in determining losses of trees > 2 m in height. This may be especially important within enclosed reserves, where forage capacity and mobility are limited due to the nature of these areas, and impacts may be seen with lower height classes of vegetation if there is sufficient foliage available. Therefore, determining impacts and recovery levels of different tree heights could complement the study in Chapter 2.

The level at which a tree can recover after elephant impact is dependent on many factors, including the regeneration time of a species, the level and therefore severity at which the impact occurs as well as other biotic constraints (Lewis, 1991; Ben-Shahar, 1996; Styles and Skinner, 2000). Most studies that have considered elephant impact on trees and any levels of recovery, fail to also address any other factors that may influence the survivability of trees. This includes insect colonisation once elephant impact occurs, which may be responsible for tree deaths that are commonly associated with elephant impact independently. The results from Chapter 3 gave new insights into insect colonisation on impacted trees, both with and without signs of recovery, within a high elephant density reserve. Findings here support existing literature that termites are likely to colonise impacted trees (Hatcher, 1995; Gould et al., 1993; N'Dri et al., 2011), however results clearly demonstrated that woodborers are likely to colonise impacted trees that showed signs of recovery. The strong relationship between impacted trees showing signs of recovery and wood borer colonisation is concerning and yet to be fully understood. Moreover, in Chapter 3, trees of varying heights (> 2 m) did not show a significant relationship with colonisations of insects,

suggesting that insects do not colonise tall trees. As this study was temporally limiting, longer term impacts to vegetation because of insect colonisation still need to be quantified. Additionally, this study could be enhanced by spatially locating termite mounds to determine the direct association between termite mounds and insect colonisation to elephant impacted trees. Given that two species (termites and wood borer) were observed, the results of this project could be complemented with the analysis of other insect species, to assess the extent of secondary impacts.

Within South Africa, the impact of elephants on large trees is viewed as unsustainable within fenced reserves (Asner et al., 2016), where efforts are focussed on protecting large trees and maintain an 'ideal' state, which remains a contentious issue among conservationists. At present, the management strategy of South African National Parks focuses resource use by elephants with regards to their distribution, rather than relying entirely on elephant densities within an area (Ferreira et al., 2017; SANParks, 2012). Historically, mitigation strategies directly reduced elephant number in response to their populations. This method, however, of maintaining elephant populations did not significantly influence the loss of large trees (> 5 m), which is inherently complicated due to their survival rates changing through different life stages. Managing the effects that elephants have on large trees has been targeted at a large-scale landscape manipulation, as well as small-scale individual tree protection methods which we identified in Chapter 2. It was suggested that in the reserve considered in Chapter 2, 3 and 4 (KPGR), methods should be utilised to protect large trees directly from elephant impact, such as creating enclosure areas (Kerley and Landman, 2006). Direct protection of trees could also include wire-net protection (Derham et al. 2016), beehives (Cook et al., 2018), and rocks (SANParks, 2012) to enable selected tree species

and size classes to recover. Even with these direct protective methods to deter elephants, secondary impacts could still influence the level of recovery (Chapter 3), which needs to be further quantified.

### **6.3 Remote sensing methods: implications and limitations**

Throughout the literature, elephant impact to vegetation has focussed on field data collection methods, often using transects as a method of sampling areas within a larger area of interest. These methods are important in quantifying levels of impact and tree recovery at fine scales, determining specific elephant associated impacts as well as recovery levels that can be applied to statistical models, which we have identified in Chapter 2 and 3.

The importance of field data collection to determine elephant impact still holds intrinsic value in research today (Ben Shahr, 1996; Shannon et al., 2011). Methods adapted from Cotezee et al. (1979) and Staub et al. (2013) highlight the importance of identifying tree characteristics including number of branches broken, whether the main stem has been broken etc. as determinants of elephant impact. The level of impact can in turn influence the recovery levels of tree species if impact is extensive and tree species don't have sufficient time to recover due to frequent utilisation in high elephant density areas (see Chapter 2). Determining tree characteristics is integral for ecological studies and wildlife habitat management, however field studies determining levels of impact through transect methods can be time consuming and not cost effective, especially within large spatial extents. This also holds true for small reserves that may have limited resources hindering effective, reliable, and repeatable field data collection. Consequently, the advancement of remote sensing methods applicable for detecting tree structure can enhance methods of understanding elephant impact, which have been identified in Chapter 4.

Concurrently, elephant densities need within an area need to be determined which have to date mainly been achieved through to aerial surveys. More recently, research has applied convolutional neural networks (CNNs) to high-resolution satellite imagery (derived from WorldView-3 and 4) (Duporge et al., 2020) and high-resolution aerial images (derived from UAVs) (Delplanque et al., 2021) to detect African elephants. Remote sensing techniques can be limited due to the skills and knowledge required for efficient analysis is high-level (Nkosi et al., 2019). Such data intensive methods necessitate suitable equipment for data acquisition and inference.

As with many field data collection methods, remote sensing studies to determine elephant impact within South Africa have also been concentrated within the Kruger National Park (KNP). Munyati and Sinthumule, (2016) used panchromatic aerial photographs and SPOT imagery, while Asner et al. (2009) and Levick (2009) used ALS to determine the large-scale impact of herbivory on the structure of vegetation. Determining the rate of treefall has been the focus for these analyses, where a subsequent algorithm intensive approach (Random Forest) was required to determine treefall rates over a large landscape scale (Asner et al., 2016). Asner et al. (2016) used ALS data captured over six years, biannually, to determine treefall at a landscape scale. Although ALS provides fine spatial resolution on tree metrics (Lim et al., 2003), this approach is one of the most expensive forms of remotely sensing, and therefore only small spatial extents are acquired. Therefore, the application of using ALS consistently across spatial and temporal extents is presently not a cost-effective approach.

Chapter 4 explored the different remote sensing approaches that had been used to determine elephant impact. ALS dominates the literature regarding tree structure detection in savanna ecosystems (Asner et al., 2009; Johansen et al., 2010; Levick et al., 2010; Levick and Rogers,

2008; Levick and Asner, 2013; Davies, Gaylard and Asner, 2018). An alternative approach was investigated here, using a point cloud derived from photogrammetry using conventional aerial survey as a novel method for estimating tree height at a reserve extent in savanna ecosystems. Results demonstrated that this method was effective. Tree metrics derived from this method can support monitoring of trees in response to elephant impact. The remote sensing approach applied here considered trees > 2 m high over a reserve extent, which gave key insights into the quantification of individual trees.

#### **6.4 Children's perception of elephant conservation**

This study has shown the importance of determining direct and indirect elephant impact, recovery and tree height to aid elephant conservation by maintaining areas with suitable elephant numbers and adequate vegetation. However, this study also considered how perceptions towards elephants and elephant conservation can be improved to enhance understanding of conservation approaches. This was deemed appropriate for this study as environmental education is a key strategy that can aid scientific research and encourage people to take informed action (Monroe and Karnsy, 2015). School children were chosen as the study demographic because there has been little focus on determining how they perceive elephants and elephant conservation, both in situ and ex situ (Makecha and Ghosal, 2017). School children are also not likely to have fully formed opinions or attitudes about the environment (Pelletier et al., 2004) and it is key to influence positive environmental attitudes at an early age as they are likely to remain once formed (Asunta, 2003).

Surveys were used across different schools that were accessible at the time of study. This was limiting (see Chapter 5) as only three schools were sampled to determine different levels of exposure based on their geographic location. Even with this limitation, this study provided key

insights into how children perceived elephants at different levels of exposure. School children feared elephants where elephants were native, and students who feared elephants where elephants were not native were more likely to want to see them in the wild. This study also showed that there was a distinct misunderstanding of how elephants benefit other species as well as their feeding behaviours. The perception that elephants were in decline due to too many species in one place, was prevalent throughout school children in South Africa. It was also determined that a wildlife guide as a career choice was not valued as highly compared to other career choices.

With these new insights in relation to location, more research is required to improve school children's perceptions of elephant conservation. The next step would be to implement workshops at different levels of exposure to improve understanding around concepts that were identified here. This would require a pre- and post-workshop evaluation to determine if perceptions changed, which would be enhanced if this could be carried out repeatedly over time. With the recent changes in online learning due to the COVID-19 pandemic, both surveys and workshops could be conducted online at a large spatial area. However, using online methods will hinder data collection in countries where there is limited access to computers, which was experienced first-hand in this study. Nevertheless, improving perceptions of elephants and elephant conservation is key within this demographic as children can act as message multipliers, transferring their knowledge to their communities (Domroese and Sterling, 1999). Our study considered secondary school children as our demographic, but a range of age groups could be tested to understand when perceptions are determined.

## **6.5 Conclusions**

In summary, this thesis has provided insights into how savanna elephants impact occurs within a fenced reserve, and how this influences the level of tree species recovery in high-use and low use habitats. Furthermore, this study considered how secondary impacts to vegetation as a result of elephant impact could be reducing the trees' ability to recover, which has rarely been comprehensively studied when considering for elephant impact. This highlights the importance of considering both recovery and secondary impacts in maintaining suitable habitat to support elephants and elephant conservation in enclosed ecosystems. Results also revealed that a complementary remote sensing method can be applied to effectively determine tree height estimates, imperative for quantitative research on elephant impact. Additionally, the perception that school children have at different levels of exposure needs to be improved to support effective elephant and other wildlife conservation. Future avenues of research, detailed above, could be used to improve best practice monitoring of elephants, and expand current knowledge and understanding of the impacts that elephants have on vegetation, and the implications of these.

## **6.6 Future research**

### **6.6.1 Ecological considerations**

This thesis has given key insights into direct and indirect effects of elephants on vegetation (both impact and recovery), which is especially important to identify when considering for constrained areas. These also need to be quantified across different spatial extents, not only considering for impact and recovery, but also in response to additional abiotic and biotic factors. These factors should be considered in areas across different regions of Africa to achieve a broader understanding on the influences of multiple factors on vegetation.

A major threat to wildlife conservation is climate change, which is increasingly being driven by human activities. Rising temperatures are being seen throughout sub-Saharan Africa, which poses problems for both the vegetation and animals alike, that are found in areas of high temperatures. Extreme temperatures effect animals in different ways, but some megafauna species cannot dissipate heat easily which makes them more vulnerable to such changes. Moreover, increasing temperatures means longer drought periods, which will affect many megafauna species that are reliant on large volumes of water and sufficient vegetation for survival.

Therefore, understanding how vegetation and megafauna species react and are reacting to climate changes is important for future conservation research. Artificial water points (AWPs) have been used in many areas to support wildlife, but more research is needed on how these can effectively be safeguarded in areas with adequate vegetation to support megafauna. This can only be achieved in areas with sufficient vegetation, which is becoming increasingly fragmented. Therefore, more research is required on suitable areas for megafauna species to ensure there are sufficient resources for survival.

Future wildlife species conservation can only be achieved if we know where wildlife is located and their associated densities. This is especially challenging to determine across large spatial extents with free roaming wildlife. However, recent technological advances in artificial intelligence have shown potential in identifying megafauna from high resolution and satellite and aerial images. Future research should be targeted at applying these approaches to not only identify megafauna species, but also accurately mapping vegetation in areas where species are present. This will also give key insights into determining suitable areas for wildlife.

Another threat that has increased in recent years is Human Wildlife Conflict, which is exacerbated with land fragmentation. Future ecological research on megafauna needs to consider how people respond to wildlife and improve measures to protect wildlife species. Local communities are especially important, and increased awareness is needed on the ecological problems species face and how this can be mitigated.

### **6.6.2 Data analyses**

Some prospective methods were discussed to improve our research earlier in this chapter. Throughout this project, different types of analyses were considered and tested. Future studies that consider other data types could be applied for different levels of impact and recovery (e.g., a percentage score of debarking, the number of broken trees, the number of coppiced branched etc.). Further detail on future studies has been explained in the following section.

#### ***6.6.2.2 Impact and recovery classifications***

Chapter 3 aimed to establish the impact and recovery levels of trees species after elephant impact occurs. Impact and recovery levels were recorded in the field as direct observations, and a subsequent classification was applied as a ‘ImpactScore’ (0-5, depending on the number of types of impacts identified) and ‘RecoveryScore’ (0-3, depending on the number of types of recovery identified) for each observed tree. Initially, adding together all impact and recovery variables was considered, but the variables were correlated and therefore not truly independent (a type of impact may be more beneficial to termite than another). The significance may also change depending on whether termites or wood borers were considered. Before the classification approach was decided, a principal component analysis (PCA) followed by a discriminant function analysis (DFA) was

considered, to see whether the variability could be reduced. PCA is used to obtain a smaller number of summary variables from a larger number of candidate independent variables. PCA was tested but was unsuitable for this study. However, these methods were explored as they could be applied to other data studies on elephant impacts.

Principal component analysis (PCA) was considered as a dimensionality reduction method. PCA is an effective method to reduce the dimensionality of higher order systems and capture most of the covariance on preferable a 2 dimensional or 1 dimensional field. The strategy is similar to the development of generalised linear models, to perform regression analysis for data belonging to the exponential family. It has been argued that PCA assumptions are not appropriate for binary, or count data and generalisations exist. Multiple Correspondence Analysis (MCA) (also known as a homogeneity analysis) could be applied to binary data (Josse and Husson, 2012; Abdi and Valentin, 2007).

### ***6.6.2.3 Modelling approaches***

Both data Chapters 2 and 3 used statistical modelling to predict outcomes based on various variables that were collected in the field and adapted accordingly for hypothesis testing. Generalised Linear Mixed Models (GLMM) were selected as the most appropriate for the data collected (categorical: binomial), however initial model investigations determined that Generalised Linear Models (GLM), could also be suitable. GLMs can be used when the residuals from a General Linear Model are not normal (based on normal/gaussian distribution) between dependent and independent variables. GLMs was performed on the datasets collected in chapter 2

and 3, using a binomial error family and logit link function. However, when GLMs were run, model validations indicated that there was high spatial autocorrelation. To correct for this, GLMMs were used where fixed and random effects were identified and included. Transect location was a suitable random factor within the models and removed identified spatial autocorrelation. GLMs that do not have spatial autocorrelation present, can be used for ecological studies. These could also be considered for studies that do not have pseudo-replication present, which will need to be determined before appropriate models are tested.

Model selection methods were also investigated for Chapter 3, where impacts and recovery were added together for the first data exploration. This was identified as unsuitable (as the variables were correlated), but methods of model selection and model averaging were still investigated within the early stages of data exploration. Designing an appropriate model for a dataset requires decisions based on terms that you want to include. Model selection can be time consuming, and a subjective approach (Crawley, 2007) when choosing the “best” model.

There are several model selection approaches that can be applied, where many scientists and researchers are now moving towards comparisons of models using an information-theory approach such as Akaike Information Criterion (AIC) (Grueber et al., 2011; Richards et al., 2011). There are numerous model selection criteria based on mathematical information theory (IT) that can be used to select models from among a set of candidate models. The most used information criteria in ecology and evolution are Akaike’s Information Criterion (AIC), the corrected AIC<sub>c</sub> (corrected for small sample sizes), and the Bayesian Information Criterion (BIC, also known as the Schwarz

Criterion) (Johnson and Omland, 2004). Advantages include not relying on a single model; models can be ranked and weighted according to their fit to the observed data and the best supported models can be averaged to get parameter estimates.

AIC is a measure of the goodness of fit of a model, where an “efficient” model explains a relatively large amount of the variance in  $y$ , using few independent variables to do so. This approach examines the model fitness. The lower the AIC value the better the model, which can then be used for further inference (Zuur et al., 2009). AIC is calculated following this equation:

$$AIC = -2\text{Log}L + 2p$$

$AIC_c$  can be used when there are small sample sizes and can correct for that limitation. It is recommended to use this when  $n/k$  is less than 40, with  $n$  being the sample size (i.e. total number of observations) and  $k$  being the total number of parameters in the most saturated model, including both fixed and random effects, as well as intercepts (Symonds and Moussalli, 2011).  $AIC_c$  is calculated as follows:

$$AIC_c = AIC + 2k(k+1)n^{-k-1}$$

Within data Chapter 3, the sample acquired applied to  $n/k$  would produce  $> 40$ , therefore AIC would be applicable. However, this approach would not have been suitable for our analyses in Chapter 3 as impacted trees and all trees (both impacted and not impacted by elephants) were used

for testing, and it isn't possible to compare AIC values between models based on different subsets of data. AIC should therefore be a consideration where no caveats are present. Consideration was also given to using the AIC approach within Chapter 3, when preliminary investigations discerned that impact and recovery were independent variables. However, as these were added together, this was later deemed an unsuitable approach. Future studies could employ the AIC method if a suitable research question and hypothesis testing approach is present within the study.

Model averaging is an approach that can follow model selection testing, such as AIC. AIC is a measure of how good a model is among candidate sets; therefore, it is prone to poor choices, in that the chosen variables must be meaningful for the chosen study. Therefore, it's important to assess the goodness of fit ( $\chi^2$  or  $R^2$ ) that includes all variables of the study, and inference from that can determine whether the approximating model (model averaging) will be a good fit. Model averaging can either be used directly within a model selection object in statistical programmes. This method is useful when model selection uncertainty is evident. This method produces parameter and error estimates that derive weighted averages across multiple models (Symonds and Moussalli, 2011). There is still uncertainty whether model averaging improves inference (Richards, 2005; Richards et al., 2011), but this approach has proved to be an effective method within ecological studies.

There are different types of model averaging approaches that can be used for: Full averaging and conditional averaging. Conditional averaging is derived from Burnham and Anderson (2002). This method keeps the averaged parameter in the original scale and is applied when there is strong

support for the best AIC model. Alternatively, unconditional variance can be used, which is an estimate of variance and is not conditional on a single model and is typically more significant than inference based on one model (Symonds and Moussalli, 2011).

Secondly, full model averaging can be applied when there is high model selection uncertainty, where for example the best AIC model is not strongly weighted (Richards et al., 2011). This is the case when inference is based on all models in the candidate set. Full model averaging produces a predictive formula for the global model, assuming that a variable is included in every model, but in some models, the corresponding coefficient (and its respective variance) is zero. This method can be used in studies where the aim is to formulate how a particular predictor relates to the response variable, which is applicable for ecological studies.

### **6.6.3 Elephant-induced vegetation change on a temporal extent**

To determine elephant impact and recovery, we considered the application of remote sensing approaches, as they provide an array of imagery with various spectral, spatial, radioactive and temporal characteristics that can be used in broad vegetation studies (Xie et al., 2008). Nkosi et al. (2019) reviewed the methods used by different studies to determine the impact of elephants on woody vegetation in sub-Saharan Africa over a 47-year period (1970 – 2017). Though there were various studies that utilised remote sensing methods, none had considered the use of 3-D photogrammetric point clouds as a means to determine elephant impact. The potential of using this remote sensing method was explored in Chapter 4.

This method proved to be effective in determining vegetation height from aerial photographs. Raw data were obtained from the National Geo-Spatial Information (NGI) and GeoSpace International. The data that was analysed in Chapter 4, was from 2018 which was acquired from a digital camera (Table 6.1) and could be tested against vegetation height data that was acquired in Chapter 2 (Appendix 3; field data). As identified in Chapter 4, there is scope to conduct a temporal study within KPGR, with data acquired for previous years.

Vertical, stereo aerial photographs have now been received from aerial surveys conducted of KPGR between 1954 and 2018. This holds great potential for future research projects based around the following research questions:

- Can aerial photographs be used to determine the change in vegetation heights pre- and post-elephant introduction within a reserve? (*Using home- and core-range of elephants since 1999 (see Appendix 3)*).
- How can we use historical aerial photographs to determine long term vegetation change over time (1954-2018)?
- Can aerial photographs be used to determine tree metrics over a large extent: within a protected area and outside of a protected area?

Even though this offers great opportunity for a study on a temporal extent, the spatial extent varies for each year that the air surveys were conducted. Due to the historical methods that have been used to acquire imagery, both digital and analogue photographs have been acquired at different

scales. According to the NGI, aerial photographs of the entire reserve are available for research purposes, however the years 2008-2012 have yet to be received in the correct format (orthophotos only) (Table 6.1).

**Table 6.1** KPGR archive airphoto details derived from the NGI and GeoSpace International

<b>Year</b>	<b>Date</b>	<b>Approximate local time</b>	<b>Analogue/Digital</b>	<b>Approximate nominal scale or GSD</b>	<b>Format</b>
<b>1954</b>	29th June	1:15pm	Analogue	1:36,000	Greyscale, individual frames
<b>1970</b>	10th July	11:00am	Analogue	1:33,000	Greyscale, individual frames
<b>1986</b>	31st May	12 Noon	Analogue	1:50,000	Greyscale, individual frames
<b>1995</b>	29th July	2:30pm	Analogue	1:20,000	Greyscale, individual frames
<b>1997</b>	10th August	1:00pm	Analogue	1:60,000	Greyscale, individual frames

<b>2003</b>	2nd & 5th October	2:15pm	Analogue	1:32,000	Greyscale, individual frames
<b>2008</b>	-	-	Digital	50cm	Orthophoto only
<b>2012</b>	-	-	Digital	50cm	Orthophoto only
<b>2015</b>	-	-	Digital	50cm	RGB orthophoto
<b>2018</b>	9th June	10:30	Digital	25cm	RGBN individual frames

Therefore, conducting a study on a temporal scale will require careful application depending on the scale of aerial photographs and additional information throughout the analysis process that was shown in Chapter 4.

## **6.6.4 Elephant management**

### ***6.6.4.1 Immunocontraception***

Various methods have been used to control elephant numbers in areas where elephants are present in high densities (see Chapter 1). One of the aims of this research project was to identify elephant impact on vegetation and the levels of recovery, which considered habitat use as a key determinant

of impact (Chapter 2). The data that was collected on the effects of elephants on vegetation was within a high-density elephant reserve: KPGR. The population of elephants within this reserve is managed by immunocontraception to limit numbers. Bull elephants on the reserve were administered with GnrH contraception in 2012 and this was stopped in 2016 (administered by Professor Bertschinger). The movement patterns of these elephants are shown to be uncharacteristic to the natural behaviours of elephants where the breeding herd and bull elephants only associate with each other during mating (Buss, 1961). However, using twice daily sightings and home range and core range analysis of these data, the bull elephants were shown to move concurrently with the breeding herd (see Appendix 3). One reason for this behaviour is that the bulls were young at the time of introduction to the reserve, so they have “grown up” with the herd, which has also been shown in a group of rewilded elephants where a single bull mirrors the herd’s movement (from a conversation with Dr Audery Delsink).

This behaviour could be explained by the age of immunocontraception administration, but this could also be attributed to there not being an unrelated mature bull within the reserve. Contrary to assumptions that older male elephants are redundant within the population, recent research has suggested that older male elephants act as repositories for ecological knowledge (Allen et al., 2020). As mentioned in Chapter 1, the long-term implications of contraceptive methods with regards to elephant’s societal change has not been evaluated (Nyakaana et al., 2001; Wittemyer et al., 2009). Within large areas without contraception, bull elephants have been shown to exert a greater impact on vegetation in comparison to cows and calves (Greyling, 2004). However, the influence that immunocontraception has on elephant herd dynamics and the subsequent impact to

vegetation is not known. This observed behaviour change should be considered in future studies of elephant impacts to vegetation, as bull elephants on contraception could show less aggressive behaviours and therefore less impact to vegetation. This would need to be investigated in other reserves, where long-term ranges of elephants are known within enclosed areas, which could give new insights into elephant impact on vegetation.

#### **6.6.4.2 Beehive deterrent project**

Strategies aimed at directly protecting resource such as large trees are a key method of management to mitigate elephant impact within fenced environments (Derham et al., 2016; Cook et al., 2018). One such method is the use of African honeybees (*Apis mellifera* subsp. *scutellata*) which is effective at protecting individual trees from elephant impact as elephants are afraid of bees and have been shown to actively trees with beehives on/near. Simultaneously, beehives used as a mitigation strategy provide pollination to flora and honey as a byproduct which can be harvested for income (Cook et al., 2018). Though beehives can provide benefits to the local communities, they can be expensive to set up, sensitive to drought and overload the environment with honeybees which may in turn exclude other pollinators.

An alternative novel approach that was a consideration during this PhD research was using Artificial Intelligence (AI) to generate AI beehives. The AI beehive would be trained (using CNN and NVIDIA) to respond when elephants are within a certain distance, which cause responses that simulate hive activity i.e., noise, pheromone production etc. Our AI solution will enable protection of trees and farmland all year round, as it will not be affected by seasonality, when resources are

low for the honeybees. In addition, this solution will not need regular monitoring, as the aim is to create a self-sustaining unit. This was beyond the scope of this project, but project planning was initiated for future research opportunities. After networking with data scientists, this project presented an opportunity to be developed with Project 15, Microsoft (<https://microsoft.github.io/project15/>). Project 15 from Microsoft is an open platform for conservation and ecologically sustainable solutions. Working with data science experts and the latest Microsoft cloud and Internet of Things (IoT) technologies, this project will aim to develop a suitable platform to develop AI beehives that can be tested for efficiency in reducing elephant impact to vegetation and can also be used as a method of deterring elephants from breaking down fences. This project will work with the following collaborators: Ajit Jaokar (University of Oxford); Ayse Mutl (Data Scientist, Feynlabs); Associate Professor Lee Scott (University College London (UCL) and Microsoft); Sarah Maston (Senior Solutions Architect, Microsoft); and Kate Gilman Williams (CEO & Founder of Kids Can Save Animals). Collaborators from the University of Oxford, UCL and Microsoft will support the project with their expertise in data science, and Kate Gilman Williams will benefit the project with her skills in advocacy and public speaking to younger demographics. The project funding is still to be determined.

### **6.6.5 Public engagement**

Alongside this research project, public engagement and outreach activities were conducted and aimed to provide impact in various ways. This involved numerous talks to the public, activities with local communities and events at science festivals (both face to face and virtually). With the aim of increasing public awareness of elephants and elephant conservation, these engagement

activities were mostly targeted at the younger demographic. The need for improved methods of engagement with this age group has been identified in Chapter 5, where focussing concepts of environmental education can foster positive attitudes towards younger generations. A future project that has been developed alongside this thesis is a children's picture book educating children on the importance of elephants within the ecosystem. Information surrounding elephant conservation that has been identified throughout this thesis has been incorporated in the text of this book. This way, research can be communicated to school children effectively.

The manuscript was submitted to Inspire Bytes Publishers in 2021 and accepted for publication. This book will form part of a series in their 'Conservation4Kids' initiative ([www.conservation4kids.com](http://www.conservation4kids.com)), and a supplementary workbook will be developed.

## References

- Abdi, H. and Valentin, D., 2007. Multiple correspondence analysis. *Encyclopedia of Measurement and Statistics*, 2(4), 651-657.
- Alagoz, B. and Akman, O., 2016. A Study towards Views of Teacher Candidates about National and Global Environmental Problems. *International Journal of Research in Education and Science*, 2(2), 483-493.
- Almeida, D.R.A., Broadbent, E.N., Zambrano, A.M.A., Wilkinson, B.E., Ferreira, M.E., Chazdon, R., Meli, P., Gorgens, E.B., Silva, C.A., Stark, S.C. and Valbuena, R., 2019. Monitoring the structure of forest restoration plantations with a drone-lidar system. *International Journal of Applied Earth Observation and Geoinformation*, 79, 192-198.
- Ancrenaz, M., Dabek, L. and O'Neil, S., 2007. The costs of exclusions: Recognizing a role for local communities in biodiversity conservation. *PLoS Biol*, 59(11), e289.
- Ardoin, N.M., Bowers, A.W. and Gaillard, E., 2020. Environmental education outcomes for conservation: A systematic review. *Biological Conservation*, 241,108-224.
- Ashenafi, Z.T. and Leader-Williams, N., 2005. Indigenous common property resource management in the Central Highlands of Ethiopia. *Human Ecology*, 33(4), 539-563.
- Asner, G., Levick, S., Kennedy-Bowdoin, T., Knapp, D., Emerson, R., Jacobson, J., Colgan, M. and Martin, R., 2009. Large-scale impacts of herbivores on the structural diversity of African savannas. *Proceedings of the National Academy of Sciences*, 106(12), 4947-4952.
- Asner, G.P., Knapp, D.E., Kennedy-Bowdoin, T., Jones, M.O., Martin, R.E., Boardman, J. and Hughes, R.F., 2008. Invasive species detection in Hawaiian rainforests using airborne imaging spectroscopy and LiDAR. *Remote Sensing of Environment*, 112(5), 1942-1955.
- Asner, G.P., Scurlock, J.M. and A. Hicke, J., 2003. Global synthesis of leaf area index observations: implications for ecological and remote sensing studies. *Global Ecology and Biogeography*, 12(3),191-205.
- Asner, G.P., Vaughn, N., Smit, I.P. and Levick, S., 2016. Ecosystem-scale effects of megafauna in African savannas. *Ecography*, 39(2), 240-252.
- Asunta, T., 2003. Knowledge of environmental issues: where pupils acquire information and how it affects their attitudes, opinions, and laboratory behaviour. *Jyväskylä Studies in Education, Psychology and Social Research* 221 (Jyväskylä. University of Jyväskylä).
- Athman, J. and Monroe, M.C., 2004. The effects of environment-based education on students'

- achievement motivation. *Journal of Interpretation Research*, 9(1), 9-25.
- Awasthy, M., Popovic, A.Z. and Linklater, W.L., 2012. Experience in local urban wildlife research enhances a conservation education programme with school children. *Pacific Conservation Biology*, 18(1), 41-46.
- Ayeni, J. S. O., 1975. Utilization of waterholes in Tsavo National Park (East). *East African Wildlife Journal*, 13, 305–323.
- Ballantyne, R., Packer, J., Hughes, K. and Dierking, L., 2007. Conservation learning in wildlife tourism settings: Lessons from research in zoos and aquariums. *Environmental Education Research*, 13(3), 367-383.
- Bandara, R. and Tisdell, C., 2003. Comparison of rural and urban attitudes to the conservation of Asian elephants in Sri Lanka: empirical evidence. *Biological Conservation*, 110 (3), 327-342.
- Bandara, R. and Tisdell, C., 2003. Use and non-use values of wild Asian elephants: a total economic valuation approach. *Sri Lanka Journal of Economics*, 4(2), 3–30.
- Barnes, R., Barnes, K. and Kapela, E., 1994. The long-term impact of elephant browsing on baobab trees at Msembe, Ruaha National Park, Tanzania. *African Journal of Ecology*, 32(3), 177-184.
- Barney, E.C., Mintzes, J.J. and Yen, C.F., 2005. Assessing knowledge, attitudes, and behavior toward charismatic megafauna: The case of dolphins. *The Journal of Environmental Education*, 36(2), 41-55.
- Barraza, L. and Walford, R.A., 2002. Environmental education: A comparison between English and Mexican school children. *Environmental Education Research*, 8(2), 171-186.
- Barrett, A.S. and Brown, L.R., 2012. A novel method for estimating tree dimensions and calculating canopy volume using digital photography. *African Journal of Range and Forage Science*, 29(3), 153-156.
- Bates, L.A., Sayialel, K.N., Njiraini, N.W., Poole, J.H., Moss, C.J. and Byrne, R.W., 2008. African elephants have expectations about the locations of out-of-sight family members. *Biology Letters*, 4(1), 34-36.
- Baxter, P., and Getz, W., 2005. A model-framed evaluation of elephant effects on tree and fire dynamics in African savannas. *Ecological Applications*, 15(4), 1331-1341.
- Bell, R. H. B. 1984. Majete Game Reserve: report of an ulendo and suggestions for management And utilisation (Malawi Government Report WRU/1//50/5/3//1). Kasungu, Malawi:

Department of National Parks and Wildlife.

- Bennett, E.L., 2015. Legal ivory trade in a corrupt world and its impact on African elephant populations. *Conservation Biology*, 29(1), 54-60.
- Ben-Shahar, R., 1993. Patterns of elephant damage to vegetation in northern Botswana. *Biological Conservation*, 65(3), 249-256.
- Ben-Shahar, R., 1996. Woodland dynamics under the influence of elephants and fire in northern Botswana. *Vegetatio*, 123(2), 153-163.
- Ben-Shahar, R., 1998. Elephant density and impact on Kalahari woodland habitats. *Transactions of the Royal Society of South Africa*, 53(2), 149-155.
- Biggs, D., Holden, M.H., Braczkowski, A., Cook, C.N., Milner-Gulland, E.J., Phelps, J., Scholes, R.J., Smith, R.J., Underwood, F.M., Adams, V.M. and Allan, J., 2017. Breaking the deadlock on ivory. *Science*, 358(6369), 1378-1381.
- Biggs, R. and Jacobs, O., 2002. The impact of the African elephant on marula trees in the Kruger National Park. *South African Journal of Wildlife Research*, 32(1), 13-22.
- Birkett, A. and Stevens-wood, B., 2005. Effect of low rainfall and browsing by large herbivores on an enclosed savannah habitat in Kenya. *African Journal of Ecology*, 43, 123–130.
- Biru, Y., and A. Bekele., 2012. Food habits of African elephant (*Loxodonta africana*) in Babile Elephant Sanctuary, Ethiopia. *Tropical Ecology*, 53, 43–52.
- Blanc, J., 2008. *Loxodonta africana*. The IUCN Red List of Threatened Species 2008: e.T12392A3339343. <http://dx.doi.org/10.2305/IUCN.UK.2008.RLTS.T12392A3339343.en>. Downloaded on 01 August 2019.
- Blanc, J.J. and Barnes, R.F., 2007. African elephant status report 2007: an update from the African elephant database (No. 33). IUCN.
- Bohlin, J., Wallerman, J. and Fransson, J.E., 2012. Forest variable estimation using photogrammetric matching of digital aerial images in combination with a high-resolution DEM. *Scandinavian Journal of Forest Research*, 27(7), 692-699.
- Bond, W.J., 1994. Keystone species. In *Biodiversity and Ecosystem Function*, 237-253. Springer, Berlin, Heidelberg.
- Bond, W.J., 2008. What limits trees in C4 grasslands and savannas?. *Annual review of Ecology, Evolution, and Systematics*, 39, 641-659.
- Boone, R.B. and Hobbs, N.T., 2004. Lines around fragments: effects of fencing on large herbivores. *African Journal of Range and Forage Science*, 21(3), 147-158.

- Bork, E.W. and Su, J.G., 2007. Integrating LIDAR data and multispectral imagery for enhanced classification of rangeland vegetation: A meta-analysis. *Remote Sensing of Environment*, 111(1), 11-24.
- Botes, A., McGeoch, M.A. and Van Rensburg, B.J., 2006. Elephant-and human-induced changes to dung beetle (Coleoptera: Scarabaeidae) assemblages in the Maputaland Centre of Endemism. *Biological Conservation*, 130(4), 573-583.
- Boundja, R., and Midgley, J., 2010. Patterns of elephant impact on woody plants in the Hluhluwe Imfolozi park, Kwazulu-Natal, South Africa. *African Journal of Ecology*, 48(1), 206-214.
- Bowd, A.D. and Bowd, A.C., 1989. Attitudes toward the treatment of animals: A study of Christian groups in Australia. *Anthrozoös*, 3(1), 20-24.
- Bowland, J. M., and R. I. Yeaton., 1997. Impact of domesticated African elephants *Loxodonta africana* on Natal bushveld. *South African Journal of Wildlife Research*, 72, 31–36.
- Brandtberg, T., Warner, T.A., Landenberger, R.E. and McGraw, J.B., 2003. Detection and analysis of individual leaf-off tree crowns in small footprint, high sampling density lidar data from the eastern deciduous forest in North America. *Remote Sensing of Environment*, 85(3), 290-303.
- Brewer, C., 2002. Outreach and partnership programs for conservation education where endangered species conservation and research occur. *Conservation Biology*, 16(1), 4-6.
- Brits, J., van Rooyen, M. and van Rooyen, N., 2000. Technique to study the impact of large herbivores on woody vegetation within piospheres. *Koedoe*, 43(2), 47–56.
- Brooks, B.R., Bruccoleri, R.E., Olafson, B.D., States, D.J., Swaminathan, S.A. and Karplus, M., 1983. CHARMM: a program for macromolecular energy, minimization, and dynamics calculations. *Journal of Computational Chemistry*, 4(2), 187-217.
- Brooks, P., Macdonald, I., and Owen-Smith, N., 1983. The Hluhluwe-Umfolozi Reserve: an ecological case history. *Management of large mammals in African conservation areas*, 51-77.
- Buchanan, G.M., Fishpool, L.D., Evans, M.I. and Butchart, S.H., 2013. Comparing field-based monitoring and remote-sensing, using deforestation from logging at Important Bird Areas as a case study. *Biological Conservation*, 167, 334-338.
- Bunney, K., Bond, W.J. and Henley, M., 2017. Seed dispersal kernel of the largest surviving megaherbivore—the African savanna elephant. *Biotropica*, 49(3), 395-401.
- Burghardt, G.M. and Herzog, H.A. 1989. Animals, evolution, and ethics. *In Perception of Animals*

- in American Culture.*, 129-151.
- Burnett, E., Sills, E., Peterson, M.N. and DePerno, C., 2016. Impacts of the conservation education program in Serra Malagueta Natural Park, Cape Verde. *Environmental Education Research*, 22(4), 538-550.
- Anderson, D. and Burnham, K., 2004. Model selection and multi-model inference. Second. NY: Springer-Verlag, 63(2020), 10.
- Buss, I., 1961. Some observations on food habits and behavior of the African elephant. *The Journal of Wildlife Management*, 25(2), 131-148.
- Byrne, R.W. and Bates, L.A., 2011. Elephant Cognition: What We Know about What Elephants Know. In *The Amboseli elephants*, 174-184. University of Chicago Press
- Calenge, C., D. Maillard, J. M. Gaillard, L. Merlot, and R. Peltier., 2002. Elephant damage to trees of wooded savanna in Zakouma National Park. *Journal of Tropical Ecology*, 18, 599–614.
- Campbell, B., Butler, J., Mapaure, I., Vermeulen, S., and Mashove, P., 1996. Elephant damage and safari hunting in *Pterocarpus angolensis* woodland in northwestern Matabeleland, Zimbabwe. *African Journal of Ecology*, 34(4), 380-388.
- Campos-Arceiz, A. and Blake, S., 2011. Megagardeners of the forest—the role of elephants in seed dispersal. *Acta Oecologica*, 37(6), 542-553.
- Capstick, P.H., 1984. *Safari: The Last Adventure*. St. Martin's Press. 165.
- Carr, J.C. and Slyder, J.B., 2018. Individual tree segmentation from a leaf-off photogrammetric point cloud. *International Journal of Remote Sensing*, 39(15-16), 5195-5210.
- Carruthers, J., Boshoff, A., Slotow, R., Biggs, H.C., Avery, G., Matthews, W., Scholes, R.J. and Mennell, K.G., 2008. The elephant in South Africa: history and distribution.
- Castelda, S. M., Napora, E. S., Nasser, N. A., Vyas, D. K. and Schulte, B. A., 2011. Diurnal co-occurrence of African elephants and other mammals at a Tanzanian waterhole. *African Journal of Ecology*, 49, 250–252.
- Caughley, G., 1976. The elephant problem—an alternative hypothesis. *African Journal of Ecology*, 14(4), 265-283.
- Chamaillé-Jammes, S., Fritz, H., and Murindagomo, F., 2007. Climate-driven fluctuations in surface-water availability and the buffering role of artificial pumping in an African savanna: Potential implication for herbivore dynamics. *Austral Ecology*, 32(7), 740–748.
- Chang'a, A., Souza de, N., Muya, J., Keyyu, J., Mwakatobe, A., Malugu, L., Ndossi, H.,

- Konuche, J., Omondi, R., Mpinge, A., and Hahn, N., 2016. Scaling-up the use of chili fences for reducing human-elephant conflict across landscapes in Tanzania. *Tropical Conservation Science*, 9(2), 921-930.
- Chase, M.J., Schlossberg, S., Griffin, C.R., Bouché, P.J., Djene, S.W., Elkan, P.W., Ferreira, S., Grossman, F., Kohi, E.M., Landen, K. and Omondi, P., 2016. Continent-wide survey reveals massive decline in African savannah elephants. *PeerJ*, 4, 23-54.
- Chatterjee, R., 2008. Protecting farmlands and conserving elephants. *Environmental Science and Technology*, 42, 7029-7029.
- Child, G., Parris, R., and Le Riche, E., 1971. Use of mineralised water by Kalahari wildlife and its effects on habitats. *East African Wildlife Journal*, 9, 125–142.
- CITES., 2018. Status of Elephant Populations, Levels of Illegal Killing and the Trade in Ivory: A Report to the CITES Standing Committee. Document 49.1 Annex I. In: Standing Committee 70 (ed.). CITES, Geneva.
- CITES., 2019. Report and Addendum to the CITES Conference of the Parties Doc.69.2. Conference of the Parties 18. Geneva.
- Clayton, S. and Karazsia, B.T., 2020. Development and validation of a measure of climate change anxiety. *Journal of Environmental Psychology*, 69, 101-434.
- Cochrane, E.P., 2003. The need to be eaten: *Balanites wilsoniana* with and without elephant seed-dispersal. *Journal of Tropical Ecology*, 19(5),579-589.
- Coetzee, B. J., A. H. Engelbrecht, S. C. J. Joubert, and P. F. Retief., 1979. Elephant impact in *Sclerocarya caffra* trees in *Acacia nigrescens* tropical plains thornveld of the Kruger National Park. *Koedoe*, 22, 39–60.
- Collins, M., Dasgupta, S., and Schapire, R. E., 2002. A generalization of principal components analysis to the exponential family. *In Advances in Neural Information Processing Systems*. 617-624.
- Collinson, R., 1983. Pilanesberg’s policy on providing artificial water points for game. Part 4: The implications of providing artificial water points indiscriminately. *Tshomarelo News*, 13, 17– 26.
- Colomina, I. and Molina, P., 2014. Unmanned aerial systems for photogrammetry and remote sensing: A review. *ISPRS Journal of Photogrammetry and Remote Sensing*, 92, 79-97.
- Conforti, V.A. and de Azevedo, F.C.C., 2003. Local perceptions of jaguars (*Panthera onca*) and pumas (*Puma concolor*) in the Iguaçú National Park area, south Brazil. *Biological conservation*, 111(2), 215-221.

- Cook, R.M. and Henley, M.D., 2019. Complexities associated with elephant impact on *Sclerocarya birrea* subsp. *caffra* in the Greater Kruger National Park. *South African Journal of Botany*, 121, 543-548.
- Cook, R.M. and Henley, M.D., 2019. The management dilemma: Removing elephants to save large trees. *Koedoe: African Protected Area Conservation and Science*, 61(1), 1-12.
- Cook, R.M., Parrini, F., King, L.E., Witkowski, E.T.F. and Henley, M.D., 2018. African honeybees as a mitigation method for elephant impact on trees. *Biological Conservation*, 217, 329-336.
- Cook, R.M., Witkowski, E.T.F., Helm, C.V., Henley, M.D. and Parrini, F., 2017. Recent exposure to African elephants after a century of exclusion: Rapid accumulation of marula tree impact and mortality, and poor regeneration. *Forest Ecology and Management*, 401, 107-116.
- Cowie, R.H., Logan, J.W., Wood, T.G., 1989. Termite (*Isoptera*) damage and control in tropical forestry with special reference to Africa and Indo-Malaysia: a review. *Bulletin of Entomological Research*, 79(2), 173–184.
- Cronje, H. P., Cronje, I. and Botha, A. J., 2005. The distribution and seasonal availability of surface water on the Manyeleti Game Reserve, Limpopo Province, South Africa. *Koedoe*, 48(2), 11–21.
- Crosmary, W., Valeix, M., Fritz, H., Madzikanda, H., and Côté, S. D., 2012. African ungulates and their drinking problems: hunting and predation risks constrain access to water. *Animal Behaviour*, 83(1), 145–153.
- Croze, H., 1974. The Seronera bull problem: the elephants. *African Journal of Ecology*, 12(1), pp.1-27.
- Cumming, D., 1981. The management of elephant and other large mammals in Zimbabwe. In *Problems in Locally Abundant Wild Animals*, 91-118. Academic Press, New York.
- Cumming, D., Brock Fenton, M., Rautenbauch, I., Taylor, R., Cumming, G., Cumming, M., Dunlop, J., Ford, A., Hovorka, M., Johnston, D., Kalcounis, M., Mahlangu, Z. and Portfors, C., 1997. Elephants, woodlands and biodiversity in southern Africa. *South African Journal of Science*, 93, 231–236.
- Cushman, S.A., Chase, M. and Griffin, C., 2010. Mapping landscape resistance to identify corridors and barriers for elephant movement in southern Africa. In *Spatial Complexity, Informatics, and Wildlife Conservation*, 349-367. Springer, Tokyo.
- Dalponte, M., Ørka, H.O., Ene, L.T., Gobakken, T. and Næsset, E., 2014. Tree crown delineation and tree species classification in boreal forests using hyperspectral and ALS data.

*Remote Sensing of Environment*, 140, 306-317.

- Damerell, P., Howe, C. and Milner-Gulland, E.J., 2013. Child-orientated environmental education influences adult knowledge and household behaviour. *Environmental Research Letters*, 8(1), 015-016.
- Dandois, J.P. and Ellis, E.C., 2013. High spatial resolution three-dimensional mapping of vegetation spectral dynamics using computer vision. *Remote Sensing of Environment*, 136, 259-276.
- Davies, A.B., Baldeck, C.A. and Asner, G.P., 2016. Termite mounds alter the spatial distribution of African savanna tree species. *Journal of Biogeography*, 43(2), 301-313.
- Davies, A.B., Gaylard, A. and Asner, G.P., 2018. Megafaunal effects on vegetation structure throughout a densely wooded African landscape. *Ecological Applications*, 28(2), 398-408.
- de Beer, Y., and R. J. van Aarde., 2008. Do landscape heterogeneity and water distribution explain aspects of elephant home range in southern Africa's arid savannas? *Journal of Arid Environments*, 72, 2017–2025.
- de Beer, Y., W. Kilian, W. Versfeld, and R. J. van Aarde., 2006. Elephants and low rainfall alter woody vegetation in Etosha National Park, Namibia. *Journal of Arid Environments*, 64, 412–421.
- de Boer, W., Ntumi, C., Correia, A. and Mafuca, J., 2000. Diet and distribution of elephant in the Maputo Elephant Reserve, Mozambique. *African Journal of Ecology*, 38(3), 188-201.
- de Boer, W., Van Oort, J., Grover, M. and Peel, M., 2015. Elephant-mediated habitat modifications and changes in herbivore species assemblages in Sabi Sand, South Africa. *European Journal of Wildlife Research*, 61(4), 491-503.
- de Boer, W.F., van Langevelde, F., Prins, H.H.T., de Ruiter, P.C., Blanc, J., Vis, M.J.P., Gaston, K.J. and Hamilton, I.D. 2013. Understanding spatial differences in African elephant densities and occurrence, a continent-wide analysis. *Biological Conservation*, 159, 468–476.
- De Garine-Wichatitsky, M., Fritz, H., Gordon, I. J. and Illius, A. W., 2004. Bush selection along foraging pathways by sympatric impala and greater kudu. *Oecologia*, 141(1), 66–75.
- Dean W, Milton S, and Jeltsch F. 1999. Large trees, fertile islands, and birds in arid savannas. *Journal of Arid Environments*, 41 (1), 61–78.

- Delplanque, A., Foucher, S., Lejeune, P., Linchant, J. and Théau, J., 2021. Multispecies detection and identification of African mammals in aerial imagery using convolutional neural networks. *Remote Sensing in Ecology and Conservation*.
- Delsink, A.K., Van Altena, J.J., Grobler, D., Bertschinger, H., Kirkpatrick, J. and Slotow, R., 2006. Regulation of a small, discrete African elephant population through immunocontraception in the Makalali Conservancy, Limpopo, South Africa. *South African Journal of Science*, 102(9), 403-405.
- Derham, K., Henley, M. D., & Schulte, B. A., 2016. Wire netting reduces African elephant (*Loxodonta africana*) impact to selected trees in South Africa. *Koedoe*, 58(1), 1-7.
- Dohn, J., Augustine, D.J., Hanan, N.P., Ratnam, J. and Sankaran, M., 2017. Spatial vegetation patterns and neighborhood competition among woody plants in an East African savanna. *Ecology*, 98(2), 478-488.
- Domroese, M.C. and Sterling, E.J., 1999. *Interpreting Biodiversity: A Manual for Environmental Educators in the Tropics*. American Museum of Natural History, Center for Biodiversity and Conservation, Central Park West at 79th Street, New York, NY 10024-5192.
- Dore, M. H., 2005. Climate change and changes in global precipitation patterns: what do we know?. *Environment International*, 31(8), 1167-1181.
- Driscoll, J.W., 1992. Attitudes toward animal use. *Anthrozoös*, 5(1), 32-39.
- Du Preez, J. S., and Grobler, I. D., 1977. Drinking times and behaviour at waterholes of some game species in the Etosha National Park. *Madoqua*, 10(1), 61-69.
- Du Toit, R., 2001. *Africa's Big Five*. Struik Publishers. ISBN 978-1-86872-582-3.
- Dublin, H., 1995. Vegetation Dynamic in the Serengeti-Mara Ecosystem: The Role of Elephants, Fire, and Other Factors. *Serengeti II: Dynamics, Management, and Conservation of an Ecosystem*, 2, 71.
- Duncanson, L.I., Dubayah, R.O., Cook, B.D., Rosette, J. and Parker, G., 2015. The importance of spatial detail: Assessing the utility of individual crown information and scaling approaches for lidar-based biomass density estimation. *Remote Sensing of Environment*, 168, 102-112.
- Duporge, I., Isupova, O., Reece, S., Macdonald, D.W. and Wang, T., 2020. Using very-high-resolution satellite imagery and deep learning to detect and count African elephants in heterogeneous landscapes. *Remote Sensing in Ecology and Conservation*. 7(3), 369-381.
- Duro, D.C., Coops, N.C., Wulder, M.A. and Han, T., 2007. Development of a large area biodiversity monitoring system driven by remote sensing. *Progress in Physical*

*Geography*, 31(3), 235-260.

- Eckhardt, H.C., Van Wilgen, B.W. and Biggs, H.C., 2000. Trends in woody vegetation cover in the Kruger National Park, South Africa, between 1940 and 1998. *African Journal of Ecology*, 38(2), 108-115.
- Eddy, T.J., Gallup Jr, G.G. and Povinelli, D.J., 1993. Attribution of cognitive states to animals: anthropomorphism in comparative perspective. *Journal of Social Issues*, 49(1), 87-101.
- Egeru, A., Bernard, B., Henry, M., & Paul, N., 2015. Piosphere syndrome and rangeland degradation in Karamoja sub-region, Uganda. *Resources and Environment*, 5(3), 73– 89.
- Ene, L., Næsset, E. and Gobakken, T., 2012. Single tree detection in heterogeneous boreal forests using airborne laser scanning and area-based stem number estimates. *International Journal of Remote Sensing*, 33(16), 5171-5193.
- Epaphras, A. M., Gereta, E., Lejora, I. A., Ole Meing'ataki, G. E., Ng'umbi, G., Kiwango, Y. and Mtahiko, M. G. G., 2007. Wildlife water utilization and importance of artificial waterholes during dry season at Ruaha National Park, Tanzania. *Wetlands Ecology and Management*, 16(3), 183–188.
- Espinosa, S. and Jacobson, S.K., 2012. Human-wildlife conflict and environmental education: Evaluating a community program to protect the Andean bear in Ecuador. *The Journal of Environmental Education*, 43(1), 55-65
- Estes, R. D., 2012. Horse Antelopes: Tribe Hippotragini. In *The Behavior Guide to African Mammals*, (2), 115–132. Berkeley: University of California Press.
- Evans, J.S. and Cushman, S.A., 2009. Gradient modeling of conifer species using randomforests. *Landscape ecology*, 24(5), 673-683.
- Farmer, H., 2010. Understanding impacts of water supplementation in a heterogeneous landscape. PhD Thesis, University of the Witwatersrand, South Africa.
- Fayrer-Hosken, R.A., Grobler, D., Van Altena, J.J., Bertschinger, H.J. and Kirkpatrick, J.F., 2000. Immunocontraception of African elephants. *Nature*, 407(6801), 149-149.
- Fernando, P., Wikramanayake, E., Weerakoon, D., Jayasinghe, L.K.A., Gunawardene, M. and Janaka, H.K., 2005. Perceptions and patterns of human–elephant conflict in old and new settlements in Sri Lanka: insights for mitigation and management. *Biodiversity and Conservation*, 14(10), 2465-2481.
- Ferreira, S.M., Greaver, C. & Simms, C., 2017, 'Elephant population growth in Kruger National Park, South Africa, under a landscape management approach', *Koedoe*, 59(1),

1427.

- Fleming, P.A. and Loveridge, J.P., 2003. Miombo woodland termite mounds: resource islands for small vertebrates?. *Journal of Zoology*, 259(2), 161-168.
- Foley, C.A.H., Papageorge, S. and Wasser, S.K., 2001. Noninvasive stress and reproductive measures of social and ecological pressures in free-ranging African elephants. *Conservation Biology*, 15(4), 1134-1142.
- Fraser, J., Gupta, R. and Krasny, M.E., 2015. Practitioners' perspectives on the purpose of environmental education. *Environmental Education Research*, 21(5), 777-800.
- Fritz, H., Duncan, P., Gordon, I.J. and Illius, A.W., 2002. Megaherbivores influence trophic guilds structure in African ungulate communities. *Oecologia*, 131(4), 620-625.
- Frost, P., 1996. The ecology of miombo woodlands. In B. Campbell (Ed.), *The Miombo in Transition: Woodlands and Welfare in Africa*, (1), 11–55.
- Fuhlendorf, S. D., Engle, D. M., Kerby, J., & Hamilton, R., 2008. Pyric herbivory: Rewilding landscapes through the recoupling of fire and grazing. *Conservation Biology*, 23(3), 588–598.
- Gadd, M., 1997. Factors influencing the impact of elephants on woody vegetation in private protected areas in South Africa's lowveld. (Doctoral dissertation).
- Gadd, M., 2002. The impact of elephants on the marula tree *Sclerocarya birrea*. *African Journal of Ecology*, 40(4), 328-336.
- Galanti, V., Preatoni, D., Martinoli, A., Wauter, L.A. and Tosi, G., 2006. Space and habitat use of the African elephant in the Tarangire-Manyara ecosystem, Tanzania: implications for conservation. *Mammalian Biology*, 71(2), 99-114.
- Gandiwa, E., Magwati, T., Zisadza, P., Chinuwo, T. and Tafangenyasha, C., 2011. The impact of African elephants on *Acacia tortilis* woodland in northern Gonarezhou National Park, Zimbabwe. *Journal of Arid Environments*, 75(9), 809-814.
- Garstang, M., 2004. Long-distance, low-frequency elephant communication. *Journal of Comparative Physiology*, 190(10), 791-805.
- Gaugris, J., and Van Rooyen, M., 2010. Effects of water dependence on the utilization pattern of woody vegetation by elephants in the Tembe Elephant Park, Maputaland, South Africa. *African Journal of Ecology*, 48(1), 126-134.
- Geerling, G.W., Labrador-Garcia, M., Clevers, J.G.P.W., Ragas, A.M.J. and Smits, A.J.M., 2007. Classification of floodplain vegetation by data fusion of spectral (CASI) and LiDAR data. *International Journal of Remote Sensing*, 28(19), 4263-4284.

- Gertenbach, W. P. D., 1983. Landscapes of the Kruger National Park. *Koedoe*, 26, 9-121.
- Getis, A., 2008. A history of the concept of spatial autocorrelation: A geographer's perspective. *Geographical Analysis*, 40(3), 297-309.
- Getz, W. M., S. Fortmann-Roe, P. C. Cross, A. J. Lyons, S. J. Ryan, and C. C. Wilmers. 2007. LoCoH: Nonparameteric kernel methods for constructing home ranges and utilization distributions. *PLoS One*, 2(2), 207.
- Gobush, K.S., Edwards, C.T.T., Balfour, D., Wittemyer, G., Maisels, F. and Taylor, R.D.,2021. *Loxodonta africana*. The IUCN Red List of Threatened Species 2021: e.T181008073A181022663.
- Goswami, V.R., Vasudev, D. and Oli, M.K., 2014. The importance of conflict-induced mortality for conservation planning in areas of human–elephant co- occurrence. *Biological Conservation*, 176, 191-198.
- Gould, M.S., Lowe, A.J., and Clarke G.P., 1993. The frequency of termite (Isoptera) damage to tree species in Namakutwa forest, Tanzania, *Sociobiology*, 23, 189-198
- Govender, N., 2005. *The effect of habitat alteration by elephants on invertebrate diversity in two small reserves in South Africa* (Doctoral dissertation).
- Graetz, R. D., & Ludwig, J. A., 1978. A method for the analysis of piosphere data applicable to range assessment. *Australian Rangeland Journal*, (1), 126–136.
- Grant, C.C. and Scholes, M.C., 2006. The importance of nutrient hot-spots in the conservation and management of large wild mammalian herbivores in semi-arid savannas. *Biological Conservation*, 130(3), 426-437.
- Grant, R. C. C., Peel, M. J. S., and Bezuidenhout, H., 2011. Evaluating herbivore management outcomes and associated vegetation impacts. *Koedoe*, 53(2), 1–15.
- Greyling, M.D., McCay, M. and Douglas-Hamilton, I., 2004. Green hunting as an alternative to lethal hunting. In *Proceedings of the Symposium on Human-Elephant Relationships and Conflicts*.
- Grodzińska-Jurczak, M., Stepska, A., Nieszporek, K. and Bryda, G., 2006. Perception of environmental problems among pre-school children in Poland. *International Research in Geographical & Environmental Education*, 15(1), 62-76.
- Grueber, C.E., Nakagawa, S., Laws, R.J. and Jamieson, I.G., 2011. Multimodel inference in ecology and evolution: challenges and solutions. *Journal of Evolutionary Biology*, 24(4), 699-711.
- Gubbi, S., Swaminath, M.H., Poornesha, H.C., Bhat, R. and Raghunath, R., 2014. An elephantine

- challenge: human–elephant conflict distribution in the largest Asiatic elephant population, southern India. *Biodiversity and Conservation*, 23(3), 633-647.
- Guldemon, R., Van Aarde, R., 2008. A meta-analysis of the impact of African elephants on savanna vegetation. *The Journal of Wildlife Management*, 72(4), 892-899.
- Guldemon, R.A., Purdon, A. and Van Aarde, R.J., 2017. A systematic review of elephant impact across Africa. *PloS one*, 12(6).
- Gunnthorsdottir, A., 2001. Physical attractiveness of an animal species as a decision factor for its preservation. *Anthrozoös*, 14(4), 204-215.
- Gurusamy, V., Tribe, A., Toukhsati, S. and Phillips, C.J., (2015). Public attitudes in India and Australia toward elephants in zoos. *Anthrozoös*, 28(1), 87-100.
- Guy, P. R. 1989. The influence of elephants and fire on a *Brachystegia- Julbernardia* woodland in Zimbabwe. *Journal of Tropical Ecology*, (5), 215-226.
- Guy, P.R., 1976. The feeding behaviour of elephant (*Loxodonta africana*) in the Sengwa area Rhodesia. *South African Journal of Wildlife Research*, 6(1), 55-63.
- Hacker, C.E. and Miller, L.J., 2016. Zoo visitor perceptions, attitudes, and conservation intent after viewing African elephants at the San Diego Zoo Safari Park. *Zoo Biology*, 35(4), 355-361.
- Hakeem, K.R., Chandna, R., Ahmad, P., Iqbal, M. and Ozturk, M., 2012. Relevance of proteomic investigations in plant abiotic stress physiology. *OmicS: a journal of integrative biology*, 16(11), 621-635.
- Halperin, J. and Geis, K.U., 1999. Lyctidae (Coleoptera) of Israel, their damage and its prevention. *Phytoparasitica*, 27(4), 257-262.
- Hamandawana, H., 2012. The impacts of herbivory on vegetation in Moremi Game Reserve, Botswana: 1967-2001. *Regional Environmental Change*, 12(1), 1–15.
- Harrington, R., Owen-smith, N., Viljoen, P. C., Biggs, H. C., Mason, D. R., & Funston, P., 1999. Establishing the causes of the roan antelope decline in the Kruger National Park, South Africa. *Biological Conservation*, 90, 69–78.
- Harris, G. M., G. J. Russell, R. J. van Aarde, and S. L. Pimm., 2008. Rules of habitat use by elephants *Loxodonta africana* in southern Africa: insights for regional management. *Oryx*, 42, 66–75.
- Harris, W.V., 1968. African termites of the genus *Schedorhinotermes* (Isoptera: *Rhinotermitidae*) and associated termitophiles (Lepidoptera: *Tineidae*). *Proceedings of the Royal Entomological Society of London. Series B, Taxonomy*, 37(8), 103-113

- Hart, B.L. and Hart, L.A., 1994. Fly switching by Asian elephants: tool use to control parasites. *Animal Behaviour*, 48(1), 35-45.
- Hart, L.A. and O'Connell, C.E., 1998. Human conflict with African and Asian elephants and associated conservation dilemmas. *Unpublished Paper. Center for Animals in Society in the School of Veterinary Medicine and Ecology. University of California, Davis*, 95616, 299-315.
- Hatcher, P.E., 1995. Three-way interactions between plant pathogenic fungi, herbivorous insects and their host plants. *Biological Reviews*, 70(4), 639-694.
- Hayward, M. W., & Hayward, M. D., 2012. Waterhole use by African Fauna. *South African Journal of Wildlife Research*, 42, 117–127.
- Hayward, M.W. and Kerley, G.I., 2009. Fencing for conservation: restriction of evolutionary potential or a riposte to threatening processes?. *Biological Conservation*, 142(1), 1- 13.
- Helm, C., and Witkowski, E., 2012. Characterising wide spatial variation in population size structure of a keystone African savanna tree. *Forest Ecology and Management*, 263, 175-188.
- Helm, C., Wilson, G., Midgley, J., Kruger, L. and Witkowski, E.T.F., 2011. Investigating the vulnerability of an African savanna tree (*Sclerocarya birrea* ssp. *caffra*) to fire and herbivory. *Austral Ecology*, 36(8), 964-973.
- Helm, C., Witkowski, E., Kruger, L., Hofmeyr, M. and Owen-Smith, N., 2009. Mortality and utilisation of *Sclerocarya birrea* subsp. *caffra* between 2001 and 2008 in the Kruger National Park, South Africa. *South African Journal of Botany*, 75(3), 475-484.
- Helm, C.V. and Witkowski, E.T., 2013. Continuing decline of a keystone tree species in the Kruger National Park, South Africa. *African Journal of Ecology*, 51(2), .270-279.
- Helm, C.V. and Witkowski, E.T.F., 2012. Characterising wide spatial variation in population size structure of a keystone African savanna tree. *Forest Ecology and Management*, 263, 175-188.
- Herzog Jr, H.A. and Burghardt, G.M., 1988. Attitudes toward animals: Origins and diversity. *Anthrozoös*, 1(4), 214-222.
- Hirschmüller, H., 2011. Semi-global matching-motivation, developments and applications. *Photogrammetric Week 11*, 173-184.
- Hiscocks, K., 1999. The impact of an increasing elephant population of the woody vegetation in southern Sabi Sand Wildtuin, South Africa. *Koedoe*, 42, 47–55.
- Hoare, R., 1999. Determinants of human–elephant conflict in a land-use mosaic. *Journal of*

- Applied Ecology*, 36(5), 689-700.
- Holdo, R., 2003. Woody plant damage by African elephants in relation to leaf nutrients in western Zimbabwe. *Journal of Tropical Ecology*, 189-196.
- Holdo, R.M. and McDowell, L.R., 2004. Termite mounds as nutrient-rich food patches for elephants. *Biotropica*, 36(2), 231-239.
- Holdo, R.M., 2005. Stem mortality following fire in Kalahari sand vegetation: effects of frost, prior damage, and tree neighbourhoods. *Plant Ecology*, 180(1), 77-86.
- Holdo, R.M., 2007. Elephants, fire, and frost can determine community structure and composition in Kalahari woodlands. *Ecological Applications*, 17(2), 558-568.
- Hopcraft, J.G.C., Olf, H. and Sinclair, A.R.E., 2010. Herbivores, resources and risks: alternating regulation along primary environmental gradients in savannas. *Trends in Ecology & Evolution*, 25(2), 119-128.
- Howes, B., Doughty, L., Thompson, S., 2020. African elephant feeding preferences in a small South African fenced game reserve. *Journal for Nature Conservation*, 53, 125700.
- Hrabar, H. and Du Toit, J.T., 2014. Interactions between megaherbivores and microherbivores: elephant browsing reduces host plant quality for caterpillars. *Ecosphere*, 5(1), 1-6.
- Hudak, A.T. and Wessman, C.A., 1998. Textural analysis of historical aerial photography to characterize woody plant encroachment in South African savanna. *Remote Sensing of Environment*, 66(3), 317-330.
- Hungerford, H.R. and Volk, T.L., 1990. Changing learner behavior through environmental education. *The Journal of Environmental Education*, 21(3), 8-21.
- Hyypä, J., Hyypä, H., Leckie, D., Gougeon, F., Yu, X. and Maltamo, M., 2008. Review of methods of small-footprint airborne laser scanning for extracting forest inventory data in boreal forests. *International Journal of Remote Sensing*, 29(5), 1339-1366.
- Hyypä, J., Kelle, O., Lehtikoinen, M. and Inkinen, M., 2001. A segmentation-based method to retrieve stem volume estimates from 3-D tree height models produced by laser scanners. *IEEE Transactions on Geoscience and Remote Sensing*, 39(5), 969-975.
- Ihwagi, F., Chira, R., Kironchi, G., Vollrath, F. and Douglas-Hamilton, I., 2012. Rainfall pattern and nutrient content influences on African elephants' debarking behaviour in Samburu and Buffalo Springs National Reserves, Kenya. *African Journal of Ecology*, 50(2), 152-159.
- Ihwagi, F.W., Vollrath, F., Chira, R.M., Douglas-Hamilton, I. and Kironchi, G., 2010. The impact

- of elephants, *Loxodonta africana*, on woody vegetation through selective debarking in Samburu and Buffalo Springs National Reserves, Kenya. *African Journal of Ecology*, 48(1), 87-95.
- Iqbal, I.A., Osborn, J., Stone, C. and Lucieer, A., 2021. A Comparison of ALS and Dense Photogrammetric Point Clouds for Individual Tree Detection in Radiata Pine Plantations. *Remote Sensing*, 13(17), 3536.
- Irie, N. and Hasegawa, T., 2009. Elephant psychology: What we know and what we would like to know. *Japanese Psychological Research*, 51(3), 177-181
- Ishida, Y., Oleksyk, T.K., Georgiadis, N.J., David, V.A., Zhao, K., Stephens, R.M., Kolokotronis, S.O. and Roca, A.L., 2011. Reconciling apparent conflicts between mitochondrial and nuclear phylogenies in African elephants. *PloS One*, 6(6), 20642.
- Jaakkola, Anttoni, Juha Hyypä, Antero Kukko, Xiaowei Yu, Harri Kaartinen, Matti Lehtomäki, and Yi Lin. "A low-cost multi-sensoral mobile mapping system and its feasibility for tree measurements." *ISPRS Journal of Photogrammetry and Remote Sensing*, 65(6), 514-522.
- Jachmann, H. and Bell, R.H.V., 1985. Utilization by elephants of the *Brachystegia* woodlands of the Kasungu National Park, Malawi. *African Journal of Ecology*, 23(4), 245-258.
- Jachmann, H., 1989. Food selection by elephants in the 'miombo' biome, in relation to leaf chemistry. *Biochemical Systematics and Ecology*, 17(1), 15-24.
- Jacobs, O., and Biggs, R., 2002. The impact of the African elephant on marula trees in the Kruger National Park. *South African Journal of Wildlife Research-24-month delayed open access*, 32(1), 13-22.
- Jakubowski, M.K., Guo, Q. and Kelly, M., 2013. Tradeoffs between lidar pulse density and forest measurement accuracy. *Remote Sensing of Environment*, 130, 245-253.
- Jarman, P. J., 1972. Seasonal distribution of large mammal populations in the unflooded middle Zambezi valley. *Journal of Applied Ecology*, 9, 283-299.
- Järnstedt, J., Pekkarinen, A., Tuominen, S., Ginzler, C., Holopainen, M. and Viitala, R., 2012. Forest variable estimation using a high-resolution digital surface model. *ISPRS Journal of Photogrammetry and Remote Sensing*, 74, 78-84.
- Johansen, K., Phinn, S. and Witte, C., 2010. Mapping of riparian zone attributes using discrete return LiDAR, QuickBird and SPOT-5 imagery: Assessing accuracy and costs. *Remote Sensing of Environment*, 114(11), 2679-2691.
- Johnson, J.B. and Omland, K.S., 2004. Model selection in ecology and evolution. *Trends in Ecology and Evolution*, 19(2), 101-108.

- Jones, B. and Barnes, J., 2007. WWF human wildlife conflict study: Namibian case study. Unpublished report, Macroeconomics Programme and Global Species Programme, WWF, Gland, Switzerland.
- Jones, C.G., Lawton, J.H. and Shachak, M., 1994. Organisms as ecosystem engineers. In *Ecosystem management*, 130-147. Springer, New York, NY.
- Joseph, G.S., Seymour, C.L., Coetzee, B.W., Ndlovu, M., Deng, L., Fowler, K., Hagan, J., Brooks, B.J., Seminara, J.A. and Foord, S.H., 2018. Elephants, termites and mound thermoregulation in a progressively warmer world. *Landscape Ecology*, 33(5), 731- 742.
- Joseph, G.S., Seymour, C.L., Cumming, G.S., Cumming, D.H. and Mahlangu, Z., 2013. Termite mounds as islands: woody plant assemblages relative to termitarium size and soil properties. *Journal of Vegetation Science*, 24(4), 702-711.
- Josse, J. and Husson, F., 2012. Selecting the number of components in principal component analysis using cross-validation approximations. *Computational Statistics and Data Analysis*, 56(6), 1869-1879.
- Jouquet, P., Traoré, S., Choosai, C., Hartmann, C. and Bignell, D., 2011. Influence of termites on ecosystem functioning. Ecosystem services provided by termites. *European Journal of Soil Biology*, 47(4), 215-222.
- Kachamba, D.J., Ørka, H.O., Gobakken, T., Eid, T. and Mwase, W., 2016. Biomass estimation using 3D data from unmanned aerial vehicle imagery in a tropical woodland. *Remote Sensing*, 8(11), 968.
- Kagoro-Rugunda, G., 2004. Crop raiding around Lake Mburo National Park, Uganda. *African Journal of Ecology*, 42(1), 32-41.
- Kandare, K., Ørka, H.O., Dalponte, M., Næsset, E. and Gobakken, T., 2017. Individual tree crown approach for predicting site index in boreal forests using airborne laser scanning and hyperspectral data. *International journal of applied earth observation and geoinformation*, 60, 72-82.
- Kankare, V., Holopainen, M., Vastaranta, M., Puttonen, E., Yu, X., Hyypä, J., Vaaja, M., Hyypä, H. and Alho, P., 2013. Individual tree biomass estimation using terrestrial laser scanning. *ISPRS Journal of Photogrammetry and Remote Sensing*, 75, 64-75.
- Kansky, R. and Knight, A.T., 2014. Key factors driving attitudes towards large mammals in conflict with humans. *Biological Conservation*, 179, 93-105.
- Kapur, R., Perceptions and Reactions of Teachers and Students towards Environmental Education.
- Kasiringua, E., Kopij, G., and Şerban, P., 2017. Daily activity patterns of ungulates at water holes

- during the dry season in the Waterberg National Park, Namibia. *Russian Journal of Theriology*, 2, 129–138.
- Kellert, S.R. and Dunlap, J., 1989. Informal learning at the zoo: A study of attitude and knowledge impacts. *Philadelphia: Zoological Society of Philadelphia*.
- Kellert, S.R., 1980. Contemporary values of wildlife in American society. *Wildlife values*.
- Kellert, S.R., 1980. *Knowledge, affection, and basic attitudes toward animals in American society: Phase III*. US Department of the Interior, Fish and Wildlife Service.
- Kellert, S.R., 1983. *Children's attitudes, knowledge and behaviors toward animals*. US Department of the Interior, Fish and Wildlife Service
- Kelly, A., 2017. Eco-anxiety at university: Student experiences and academic perspectives on cultivating healthy emotional responses to the climate crisis. (Independent Study Project) Collection. 2642.
- Kelly, J.D., 1997. Effective conservation in the twenty-first century: the need to be more than a zoo. One organization's approach. *International Zoo Yearbook*, 35(1), 1-14.
- Kendall, H.A., Lobao, L.M. and Sharp, J.S., 2006. Public concern with animal well-being: Place, social structural location, and individual experience. *Rural Sociology*, 71(3), 399-428.
- Kerley, G., and Landman, M., 2006. The impacts of elephants on biodiversity in the EasternCape Subtropical Thickets: elephant conservation. *South African Journal of Science*, 102(9-10), 395-402.
- Kerley, G.I., Landman, M. and De Beer, S., 2010. How do small browsers respond to resource changes? Dietary response of the Cape grysbok to clearing alien Acacias. *Functional Ecology*, 24(3), 670-675.
- Kideghesho, J.R., Røskaft, E. and Kaltenborn, B.P., 2007. Factors influencing conservation attitudes of local people in Western Serengeti, Tanzania. *Biodiversity and Conservation*, 16(7), 2213-2230.
- Kie, J.G., 2013. A rule-based ad hoc method for selecting a bandwidth in kernel home-range analyses. *Animal Biotelemetry*, 1-13.
- Kim, H.J. and Wasser, S.K., 2019. *Loxodonta africana* subspecies distribution across African Elephant Database Input Zones.
- King, L., Douglas-Hamilton, I., and Vollrath, F., 2011. Beehive fences as effective deterrents for crop-raiding elephants: field trials in northern Kenya. *African Journal of Ecology*, 49(4), 431-439.

- Knafo, A. and Galansky, N., 2008. The influence of children on their parents' values. *Social and Personality Psychology Compass*, 2(3), 1143-1161.
- Koch, B., Heyder, U. and Weinacker, H., 2006. Detection of individual tree crowns in airborne lidar data. *Photogrammetric Engineering and Remote Sensing*, 72(4), 357- 363.
- Koch, B., Heyder, U. and Weinacker, H., 2006. Detection of individual tree crowns in airborne lidar data. *Photogrammetric Engineering and Remote Sensing*, 72(4), 357- 363.
- Kohi, E., de Boer, W., Peel, M., Slotow, R., van der Waal, C., Heitkönig, I., Skidmore, A., and Prins, H., 2011. African elephants *Loxodonta africana* amplify browse heterogeneity in African savanna. *Biotropica*, 43(6), 711-721.
- Koukoulas, S. and Blackburn, G.A., 2005. Mapping individual tree location, height and species in broadleaved deciduous forest using airborne LIDAR and multi-spectral remotely sensed data. *International Journal of Remote Sensing*, 26(3), 431-455.
- Kramer, R., Langholz, J. and Salafsky, N., 2002. The Role of the Private Sector in Protected Area Implementation and Management A Conceptual Framework for Analyzing Effectiveness. *Unpublished document*.
- Kremen, C., Merenlender, A., Murphy, D., 1994 Ecological monitoring: a vital need for integrated conservation and development programs in the tropics. *Conservation Biology*, 8(2), 388-397.
- Kruse, C.K. and Card, J.A., 2004. Effects of a conservation education camp program on campers' self-reported knowledge, attitude, and behavior. *The Journal of Environmental Education*, 35(4), 33-45.
- Kuhar, C.W., Bettinger, T.L., Lehnhardt, K., Townsend, S. and Cox, D., 2007. Into the forest: the evolution of a conservation education program at Kalinzu Forest Reserve, Uganda. *Applied Environmental Education and Communication*, 6(2), 159-166.
- Kwamboka, V. A. 2013. Living in harmony with elephants. *International Zoo Educators Journal*, 49, 43–46.
- Lande, R. and Arnold, S. 1983. The measurement of selection on correlated characters. *Evolution*, 37, 1210-1226.
- Landman, M., Kerley, G., and Schoeman, D., 2008. Relevance of elephant herbivory as a threat to Important Plants in the Addo Elephant National Park, South Africa. *Journal of Zoology*, 274(1), 51-58.
- Lange, R. T., 1969. The piosphere, sheep track and dung patterns. *Journal of Range Management*, 22, 396–400.

- Laws, R., 1970. Elephants as agents of habitat and landscape change in East Africa. *Oikos*, 1-15.
- Leberl, F., Irschara, A., Pock, T., Meixner, P., Gruber, M., Scholz, S. and Wiechert, A., 2010. Point clouds. *Photogrammetric Engineering & Remote Sensing*, 76(10), 1123-1134.
- Lefsky, M.A., Cohen, W.B., Parker, G.G. and Harding, D.J., 2002. Lidar remote sensing for ecosystem studies: Lidar, an emerging remote sensing technology that directly measures the three-dimensional distribution of plant canopies, can accurately estimate vegetation structural attributes and should be of particular interest to forest, landscape, and global ecologists. *BioScience*, 52(1), 19-30.
- Lehmann, C., Prior, L., Williams, R., Bowman, D., 2008. Spatio-temporal trends in tree cover of a tropical mesic savanna are driven by landscape disturbance. *Journal of Applied Ecology*, 45(4), 1304-1311.
- Lehmann, M. B., Funston, P. J., Owen, C. R., Slotow, R., 2008. Home Range Utilisation and Territorial Behaviour of Lions (*Panthera leo*) on Karongwe Game Reserve, South Africa. *PLoS One*, 3(12).
- Leimgruber, P., Gagnon, J.B., Wemmer, C., Kelly, D.S., Songer, M.A. and Selig, E.R., 2003, November. Fragmentation of Asia's remaining wildlands: implications for Asian elephant conservation. In *Animal Conservation forum 6* (4), 347-359. Cambridge University Press.
- Lemieux, A., and Clarke, R., 2009. The international ban on ivory sales and its effects on elephant poaching in Africa. *The British Journal of Criminology*, 49(4), 451-471.
- Leuthold, W., 1977. Changes in tree populations of Tsavo East National Park, Kenya. *East African Wildlife Journal*, 15, 61-69.
- Leuthold, W., 1977. Spatial organization and strategy of habitat utilization of elephants in Tsavo National Park, Kenya. *Zeitschrift für Saugetierkunde*, 42(6), 358-379.
- Leuthold, W., 1996. Recovery of woody vegetation in Tsavo National Park, Kenya, 1970-94. *African Journal of Ecology*, 34(2), 101-112.
- Levick, S. and Rogers, K., 2008. Patch and species-specific responses of savanna woody vegetation to browser exclusion. *Biological Conservation*, 141(2), 489-498.
- Levick, S. R., and K. H. Rogers., 2008. "Structural biodiversity monitoring in savanna ecosystems: Integrating LiDAR and high-resolution imagery through object-based image analysis." In *Object-Based Image Analysis*, 477-491. Springer, Berlin, Heidelberg
- Levick, S., and Asner, G., 2013. The rate and spatial pattern of treefall in a savanna landscape.

- Biological Conservation*, 157, 121-127.
- Levick, S.R., Asner, G.P., Kennedy-Bowdoin, T. and Knapp, D.E., 2009. The relative influence of fire and herbivory on savanna three-dimensional vegetation structure. *Biological Conservation*, 142(8), 1693-1700.
- Lewis, D., 1991. Observations of tree growth, woodland structure and elephant damage on *Colophospermum mopane* in Luangwa Valley, Zambia. *African Journal of Ecology*, 29(3), 207-221.
- Libanda, B. and Blignaut, J., 2007. Tourism's local benefits for Namibia's community-based natural resource management areas. *International Journal for Ecological Economics and Statistics*, 10(W08), 249-259
- Lim, K., Treitz, P., Wulder, M., St-Onge, B. and Flood, M., 2003. LiDAR remote sensing of forest structure. *Progress in Physical Geography*, 27(1), 88-106.
- Lindsey, P., Allan, J., Brehony, P., Dickman, A., Robson, A., Begg, C., Bhammar, H., Blanken, L., Breuer, T., Fitzgerald, K. and Flyman, M., 2020. Conserving Africa's wildlife and wildlands through the COVID-19 crisis and beyond. *Nature Ecology and Evolution*, 4(10), 1300-1310.
- Lisein, J., Pierrot-Deseilligny, M., Bonnet, S. and Lejeune, P., 2013. A photogrammetric workflow for the creation of a forest canopy height model from small unmanned aerial system imagery. *Forests*, 4(4), 922-944.
- Liu, H., Bauer, L.S., Gao, R., 2003. Exploratory survey for the emerald ash borer, *Agrilus planipennis* (Coleoptera: *Buprestidae*), and its natural enemies in China. *Great Lakes Entomologist*, 36, 191-204
- Liu, J., Skidmore, A.K., Jones, S., Wang, T., Heurich, M., Zhu, X. and Shi, Y., 2018. Large off-nadir scan angle of airborne LiDAR can severely affect the estimates of forest structure metrics. *ISPRS Journal of Photogrammetry and Remote Sensing*, 136, 13- 25.
- Liu, K., Shen, X., Cao, L., Wang, G. and Cao, F., 2018. Estimating forest structural attributes using UAV-LiDAR data in Ginkgo plantations. *ISPRS Journal of Photogrammetry and Remote Sensing*, 146, 465-482.
- Loarie, S., Van Aarde, R., P and Pimm, S., 2009. Fences and artificial water affect African savannah elephant movement patterns. *Biological Conservation*, 142(12), 3086-3098.
- Lock, J., 1977. Preliminary results from fire and elephant exclusion plots in Kabalega National Park, Uganda. *African Journal of Ecology*, 15(3), 229-232.
- Lock, J.M., 1993. Vegetation change in Queen Elizabeth National Park, Uganda: 1970– 1988.

- African Journal of Ecology*, 31(2), 106-117.
- Lombard, A., Johnson, C., Cowling, R., and Pressey, R., 2001. Protecting plants from elephants: botanical reserve scenarios within the Addo Elephant National Park, *South Africa. Biological Conservation*, 102(2), 191-203.
- Lombard, A.T., 1995. The problems with multi-species conservation: do hotspots, ideal reserves and existing reserves coincide?. *African Zoology*, 30(3), 145-163.
- Low, A.B. and Rebelo, A.G., 1998. Vegetation of South Africa, Lesotho and Swaziland: A companion to the vegetation map of South Africa, Lesotho and Swaziland.
- Ludwig, F., De Kroon, H. and Prins, H.H., 2008. Impacts of savanna trees on forage quality for a large African herbivore. *Oecologia*, 155(3), 487-496.
- Lueders, I., Young, D., Maree, L., Van der Horst, G., Luther, I., Botha, S., Tindall, B., Fosgate, G., Ganswindt, A. and Bertschinger, H.J., 2017. Effects of GnRH vaccination in wild and captive African Elephant bulls (*Loxodonta africana*) on reproductive organs and semen quality. *PLoS One*, 12(9), 0178270.
- MacGregor, S. D., and T. G. O'Connor., 2004. Response of *Acacia tortilis* to utilization by elephants in a semi- arid African savanna. *South African Journal of Wildlife Research*, 34, 55–66.
- Mackey, R.L., Page, B.R., Duffy, K.J. and Slotow, R., 2006. Modelling elephant population growth in small, fenced, South African reserves. *South African Journal of Wildlife Research-24-month delayed open access*, 36(1), .33-43.
- Makecha, R.N. and Ghosal, R., 2017. Elephant conservation: Reviewing the need and potential impact of cognition-based education. *International Journal of Comparative Psychology*, 30.
- Makhabu, S. W., Balisana, M., and Perkins, J., 2002. Vegetation gradients around artificial waterpoints in the Central Kalahari Game Reserve of Botswana. *African Journal of Ecology*, 40, 103–109.
- Makhabu, S., Skarpe, C., and Hytteborn, H., 2006. Elephant impact on shoot distribution on trees and on rebrowsing by smaller browsers. *Acta Oecologica*, 30(2), 136-146
- Maltamo, M., Mustonen, K., Hyypä, J., Pitkänen, J. and Yu, X., 2004. The accuracy of estimating individual tree variables with airborne laser scanning in a boreal nature reserve. *Canadian Journal of Forest Research*, 34(9), 1791-1801.
- Manning, A.D., Fischer, J. and Lindenmayer, D.B., 2006. Scattered trees are keystone structures—implications for conservation. *Biological Conservation*, 132(3), 311-321.

- Mapaure, I., and Mhlanga, L., 2000. Patterns of elephant damage to *Colophospermum mopane* on selected islands in Lake Kariba, Zimbabwe. *Kirkia*, 189-198.
- Marcinkowski, T. and Reid, A., 2019. Reviews of research on the attitude–behavior relationship and their implications for future environmental education research, *Environmental Education Research*, 25(4), 459-471.
- Margules, C.R. and Pressey, R.L., 2000. Systematic conservation planning. *Nature*, 405(6783), 243-253.
- Maron, J. L., and Crone, E., 2006. Herbivory: effects on plant abundance, distribution and population growth. *Proceedings of the Royal Society Biological Sciences*, 273, 2575– 2584.
- McDuff, M. and Jacobson, S., 2000. Impacts and future directions of youth conservation organizations: Wildlife clubs in Africa. *Wildlife Society Bulletin*, 414-425.
- Mehta, J.N. and Heinen, J.T., 2001. Does community-based conservation shape favorable attitudes among locals? An empirical study from Nepal. *Environmental Management*, 28(2), 165-177.
- Meijer, W., Scheer, S., Whan, E., Yang, C. and Kritski, E. 2018. Demand under the Ban – China Ivory Consumption Research Post-ban 2018.
- Meinhold, J.L. and Malkus, A.J., 2005. Adolescent environmental behaviors: Can knowledge, attitudes, and self-efficacy make a difference?. *Environment and Behavior*, 37(4), 511-532.
- Midgley, J. J., D. Balfour, and G. I. Kerley., 2005. Why do elephants damage savanna trees? *South African Journal of Science*, 101, 213–215.
- Midgley, J.J., Coetzee, B.W., Tye, D. and Kruger, L.M., 2020. Mass sterilization of a common palm species by elephants in Kruger National Park, South Africa. *Scientific Reports*, 10(1), 1-5.
- Midgley, J.J., Lawes, M.J. and Chamailé-Jammes, S., 2010. Savanna woody plant dynamics: the role of fire and herbivory, separately and synergistically. *Australian Journal of Botany*, 58(1), 1-11.
- Milliken, T., Burn, R.W. and Sangalakula, L., 2016. The Elephant Trade Information System (ETIS) and the Illicit Trade in Ivory: A report to the 17th meeting of the Conference of the Parties to CITES. *CoP17 Doc*, 56.
- Mitchell, B.A., Fitzsimons, J.A., Stevens, C.M. and Wright, D.R., 2018. PPA or OECM? Differentiating between privately protected areas and other effective area-based conservation measures on private land. *Parks*, 24(Special Issue), 49-60.

- Mobæk, R., Narmo, A.K. and Moe, S.R., 2005. Termitaria are focal feeding sites for large ungulates in Lake Mburo National Park, Uganda. *Journal of Zoology*, 267(1), 97- 102.
- Moe, S.R., Rutina, L.P., Hytteborn, H. and Du Toit, J.T., 2009. What controls woodland regeneration after elephants have killed the big trees? *Journal of Applied Ecology*, 46(1), 223-230.
- Moncrieff, G.R., Kruger, L.M. and Midgley, J.J., 2008. Stem mortality of *Acacia nigrescens* induced by the synergistic effects of elephants and fire in Kruger National Park, South Africa. *Journal of Tropical Ecology*, 655-662.
- Mondol, S., Moltke, I., Hart, J., Keigwin, M., Brown, L., Stephens, M. and Wasser, S.K., 2015. New evidence for hybrid zones of forest and savanna elephants in Central and West Africa. *Molecular Ecology*, 24(24), 6134-6147.
- Monroe, M. and Krasny, M., 2015. Across the spectrum. *North American Association for Environmental Education: Washington, DC, USA*.
- Morris, B., 2006. The history and conservation of mammals in Malawi. In *Kachere Monographs* (21). East Lansing: African Books Collective Oxford, Michigan State University Press.
- Morrison, T. A., Holdo, R. M., and Anderson, T. M., 2016. Elephant damage, not fire or rainfall, explains mortality of overstorey trees in Serengeti. *Journal of Ecology*, 104(2), 409–418.
- Moseby, K., Lollback, G., and Lynch, C., 2018. Too much of a good thing; successful reintroduction leads to overpopulation in a threatened mammal. *Biological Conservation*, 219, 78-88.
- Mosugelo, D. K., Moe, S. R., Ringrose, S., and Nellemann, C., 2002. Vegetation changes during a 36-year period in northern Chobe National Park, Botswana. *African Journal of Ecology*, 40(3), 232–240.
- Mucina, L. and Rutherford, M.C., 2006. The vegetation of South Africa, Lesotho and Swaziland. *Strelitzia* 19., (South African National Biodiversity Institute: Pretoria, South Africa). *Memoirs of the Botanical Survey of South Africa*.
- Mukinya, J. G., 1977. Feeding and drinking habits of the black rhinoceros in Masai Mara Game Reserve. *East African Wildlife Journal*, 15, 125–138.
- Munyati, C. and Sinthumule, N.I., 2016. Change in woody cover at representative sites in the Kruger National Park, South Africa, based on historical imagery. *SpringerPlus*, 5(1),1-23.
- Musgrave, M.K. and Compton, S.G., 1997. Notes and Records. *African Journal of Ecology*, 35(4), 370-373.
- Mutanga, O., Dube, T. and Ahmed, F., 2016. Progress in remote sensing: vegetation monitoring

- in South Africa. *South African Geographical Journal*, 98(3), 461-471.
- Muvengwi, J., Mbiba, M. and Nyenda, T., 2013. Termite mounds may not be foraging hotspots for mega-herbivores in a nutrient-rich matrix. *Journal of Tropical Ecology*, 551-558.
- Myers, G., 2002. Symbolic animals and the developing self. *Anthrozoös*, 15(1), 19-36.
- Foster, C.N., Barton, P.S. and Lindenmayer, D.B., 2014. Effects of large native herbivores on other animals. *Journal of Applied Ecology*, 51(4), 929-938.
- Næsset, E., 2002. Predicting forest stand characteristics with airborne scanning laser using a practical two-stage procedure and field data. *Remote Sensing of Environment*, 80(1), 88-99.
- Naidoo, R., Fisher, B., Manica, A. and Balmford, A., 2016. Estimating economic losses to tourism in Africa from the illegal killing of elephants. *Nature Communications*, 7(1), 1-9.
- Nair, K.S., 2007. Tropical forest insect pests: ecology, impact, and management. Cambridge University Press.
- Nakajima, S., Arimitsu, K. and Lattal, K.M., 2002. Estimation of animal intelligence by university students in Japan and the United States. *Anthrozoös*, 15(3), 194-205.
- Nasseri, N.A., McBrayer, L.D. and Schulte, B.A., 2011. The impact of tree modification by African elephant (*Loxodonta africana*) on herpetofaunal species richness in northern Tanzania. *African Journal of Ecology*, 49(2), 133-140.
- Navarro, A., Young, M., Allan, B., Carnell, P., Macreadie, P. and Ierodiaconou, D., 2020. The application of Unmanned Aerial Vehicles (UAVs) to estimate above-ground biomass of mangrove ecosystems. *Remote Sensing of Environment*, 242, 111747.
- N'Dri, A.B., Gignoux, J., Konaté, S., Dembélé, A., and Aïdara, D., 2011. Origin of trunk damage in West African savanna trees: the interaction of fire and termites, *Journal of Tropical Ecology*, 27, 269-278,
- Nelson, A., Bidwell, P. and Sillero-Zubiri, C., 2003. A review of human-elephant conflict management strategies. *People & Wildlife, A Wildlife Conservation Research Unit, Born Free Foundation Partnership*.
- Nelson, R., Krabill, W. and Tonelli, J., 1988. Estimating forest biomass and volume using airborne laser data. *Remote Sensing of Environment*, 24(2), 247-267.
- Neupane, D., Johnson, R.L. and Risch, T.S., 2017. How do land-use practices affect human—elephant conflict in nepal?. *Wildlife Biology*, 2017(4).
- Newsome, D., 2020. The collapse of tourism and its impact on wildlife tourism destinations.

*Journal of Tourism Futures*. 7 (3), 295-302.

- Ngama, S., Korte, L., Bindelle, J., Vermeulen, C., and Poulsen, J., 2016. How bees deter elephants: beehive trials with forest elephants (*Loxodonta africana cyclotis*) in Gabon. *PLoS One*, 11(5), 0155690.
- Nicholls, A. O., Viljoen, P. C., Knight, M. H., and van Jaarsveld, a. S., 1996. Evaluating population persistence of censused and unmanaged herbivore populations from the Kruger National Park, South Africa. *Biological Conservation*, 76(1), 57–67.
- Nkosi, S.E., Adam, E., Barrett, A.S. and Brown, L.R., 2019. A synopsis of field and remote sensing-based methods for studying African elephant (*Loxodonta Africana*) impact on woody vegetation in Africa. *Applied Ecology and Environmental Research*, 17(2), 4045-4066.
- Noordermeer, L., Gobakken, T., Næsset, E. and Bollandsås, O.M., 2021. Economic utility of 3D remote sensing data for estimation of site index in Nordic commercial forest inventories: a comparison of airborne laser scanning, digital aerial photogrammetry and conventional practices. *Scandinavian Journal of Forest Research*, 36(1), 55- 67.
- Noss, R.F., Noss, R.F. and Cooperrider, A., 1994. *Saving nature's legacy: protecting and restoring biodiversity*. Island Press.
- Novelli, M., Barnes, J.I. and Humavindu, M., 2006. The other side of the ecotourism coin: consumptive tourism in Southern Africa. *Journal of Ecotourism*, 5(1-2), 62-79.
- Ntumi, C. P., R. J. van Aarde, N. Fairall, and W. F. de Boer. 2005. Use of space and habitat by elephants (*Loxodonta africana*) in the Maputo Elephant Reserve, Mozambique. *South African Journal of Wildlife Research*, 35, 139–146.
- Nurminen, K., Karjalainen, M., Yu, X., Hyyppä, J. and Honkavaara, E., 2013. Performance of dense digital surface models based on image matching in the estimation of plot-level forest variables. *ISPRS Journal of Photogrammetry and Remote Sensing*, 83, 104-115.
- Nyakaana, S., Abe, E.L., Arctander, P. and Siegismund, H.R., 2001. DNA evidence for elephant social behaviour breakdown in Queen Elizabeth National Park, Uganda. *Animal Conservation*, 4(3), 231-237.
- Nyakaana, S., Abe, E.L., Arctander, P. and Siegismund, H.R., 2001, August. DNA evidence for elephant social behaviour breakdown in Queen Elizabeth National Park, Uganda. In *Animal Conservation forum*, 4(3), 231-237. Cambridge University Press.
- O'Connor, T. G., P. S. Goodman, and B. Clegg., 2007. A functional hypothesis of the threat

- of local extirpation of woody plant species by elephant in Africa. *Biological Conservation*, 136, 329–345.
- Oates, L. and Rees, P.A., 2013. The historical ecology of the large mammal populations of Ngorongoro Crater, Tanzania, east Africa. *Mammal Review*, 43(2), 124-141.
- Ogutu, J. O., Reid, R. S., Piepho, H. P., Hobbs, N. T., Rainy, M. E., Kruska, R. L., & Nyabenge, M., 2014. Large herbivore responses to surface water and land use in an East African savanna: implications for conservation and human-wildlife conflicts. *Biodiversity and Conservation*, 23(3), 573-596.
- Okello, M.M., 2005. Land use changes and human–wildlife conflicts in the Amboseli Area, Kenya. *Human Dimensions of Wildlife*, 10(1), 19-28.
- Okullo, P. and Moe, S.R., 2012. Large herbivores maintain termite-caused differences in herbaceous species diversity patterns. *Ecology*, 93(9), 2095-2103.
- Oliveras, I. and Malhi, Y., 2016. Many shades of green: the dynamic tropical forest–savannah transition zones. *Philosophical Transactions of the Royal Society B: Biological Sciences*, 371(1703), 20150308.
- Osborn, F.V. and Parker, G.E., 2003. Towards an integrated approach for reducing the conflict between elephants and people: a review of current research. *Oryx*, 37(1), 80-84.
- Owen-Smith, N. and Chafota, J., 2012. Selective feeding by a megaherbivore, the African elephant (*Loxodonta africana*). *Journal of Mammalogy*, 93(3), 698-705.
- Owen-Smith, N., 1988. *Megaherbivores: the influence of very large body size on ecology*. Cambridge university press.
- Owen-Smith, N., 1996. Ecological guidelines for waterpoints in extensive protected areas. *South African Journal of Wildlife Research*, 26(4), 107–112.
- Owen-Smith, N., Le Roux, E., and Macandza, V., 2013. Are relatively rare antelope narrowly selective feeders? A sable antelope and zebra comparison. *Journal of Zoology*, 291(3), 1–8.
- Owen-Smith, N., Slotow, R., Kerley, G., Van Aarde, R., and Page, B., 2006. A scientific perspective on the management of elephants in the Kruger National Park and elsewhere: elephant conservation. *South African Journal of Science*, 102(9), 389-394.
- Owen-Smith, R. N., 2002. Adaptive herbivore ecology: from resources to populations in variable environments. *Cambridge University Press*
- Owen-Smith, R.N., 1992. *Megaherbivores: the influence of very large body size on ecology*. Cambridge university press

- Palkopoulou, E., Lipson, M., Mallick, S., Nielsen, S., Rohland, N., Baleka, S., Karpinski, E., Ivancevic, A.M., To, T.H., Kortschak, R.D. and Raison, J.M., 2018. A comprehensive genomic history of extinct and living elephants. *Proceedings of the National Academy of Sciences*, 115(11), E2566-E2574.
- Palmer, R., 2005. Beyond the basics: post-basic education, training and poverty reduction in Ghana. *Post-Basic Education and Training Working Paper Series*, 4.
- Palmer, T.M. and Brody, A.K., 2013. Enough is enough: the effects of symbiotic ant abundance on herbivory, growth, and reproduction in an African acacia. *Ecology*, 94(3), 683-691.
- Parker, A. H., and Witkowski, E. T. F., 1999. Long-term impacts of abundant perennial water provision for game on herbaceous vegetation in a semi-arid African savanna woodland. *Journal of Arid Environments*, 41, 309–321.
- Parker, G.E. and Osborn, F.V., 2006. Investigating the potential for chilli *Capsicum* spp. to reduce human-wildlife conflict in Zimbabwe. *Oryx*, 40(3), 343-346.
- Parker, G.E., Osborn, F.V. and Hoarse, R.E., 2007. Human-elephant conflict mitigation: a training course for community-based approaches in Africa (Participant's Manual).
- Payne, K.B., Langbauer, W.R. and Thomas, E.M., 1986. Infrasonic calls of the Asian elephant (*Elephas maximus*). *Behavioral Ecology and Sociobiology*, 18(4), 297-301.
- Pelletier, J. and Wilde Astington, J., 2004. Action, consciousness and theory of mind: Children's ability to coordinate story characters' actions and thoughts. *Early Education and Development*, 15(1), 5-22.
- Pellew, R., 1983. The impacts of elephant, giraffe and fire upon the *Acacia tortilis* woodlands of the Serengeti. *African Journal of Ecology*, 21(1), 41-74.
- Pennisi, E., 2015. Africa's soil engineers: Termites.
- Perera, B.M.A.O., 2009. The human-elephant conflict: A review of current status and mitigation methods. *Gajah*, 30, 41-52.
- Périquet, S., Valeix, M., Loveridge, A. J., Madzikanda, H., Macdonald, D. W., & Fritz, H., 2010. Individual vigilance of African herbivores while drinking: the role of immediate predation risk and context. *Animal Behaviour*, 79, 665–671.
- Perkins, J. S., & Thomas, D. S. G., 1993. Land degradation and cattle ranching in the Kalahari desert of Botswana. *Land Degradation and Rehabilitation*, 4, 179–194.
- Persson, H.J. and Fransson, J.E., 2017. Comparison between TanDEM-X and ALS-based estimation of aboveground biomass and tree height in boreal forests. *Scandinavian Journal of Forest Research*, 32(4), 306-319.

- Peters, B.C., Creffield, J.W. and Eldridge, R.H., 2002. Lyctine (Coleoptera: Bostrichidae) pests of timber in Australia: a literature review and susceptibility testing protocol. *Australian Forestry*, 65(2), 107-119.
- Pinheiro, J., and Bates, D., 2000. Fitting linear mixed-effects models. *Mixed-effects models in S and S-PLUS*, 133-199.
- Pirie, T.J., Thomas, R.L. and Fellowes, M.D., 2017. Game fence presence and permeability influences the local movement and distribution of South African mammals. *African Zoology*, 52(4), 217-227.
- Plotnik, J.M., De Waal, F.B. and Reiss, D., 2006. Self-recognition in an Asian elephant. *Proceedings of the National Academy of Sciences*, 103(45), 17053-17057.
- Plotnik, J.M., de Waal, F.B., Moore III, D. and Reiss, D., 2010. Self-recognition in the Asian elephant and future directions for cognitive research with elephants in zoological settings. *Zoo Biology*, 29(2), 179-191.
- Plous, S., (1993). Psychological mechanisms in the human use of animals. *Journal of Social Issues*, 49(1), 11-52.
- Poole, J.H., 1989. Mate guarding, reproductive success and female choice in African elephants. *Animal Behaviour*, 37, 842-849.
- Popescu, S.C., Wynne, R.H. and Nelson, R.F., 2003. Measuring individual tree crown diameter with lidar and assessing its influence on estimating forest volume and biomass. *Canadian Journal of Remote Sensing*, 29(5), 564-577.
- Pozo, R., Coulson, T., McCulloch, G., Stronza, A., and Songhurst, A., 2019. Chilli-briquettes modify the temporal behaviour of elephants, but not their numbers. *Oryx*, 53(1), 100-108.
- Pringle, R.M., 2008. Elephants as agents of habitat creation for small vertebrates at the patch scale. *Ecology*, 89(1), 26-33.
- Pringle, R.M., Kimuyu, D.M., Sensenig, R.L., Palmer, T.M., Riginos, C., Veblen, K.E. and Young, T.P., 2015. Synergistic effects of fire and elephants on arboreal animals in an African savanna. *Journal of Animal Ecology*, 84(6), 1637-1645.
- R Core Team (2014). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>
- Raihan Sarker, A.H.M. and Røskaft, E., 2010. Human–wildlife conflicts and management options in Bangladesh, with special reference to Asian elephants (*Elephas maximus*). *International Journal of Biodiversity Science, Ecosystem Services and Management*, 6(3-

4), 164-175.

- Rakotomamonjy, S.N., Jones, J.P.G., Razafimanahaka, J.H., Ramamonjisoa, B. and Williams, S.J., 2015. The effects of environmental education on children's and parents' knowledge and attitudes towards lemurs in rural Madagascar. *Animal Conservation*, 18(2), 157-166.
- Rausher, M. 1992. The measurement of selection on quantitative traits: biases due to environmental covariances between traits and fitness. *Evolution*, 46, 616-626.
- Redfern, J. V., Grant, R., Biggs, H., and Getz, W. M., 2003. Surface-Water Constraints on Herbivore Foraging in the Kruger National Park, South Africa. *Ecology*, 84(8), 2092–2107.
- Reitberger, J., Schnörr, C., Krzystek, P. and Stilla, U., 2009. 3D segmentation of single trees exploiting full waveform LIDAR data. *ISPRS Journal of Photogrammetry and Remote Sensing*, 64(6), 561-574.
- Rensch, B., 1957. The intelligence of elephants. *Scientific American*, 196(2), 44-49.
- Richards, S.A., 2005. Testing ecological theory using the information-theoretic approach: examples and cautionary results. *Ecology*, 86(10), 2805-2814.
- Richards, S.A., Whittingham, M.J. and Stephens, P.A., 2011. Model selection and model averaging in behavioural ecology: the utility of the IT-AIC framework. *Behavioral Ecology and Sociobiology*, 65(1), 77-89.
- Riginos, C., and Grace, J. B., 2008. Savanna tree density, herbivores, and the herbaceous community: Bottom-up vs. top-down effects. *Ecology*, 89(8), 2228–2238.
- Ripple, W.J., Newsome, T.M., Wolf, C., Dirzo, R., Everatt, K.T., Galetti, M., Hayward, M.W., Kerley, G.I., Levi, T., Lindsey, P.A. and Macdonald, D.W., 2015. Collapse of the world's largest herbivores. *Science Advances*, 1(4).
- Roca, A.L., Georgiadis, N. and O'Brien, S.J., 2007. Cyto-nuclear genomic dissociation and the African elephant species question. *Quaternary International*, 169, 4-16.
- Roca, A.L., Ishida, Y., Brandt, A.L., Benjamin, N.R., Zhao, K. and Georgiadis, N.J., 2015. Elephant natural history: a genomic perspective. *Annual Review of Animal Biosciences*, 3(1), 139-167.
- Rohner, B., Waldner, P., Lischke, H., Ferretti, M. and Thürig, E., 2018. Predicting individual- tree growth of central European tree species as a function of site, stand, management, nutrient, and climate effects. *European Journal of Forest Research*, 137(1), 29-44.
- Roussel, J.R. and Auty, D., 2017. lidR: Airborne LiDAR Data Manipulation and Visualization

for Forestry Applications. R Package Version 1.2. 0. *Cit. on*, 32.

Roussel, J.R., Auty, D., Coops, N.C., Tompalski, P., Goodbody, T.R., Meador, A.S., Bourdon, J.F., de Boissieu, F. and Achim, A., 2020. lidR: An R package for analysis of Airborne Laser Scanning (ALS) data. *Remote Sensing of Environment*, 251, 112061.

- Ruess, R.W. and Halter, F.L., 1990. The impact of large herbivores on the Seronera woodlands, Serengeti National Park, Tanzania. *African Journal of Ecology*, 28(4), 259-275.
- Russell, J. and Ward, D., 2014. Vegetation change in northern KwaZulu-Natal since the Anglo-Zulu War of 1879: local or global drivers?. *African Journal of Range and Forage Science*, 31(2), 89-105.
- Rutherford, M.C. and Westfall, R.H., 1994. Biomes of southern Africa: an objective categorization. National Botanical Institute.
- SANParks., 2012. Elephant management plan. Kruger National Park. 2013-2022. SANParks, Skukuza, South Africa.  
[https://www.sanparks.org/assets/docs/parks\\_kruger/elephants/knp- elephantmanagement-plan.pdf](https://www.sanparks.org/assets/docs/parks_kruger/elephants/knp- elephantmanagement-plan.pdf)
- Santiapillai, C., 1997. The Asian elephant conservation: a global strategy. *Gajah*, 18, 21- 39.
- Schippers, P., van Teeffelen, A. J. A., Verboom, J., Vos, C. C., Kramer, K., and WallisDeVries, M. F. 2014. The impact of large herbivores on woodland-grassland dynamics in fragmented landscapes: The role of spatial configuration and disturbance. *Ecological Complexity*, 17(1), 20–31.
- Scholtz, R., Kiker, G.A., Smit, I.P.J. and Venter, F.J., 2014. Identifying drivers that influence the spatial distribution of woody vegetation in Kruger National Park, South Africa. *Ecosphere*, 5(6), 1-12.
- Scogings, P., Johansson, T., Hjältén, J., and Kruger, J., 2012. Responses of woody vegetation to exclusion of large herbivores in semi-arid savannas. *Austral Ecology*, 37(1), 56-66.
- Seloana, M.Q., Kruger, J.W., Potgieter, M.J. and Jordaan, J.J., 2017. Elephant damage to *Sclerocarya birrea* on different landscapes. *International Journal of Biodiversity and Conservation*, 9(4), 97-106.
- Senzota, R. B. M., & Mtahko, G., 1990. Effect on wildlife of a waterhole in Mikumi National Park, Tanzania. *African Journal of Ecology*, 28, 147–151.
- Seydack, A. H., Grant, C. C., Smit, I. P., Vermeulen, W. J., Baard, J., and Zambatis, N., 2012. Large herbivore population performance and climate in a South African semi-arid savanna. *Koedoe*, 54(1), 1–20.
- Shafer, C.L., 1995. Values and shortcomings of small reserves. *BioScience*, 45(2), 80-88.
- Shannon, G., Druce, D., Page, B., Eckhardt, H., Grant, R., and Slotow, R., 2008. The

- utilization of large savanna trees by elephant in southern Kruger National Park. *Journal of Tropical Ecology*, 281-289.
- Shannon, G., Matthews, W. S., Page, B. R., Parker, G. E., and Smith, R. J., 2009. The affects of artificial water availability on large herbivore ranging patterns in savanna habitats: a new approach based on modelling elephant path distributions. *Diversity and Distributions*, 15(5), 776–783.
- Shannon, G., Thaker, M., Vanak, A.T., Page, B.R., Grant, R. and Slotow, R., 2011. Relative impacts of elephant and fire on large trees in a savanna ecosystem. *Ecosystems*, 14(8), 1372-1381.
- Sherry, B. Y., 1989. Aspects of the biology of the elephant in the middle Shire valley, Southern Malawi. MSc Thesis, University of Malawi, Malawi.
- Sieber, J. E. 1986. Students' and scientists' attitudes on animal research. *The American Biology Teacher*, 48(2), 85–91.
- Simms, C., 2009. *The utilisation of satellite images for the detection of elephant induced vegetation change patterns* (Doctoral dissertation, University of South Africa).
- Sirami, C., Seymour, C., Midgley, G. and Barnard, P., 2009. The impact of shrub encroachment on savanna bird diversity from local to regional scale. *Diversity and Distributions*, 15(6), 948-957.
- Sirost, E., Renaud, P. C., and Pays, O., 2016. How competition and predation shape patterns of waterhole use by herbivores in arid ecosystems. *Animal Behaviour*, 118, 19–26.
- Sitati, N.W. and Ipara, H., 2012. Indigenous ecological knowledge of a human-elephant interaction in Transmara district, Kenya: Implications for research and management. *Advances in Anthropology*, 2(3),107.
- Sitati, N.W. and Walpole, M.J., 2006. Assessing farm-based measures for mitigating human-elephant conflict in Transmara District, Kenya. *Oryx*, 40(3), 279-286.
- Skarpe, C., 1990. Shrub Layer Dynamics Under Different Herbivore Densities in an Arid Savanna, Botswana. *Journal of Applied Ecology*, 27(3), 873–885.
- Skarpe, C., 1992. Dynamics of savanna ecosystems. *Journal of Vegetation Science*, 3(3), 293-300.
- Skarpe, C., Aarrestad, P.A., Andreassen, H.P., Dhillion, S.S., Dimakatso, T., du Toit, J.T., Duncan, J.H., Hytteborn, H., Makhabu, S., Mari, M. and Marokane, W., 2004. The

- return of the giants: ecological effects of an increasing elephant population. *AMBIO: A Journal of the Human Environment*, 33(6), 276-282.
- Slotow, R., Garai, M., Reilly, B., Page, B., and Carr, R., 2005. Population dynamics of elephants re-introduced to small, fenced reserves in South Africa. *South African Journal of Wildlife Research*, 35(1), 23-32.
- Slotow, R., Garai, M.E., Reilly, B., Page, B. and Carr, R.D., 2005. Population dynamics of elephants re-introduced to small, fenced reserves in South Africa. *South African Journal of Wildlife Research-24-month delayed open access*, 35(1), 23-32.
- Smallie, J.J. and O'connor, T.G., 2000. Elephant utilization of *Colophospermum mopane*: possible benefits of hedging. *African Journal of Ecology*, 38(4), 352-359.
- Smit, I. P. J., and Grant, C. C., 2009. Managing surface-water in a large semi-arid savanna park: Effects on grazer distribution patterns. *Journal for Nature Conservation*, 17(2), 61–71.
- Smit, I. P. J., Grant, C. C., and Devereux, B. J., 2007. Do artificial waterholes influence the way herbivores use the landscape? Herbivore distribution patterns around rivers and artificial surface water sources in a large African savanna park. *Biological Conservation*, 136, 85–99.
- Smit, I.P.J., Grant, C.C. and Whyte, I.J., 2007. Landscape-scale sexual segregation in the dry season distribution and resource utilization of elephants in Kruger National Park, South Africa. *Diversity and Distributions*, 13(2), 225-236.
- Soule, M.E. and Simberloff, D., 1986. What do genetics and ecology tell us about the design of nature reserves?. *Biological Conservation*, 35(1), 19-40.
- Soulé, M.E. and Terborgh, J., 1999. Conserving nature at regional and continental scales—a scientific program for North America. *BioScience*, 49(10), 809-817.
- Spies, K. S., 2015. Ecology of impala (*Aepyceros melampus*) and waterbuck (*Kobus ellipsiprymnus*) in Majete Wildlife Reserve, Malawi. MSc Thesis, University of Stellenbosch, South Africa.
- Ssali, F., Sheil, D., Nkurunungi, J., 2013. How selective are elephants as agents of forest tree damage in Bwindi Impenetrable National Park, Uganda? *African Journal of Ecology*, 51(1), 55-65.
- Staub, C. G., M. W. Binford, and F. R. Stevens. 2013. Elephant herbivory in Majete Wildlife Reserve, Malawi. *African Journal of Ecology*, 51, 536–543.

- Staub, C.G., Binford, M.W. and Stevens, F.R., 2013. Elephant herbivory in Majete wildlife reserve, Malawi. *African Journal of Ecology*, 51(4), 536-543.
- Stephenson, P.J. and Ntiamoa-Baidu, Y., 2010. Conservation planning for a widespread, threatened species: WWF and the African elephant *Loxodonta africana*. *Oryx*, 44(2), 194-204.
- Stepper, C., Straub, C. and Pretzsch, H., 2015. Using semi-global matching point clouds to estimate growing stock at the plot and stand levels: Application for a broadleaf-dominated forest in central Europe. *Canadian Journal of Forest Research*, 45(1), 111-123.
- Stereńczak, K., Kraszewski, B., Mielcarek, M., Piasecka, Ż., Lisiewicz, M. and Heurich, M., 2020. Mapping individual trees with airborne laser scanning data in an European lowland forest using a self-calibration algorithm. *International Journal of Applied Earth Observation and Geoinformation*, 93, 102191.
- Stiles, D., 2004. The ivory trade and elephant conservation. *Environmental Conservation*, 31(4), 309-321.
- Stinchcombe, J.R., Rutter, M.T., Burdick, D.S., Tiffin, P., Rausher, M.D. and Mauricio, R., 2002. Testing for environmentally induced bias in phenotypic estimates of natural selection: theory and practice. *The American Naturalist*, 160(4), 511-523.
- Störmer, N., Weaver, L.C., Stuart-Hill, G., Diggle, R.W. and Naidoo, R., 2019. Investigating the effects of community-based conservation on attitudes towards wildlife in Namibia. *Biological Conservation*, 233, 193-200.
- Straub, C., Stepper, C., Seitz, R. and Waser, L.T., 2013. Potential of UltraCamX stereo images for estimating timber volume and basal area at the plot level in mixed European forests. *Canadian Journal of Forest Research*, 43(8), 731-741.
- Stretch, A., Duffy, K., Van Os, R., Vos, S., Van Aarde, J., and Elish, G., 2002. Estimating impact of reintroduced elephant on trees in a small reserve. *South African Journal of Wildlife Research*, 32(1), 23-29.
- Stuart-Hill, G.C., 1992. Effects of elephants and goats on the Kaffrarian succulent thicket of the eastern Cape, South Africa. *Journal of Applied Ecology*, 699-710.
- Styles, C., and Skinner, J., 2000. The influence of large mammalian herbivores on growth form and utilization of mopane trees, *Colophospermum mopane*, in Botswana's Northern Tuli Game Reserve. *African Journal of Ecology*, 38(2), 95-101.

- Suárez, J.C., Ontiveros, C., Smith, S. and Snape, S., 2005. Use of airborne LiDAR and aerial photography in the estimation of individual tree heights in forestry. *Computers and Geosciences*, 31(2), 253-262.
- Sungirai, M., and Ngwenya, M., 2016. An Investigation into the Efficiency of Utilization of Artificial Game Water Supplies by Wildlife Species in the North Eastern Kalahari Region of Hwange National Park in Zimbabwe. *Applied Ecology and Environmental Sciences*, 4(1), 7–14.
- Sutherland, K., Ndlovu, M., and Perez-Rodriguez, A., 2018. Use of artificial waterholes by animals in the southern region of the Kruger National Park, South Africa. *African Journal of Wildlife Research*, 48(2), 1–14.
- Swanagan, J.S., 2000. Factors influencing zoo visitors' conservation attitudes and behavior. *The Journal of Environmental Education*, 31(4), 26-31.
- Symonds, M.R. and Moussalli, A., 2011. A brief guide to model selection, multimodel inference and model averaging in behavioural ecology using Akaike's information criterion. *Behavioral Ecology and Sociobiology*, 65(1),13-21.
- Takooshian, H. 1988. Opinions on animal research: Scientists vs. the public. *PsyETA Bulletin*, 7, 5–9
- Tao, S., Wu, F., Guo, Q., Wang, Y., Li, W., Xue, B., Hu, X., Li, P., Tian, D., Li, C. and Yao, H., 2015. Segmenting tree crowns from terrestrial and mobile LiDAR data by exploring ecological theories. *ISPRS Journal of Photogrammetry and Remote Sensing*, 110, 66-76.
- Taruvinga, A. and Mushunje, A., 2014. Society's perceptions of African elephants and their relative influence towards the conservation of elephants. *APCBEE procedia*, 10, 299-304.
- Taylor, R.D. 2009. Community based natural resource management in Zimbabwe: the experience of CAMPFIRE. *Biological Conservation*, 18(10): 2563–2583.
- Tefempa, H. B., Ngassam, P., Nkongmeneck, B. A., Mapongmetsem, P. M., & Gubbuk, H., 2008. Behaviour of mammals around artificial waterholes in the Waza National Park (Cameroon). *Mediterranean Agricultural Sciences*, 21(1), 7–13.
- Thompson, K.A. and Johnson, M.T.J. 2016. Antiherbivore defenses alter natural selection on plant reproductive traits. *Evolution*, 70: 796-810.
- Thomson, P., 1975. The role of elephants, fire and other agents in the decline of a *Brachystegia boehmii* woodland. *South African Journal of Wildlife Research*, 5(1), 11-18.

- Thouless, C., Dublin, H.T., Blanc, J., Skinner, D., Daniel, T., Taylor, R., Maisels, F., Frederick, H. and Bouché, P., 2016. African elephant status report 2016. *An update from the African Elephant Database*.
- Thrash, I., and Derry, J. F., 1999. The nature and modelling of piospheres: A review. *Koedoe*, 42(2), 73–94.
- Thrash, I., 2000. Determinants of the extent of indigenous large herbivore impact on herbaceous vegetation at watering points in the north-eastern lowveld, South Africa. *Journal of Arid Environments*, 44, 61–72.
- Thrash, I., Nel, P. J., Theron, G. K., and Bothma J. du P., 1991. The impact of the provision of water for game on the woody vegetation around a dam in the Kruger National Park. *Koedoe*, 34(2), 131–148.
- Thrash, I., Theron, G. K., and Bothma, J. du P., 1993. Impact of water provision on herbaceous community composition in the Kruger National Park, South Africa. *African Journal of Range and Forage Science*, 10(1), 31–35.
- Tolsma, D.J., Ernst, W. H. O., & Verwey, R. A., 1987. Nutrients in soil and vegetation around two artificial water points in eastern Botswana. *Journal of Applied Ecology*, 24, 991– 1000.
- Tompalski, P., White, J.C., Coops, N.C. and Wulder, M.A., 2019. Quantifying the contribution of spectral metrics derived from digital aerial photogrammetry to area-based models of forest inventory attributes. *Remote Sensing of Environment*, 234, 111434.
- Toomey, A.H., Knight, A.T. and Barlow, J., 2017. Navigating the space between research and implementation in conservation. *Conservation Letters*, 10(5), 619-625.
- Trollope, W.S.W., Trollope, L.A., Biggs, H.C., Pienaar, D. and Potgieter, A.L.F., 1998. Long-term changes in the woody vegetation of the Kruger National Park, with special reference to the effects of elephants and fire. *Koedoe*, 41(2), 103-112.
- Underwood, F.M., Burn, R.W. and Milliken, T., 2013. Dissecting the illegal ivory trade: an analysis of ivory seizures data. *PloS one*, 8(10), p.e76539.
- Valeix, M., 2011. Temporal dynamics of dry-season water-hole use by large African herbivores in two years of contrasting rainfall in Hwange National Park, Zimbabwe. *Journal of Tropical Ecology*, 27, 163–170.
- Valeix, M., Fritz, H., Loveridge, A. J., Davidson, Z., Hunt, J. E., Murindagomo, F., & Macdonald, D. W., 2009. Does the risk of encountering lions influence African herbivore behaviour at waterholes? *Behavioural Ecology and Socio-biology*, 63, 1483– 1494.
- Van der Plas, F., Howison, R., Reinders, J., Fokkema, W. and Olf, H., 2013. Functional traits

- of trees on and off termite mounds: understanding the origin of biotically-driven heterogeneity in savannas. *Journal of Vegetation Science*, 24(2), 227-238.
- van Hoven, W., 2015. Private game reserves in southern Africa. In *Institutional arrangements for conservation, development and tourism in eastern and southern Africa* 101-118. Springer, Dordrecht.
- Van Wyk, P. and Fairall, N., 1969. The influence of the African elephant on the vegetation of the Kruger National Park. *Koedoe*, 12(1), 57-89.
- Vanak, A., Thaker, M. and Slotow, R., 2010. Do fences create an edge-effect on the movement patterns of a highly mobile mega-herbivore?. *Biological Conservation*, 143(11), 2631-2637.
- Vanak, A.T., Shannon, G., Thaker, M., Page, B., Grant, R. and Slotow, R., 2012. Biocomplexity in large tree mortality: interactions between elephant, fire and landscape in an African savanna. *Ecography*, 35(4), 315-321.
- Van Valkenburgh, P., Cushman, K.C., Butters, L.J.C., Vega, C.R., Roberts, C.B., Kepler, C. and Kellner, J., 2020. Lasers without lost cities: Using drone lidar to capture architectural complexity at Kuelap, Amazonas, Peru. *Journal of Field Archaeology*, 45, S75-S88.
- Vastaranta, M., Wulder, M.A., White, J.C., Pekkarinen, A., Tuominen, S., Ginzler, C., Kankare, V., Holopainen, M., Hyypä, J. and Hyypä, H., 2013. Airborne laser scanning and digital stereo imagery measures of forest structure: Comparative results and implications to forest mapping and inventory update. *Canadian Journal of Remote Sensing*, 39(5), 382-395.
- Vauhkonen, J., Ene, L., Gupta, S., Heinzl, J., Holmgren, J., Pitkänen, J., Solberg, S., Wang, Y., Weinacker, H., Hauglin, K.M. and Lien, V., 2012. Comparative testing of single-tree detection algorithms under different types of forest. *Forestry*, 85(1), 27-40.
- Vesey-FitzGerald, D.F., 1973. Animal impact on vegetation and plant succession in Lake Manyara National Park, Tanzania. *Oikos*, 314-324.
- Vigne, L. and Martin, E., 2017. Decline in the legal ivory trade in China in anticipation of a ban. *Nairobi: Save The Elephants*.
- Vogel, S.M., Henley, M.D., Rode, S.C., van de Vyver, D., Meares, K.F., Simmons, G. and de Boer, W.F., 2014. Elephant (*Loxodonta africana*) impact on trees used by nesting vultures and raptors in South Africa. *African Journal of Ecology*, 52(4), 458-465.
- Waithaka, J., 2012, January. Historical factors that shaped wildlife conservation in Kenya. In *The George Wright Forum* 29(1) 21-29. George Wright Society.

- Walker, S.H., 1976. An approach to the monitoring of changes in the composition and utilization of woodland and savanna vegetation. *South African Journal of Wildlife Research-24-month delayed open access*, 6(1), 1-32.
- Wargo, P.M., 1996. Consequences of environmental stress on oak: predisposition to pathogens. *Annales des sciences forestières*, (53), 359-368.
- Weaver, S., 1995. *Habitat utilisation by selected herbivores in the Klaserie Private Nature Reserve, South Africa*. M.Sc. thesis, University of Pretoria, Pretoria.
- Weir, J. S., 1971. The effect of creating additional water supplies in a Central African National Park. In E. Duffey & A. S. Watt (Eds.), *The scientific management of animal and plant communities for conservation*. 367–385. *Oxford: Blackwell Scientific Publications*.
- Weir, J., and Davison, E., 1965. Daily Occurrence of African Game Animals at Water Holes During Dry Weather. *Zoologica Africana*, 1(2), 353–368.
- Werner, P.A., Prior, L.D., Forner, J., 2008. Growth and survival of termite-piped *Eucalyptus tetradonta* and *E. miniata* in northern Australia: Implications for harvest of trees for didgeridoos. *Forest Ecology and Management* (256), 328–334.
- West, S.E., 2015. Understanding participant and practitioner outcomes of environmental education. *Environmental Education Research*, 21(1), 45-60.
- Western, D. 1989. The ecological value of elephants: a keystone role In Africa's ecosystems. In: *Ivory Trade and the future of the African elephant*. Vol.2. A report by the Ivory Trade Review Group to CITES.
- Western, D. and Lindsay, W.K., 1984. Seasonal herd dynamics of a savanna elephant population. *African Journal of Ecology*, 22(4), 229-244.
- Western, D., 1989, The ecological role of elephants in Africa. 42-45.
- Western, D., 1975. Water availability and its influence on the structure and dynamics of a savanna large mammal community. *East African Wildlife Journal*, 13, 265–286.
- Western, D., and Maitumo, D., 2004. Woodland loss and restoration in a savanna park: a 20-year experiment. *African Journal of Ecology*, 42(2), 111-121.
- Western, D., Waithaka, J. and Kamanga, J., 2015. Finding space for wildlife beyond national parks and reducing conflict through community-based conservation: the Kenya experience. *Parks*, 21(1), 51-62.
- White, A.M., and Goodman, P.S., 2009. Differences in woody vegetation are unrelated to use by African elephants (*Loxodonta africana*) in Mkhuzi Game Reserve, South Africa. *African Journal of Ecology*, (48), 215– 223.

- White, J.C., Stepper, C., Tompalski, P., Coops, N.C. and Wulder, M.A., 2015. Comparing ALS and image-based point cloud metrics and modelled forest inventory attributes in a complex coastal forest environment. *Forests*, 6(10), 3704-3732.
- Whitehouse, A.M. and Schoeman, D.S., 2003. Ranging behaviour of elephants within a small, fenced area in Addo Elephant National Park, South Africa. *African Zoology*, 38(1), 95-108.
- Whyte, I., van Aarde, R., Pimm, S., 2003. Kruger's elephant population: its size and consequences for ecosystem heterogeneity. *The Kruger experience: ecology and management of savanna heterogeneity*, 332-348.
- Wienand, J. J., 2013. Woody vegetation change and elephant water point use in Majete Wildlife Reserve: implications for water management strategies. MSc Thesis, University of Stellenbosch, South Africa.
- Wigley, B.J., Bond, W.J. and Hoffman, M.T., 2010. Thicket expansion in a South African savanna under divergent land use: local vs. global drivers?. *Global Change Biology*, 16(3), 964-976.
- Wigley, B.J., Coetsee, C., Kruger, L.M., Ratnam, J. and Sankaran, M., 2019. Ants, fire, and bark traits affect how African savanna trees recover following damage. *Biotropica*, 51(5), 682-691.
- Wittemyer, G., 2011. Effects of economic downturns on mortality of wild African elephants. *Conservation Biology*, 25(5), 1002-1009.
- Wittemyer, G., Northrup, J.M., Blanc, J., Douglas-Hamilton, I., Omondi, P. and Burnham, K.P., 2014. Illegal killing for ivory drives global decline in African elephants. *Proceedings of the National Academy of Sciences*, 111(36), 13117-13121.
- Wittemyer, G., Okello, J.B., Rasmussen, H.B., Arctander, P., Nyakaana, S., Douglas-Hamilton, I. and Siegismund, H.R., 2009. Where sociality and relatedness diverge: the genetic basis for hierarchical social organization in African elephants. *Proceedings of the Royal Society B: Biological Sciences*, 276(1672), 3513-3521.
- Wolf, P.R., Dewitt, B.A. and Wilkinson, B.E., 2014. *Elements of Photogrammetry with Applications in GIS*. McGraw-Hill Education.
- Woolley, L., Page, B. and Slotow, R., 2011. Foraging strategy within African elephant familyunits: why body size matters. *Biotropica*, 43(4), 489-495.
- Woolley, L.A., Millspaugh, J.J., Woods, R.J., Van Rensburg, S.J., Mackey, R.L., Page, B. and Slotow, R., 2008. Population and individual elephant response to a catastrophic fire

- in Pilanesberg National Park. *PLoS One*, 3(9), 3233.
- Worton, B., 1989. Kernel methods for estimating the utilization distribution in home range studies. *Ecology*, 70, 164-168
- Worton, B.J., 1989. Kernel methods for estimating the utilization distribution in home-range studies. *Ecology Society of America*, 70, 164-168.
- Wulder, M.A., Loveland, T.R., Roy, D.P., Crawford, C.J., Masek, J.G., Woodcock, C.E., Allen, R.G., Anderson, M.C., Belward, A.S., Cohen, W.B. and Dwyer, J., 2019. Current status of Landsat program, science, and applications. *Remote Sensing of Environment*, 225, 127-147.
- Wulder, M.A., White, J.C., Nelson, R.F., Næsset, E., Ørka, H.O., Coops, N.C., Hilker, T., Bater, C.W. and Gobakken, T., 2012. Lidar sampling for large-area forest characterization: A review. *Remote Sensing of Environment*, 121, 196-209.
- Xie, Y., Sha, Z. and Yu, M., 2008. Remote sensing imagery in vegetation mapping: a review. *Journal of Plant Ecology*, 1(1), 9-23.
- Yao, W., Krzystek, P. and Heurich, M., 2012. Tree species classification and estimation of stem volume and DBH based on single tree extraction by exploiting airborne full-waveform LiDAR data. *Remote Sensing of Environment*, 123, 368-380.
- Young, E., 1972. The value of waterhole counts in estimating wild animal populations. *Journal of the South African Wildlife Management Association*, 2(1), 22–23.
- Young, K. D., Ferreira, S. M., and Van Aarde, R. J., 2009. The influence of increasing population size and vegetation productivity on elephant distribution in the Kruger National Park. *Austral Ecology*, 34(3), 329–342.
- Young, K., Ferreira, S., and Van Aarde, R., 2009. Elephant spatial use in wet and dry savannas of southern Africa. *Journal of Zoology*, 278(3), 189-205.
- Zarco-Tejada, P.J., Diaz-Varela, R., Angileri, V. and Loudjani, P., 2014. Tree height quantification using very high-resolution imagery acquired from an unmanned aerial vehicle (UAV) and automatic 3D photo-reconstruction methods. *European Journal of Agronomy*, 55, 89-99.
- Zhao, Kaiguang, Sorin Popescu, and Ross Nelson. "Lidar remote sensing of forest biomass: A scale-invariant estimation approach using airborne lasers." *Remote Sensing of Environment*, 113(1), 182-196.
- Zuur, A., Ieno, E.N., Walker, N., Saveliev, A.A. and Smith, G.M., 2009. Mixed effects models and extensions in ecology with R. Springer Science and Business Media.
- Zuur, A.F., Ieno, E.N. and Elphick, C.S., 2010. A protocol for data exploration to avoid common

statistical problems. *Methods in Ecology and Evolution*, 1(1), 3-14.

Zvidzai, M., Murwira, A., Caron, A., and Wichtatitsky, G., 2013. Waterhole use patterns at the wildlife/livestock interface in a semi-arid savanna of Southern Africa. *International Journal of Development and Sustainability*, 2(2), 455–471.

## Appendices

### Appendix 1.

**Table A.1** The number of species (n) found within the area (Karongwe Private Game Reserve)

with common name, family and genus

Species	Common name	Family	Genus	Count (n)
<i>Acacia nigrescens</i>	Knob Thorn	<i>Fabaceae</i>	<i>Senegalia</i>	240
<i>Sclerocarya birrea</i>	Marula	<i>Anacardiaceae</i>	<i>Sclerocarya</i>	153
<i>Commiphora mollis</i>	Velvet-Leaved Corkwood	<i>Burseraceae</i>	<i>Commiphora</i>	107
<i>Combretum apiculatum</i>	Red bush willow	<i>Combretaceae</i>	<i>Combretum</i>	93
<i>Combretum imberbe</i>	Leadwood	<i>Combretaceae</i>	<i>Combretum</i>	43
<i>Combretum hereroense</i>	Russet bush willow	<i>Combretaceae</i>	<i>Combretum</i>	21
<i>Diospyros mespiliformis</i>	Jackalberry	<i>Ebenaceae</i>	<i>Diospyros</i>	17
<i>Acacia tortilis</i>	Umbrella thorn	<i>Fabaceae</i>	<i>Vachellia</i>	16
<i>Acacia nilotica</i>	Gum arabic	<i>Fabaceae</i>	<i>Vachellia</i>	13
<i>Bolusanthus speciosus</i>	Wisteria	<i>Fabaceae</i>	<i>Bolusanthus</i>	11
<i>Lannea schweinfurthii</i>	False marula	<i>Anacardiaceae</i>	<i>Lannea</i>	11
<i>Philenoptera violacea</i>	Apple leaf	<i>Fabaceae</i>	<i>Philenoptera</i>	11
<i>Peltophorum africanum</i>	Weeping wattle	<i>Fabaceae</i>	<i>Peltophorum</i>	10
<i>Searsia pyroides</i>	Common currant-rhus	<i>Anacardiaceae</i>	<i>Searsia</i>	6
<i>Spirostachys africana</i>	Tamboti	<i>Euphorbiaceae</i>	<i>Spirostachys</i>	6

<i>Combretum collinum</i>	Weeping bush willow	<i>Combretaceae</i>	<i>Combretum</i>	4
<i>Olea europaea</i>	European olive	<i>Oleaceae</i>	<i>Olea</i>	4
<i>Commiphora glandulosa</i>	Tall firethorn corkwood	<i>Burseraceae</i>	<i>Commiphora</i>	3
<i>Schotia brachypetala</i>	Weeping boer-bean	<i>Fabaceae</i>	<i>Schotia</i>	3
<i>Ziziphus mucronata</i>	Buffalo thorn	<i>Rhamnaceae</i>	<i>Ziziphus</i>	3
<i>Acacia gerrardii</i>	Red Thorn	<i>Fabaceae</i>	<i>Acacia</i>	2
<i>Balanites maughamii</i>	Torchwood	<i>Zygophyllaceae</i>	<i>Balanites</i>	2
<i>Combretum molle</i>	Velvet bushwillow	<i>Combretaceae</i>	<i>Combretum</i>	2
<i>Ozoroa paniculosa</i>	Common resin tree	<i>Anacardiaceae</i>	<i>Ozoroa</i>	2
<i>Strychnos spinosa</i>	Green monkey orange	<i>Loganiaceae</i>	<i>Strychnos</i>	2
<i>Acacia erubescens</i>	Blue thorn	<i>Fabaceae</i>	<i>Vachellia</i>	1
<i>Acacia robusta</i>	Splendid thorn	<i>Fabaceae</i>	<i>Vachellia</i>	1
<i>Albizia harveyi</i>	Common false thorn	<i>Fabaceae</i>	<i>Albizia</i>	1
<i>Dovyalis caffra</i>	Kei-apple	<i>Salicaceae</i>	<i>Dovyalis</i>	1
<i>Gymnosporia buxifolia</i>	Common spike-thorn	<i>Celastraceae</i>	<i>Gymnosporia</i>	1
<i>Strychnos madagascariensis</i>	Black monkey orange	<i>Loganiaceae</i>	<i>Strychnos</i>	1
<i>Terminalia prunioides</i>	Purple-pod Clusterleaf	<i>Combretaceae</i>	<i>Terminalia</i>	1

**Appendix 2.**

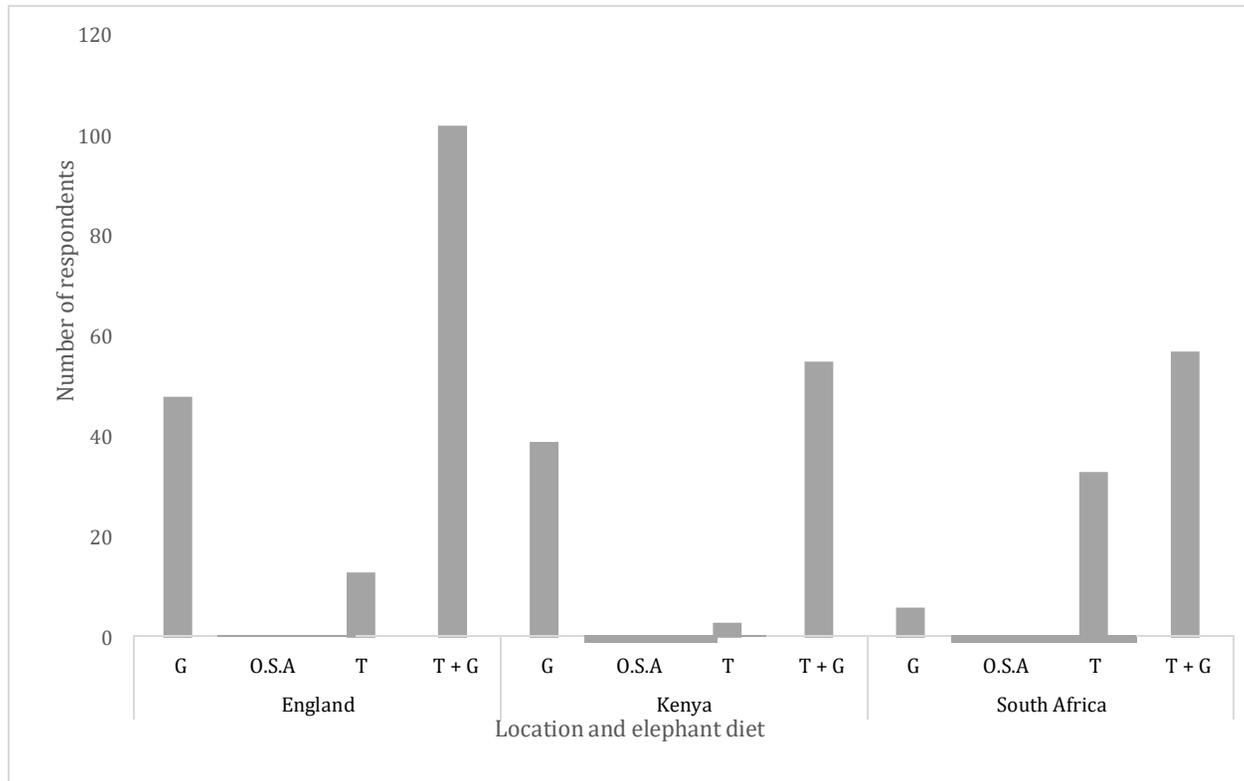
**Table A.2.1** Contingency table showing aggregated career choice data for male and female students within each country

Country	Gender	Ranking	<i>Career type</i>							
			Teacher	Doctor	Veterinarian	Artist	Musician	Athlete	Wildlife Guide	Business manager
England	<i>Male</i>	Most favourite 1	7	14	1	6	2	18	3	14
		2	10	14	6	6	5	9	5	10
		3	12	10	11	6	5	8	6	7
		4	9	11	6	11	8	7	9	5
		5	6	5	19	10	7	5	9	5
		6	4	6	6	8	17	5	10	8
		7	5	2	10	8	9	5	18	7
		Least favourite 8	12	3	6	10	12	8	5	9
England	<i>Female</i>	Most favourite 1	10	34	8	13	9	8	10	6

		2	21	13	13	10	10	2	15	14
		3	21	17	17	12	10	5	9	7
		4	11	10	20	14	9	12	12	10
		5	8	11	15	18	11	9	16	10
		6	13	5	8	14	12	17	10	19
		7	5	4	9	8	16	25	19	12
		Least favourite 8	9	4	8	9	21	20	7	20
Kenya	<i>Male</i>	Most favourite 1	1	4	3	2	10	2	6	6
		2	2	6	2	8	3	2	7	5
		3	3	4	3	4	7	2	3	11
		4	5	7	2	9	1	4	5	2
		5	3	4	10	1	5	4	4	2
		6	1	6	6	8	3	4	5	3
		7	8	3	5	2	1	9	2	5

		Least favourite 8	12	1	4	1	5	8	3	1
Kenya	<i>Female</i>	Most favourite 1	4	29	3	4	11	4	3	4
		2	9	13	4	7	5	3	11	7
		3	15	5	1	7	5	8	7	16
		4	9	6	8	8	8	5	7	10
		5	3	3	9	6	7	13	11	9
		6	6	2	9	16	7	7	12	1
		7	8	2	11	10	11	9	2	6
		Least favourite 8	7	1	16	3	7	12	8	8
South	<i>Male</i>	Most favourite 1	7	10	6	3	2	2	5	7
Africa		2	11	6	5	7	4	5	3	2
		3	7	12	4	4	2	7	1	5
		4	7	5	8	6	6	3	2	4
		5	2	1	6	9	6	5	8	5

		6	2	4	1	5	9	6	10	5
		7	4	3	7	4	4	9	5	6
		Least favourite 8	2	1	5	4	9	5	8	8
South	<i>Female</i>	Most favourite 1	5	30	4	3	3	2	4	3
Africa		2	12	7	7	3	6	1	8	9
		3	9	6	9	10	6	3	8	4
		4	10	2	6	11	7	7	9	4
		5	6	3	6	8	11	9	8	6
		6	6	3	8	10	7	7	5	9
		7	5	2	7	5	11	13	7	6
		Least favourite 8	2	2	8	5	4	13	6	14

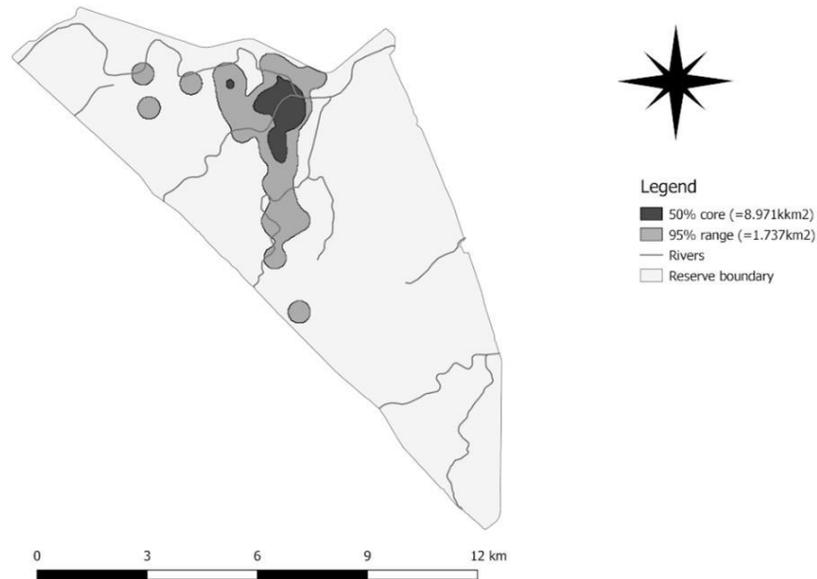


**Figure A.2.2.** Total participant response of what elephants eat: G = Grasses; O.S.A = Other small animals; T = Trees, and T + G = Trees and Grasses.

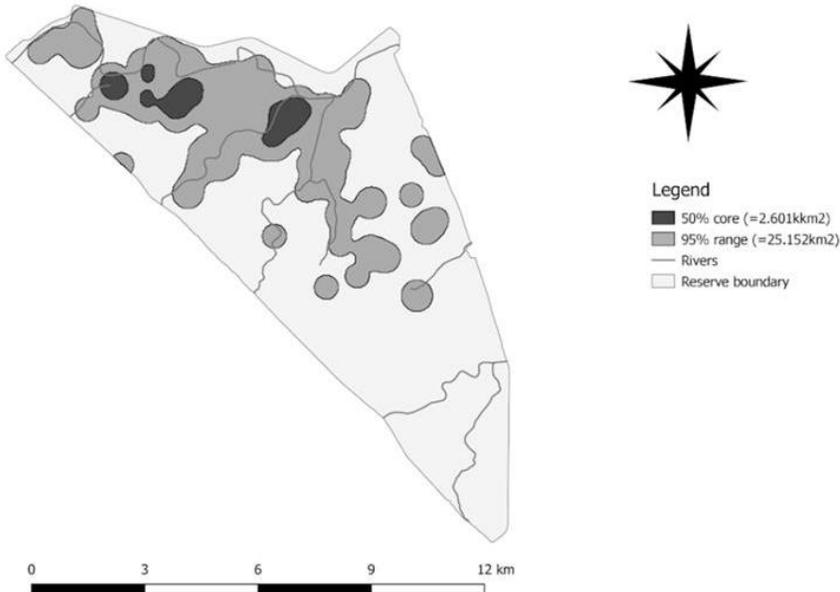
### Appendix 3.

**Figure A.3:** Elephant home- (95%) and core-range (50%) analysis from introduction (1999) until 2018 for breeding herd and bulls. Data were compiled for each month, and maps were generated from combined yearly data.

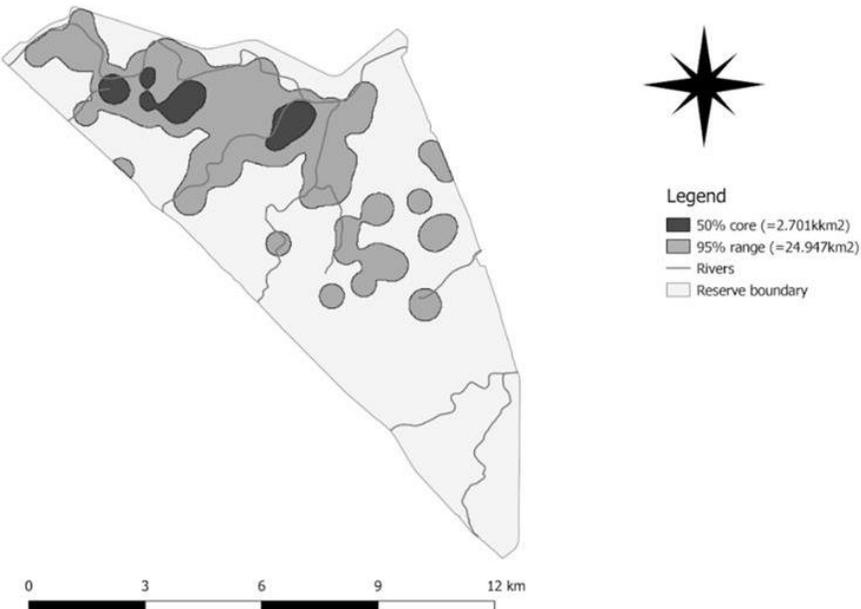
All elephants 1999



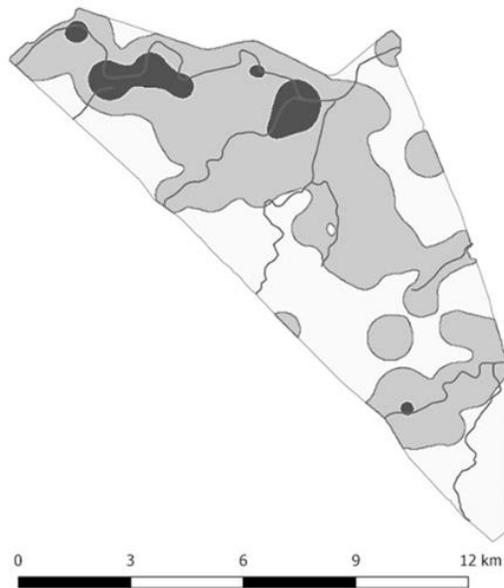
Breeding Herd 2000



Bull 2000



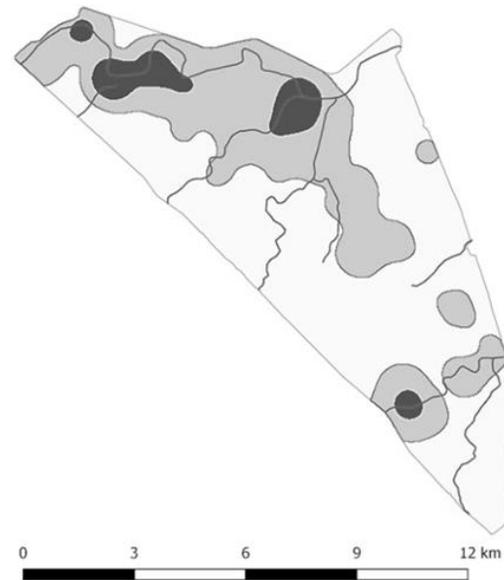
Breeding Herd 2001



Legend

- 50% core (=4.656kr)
- 95% range (=47.22t)
- Rivers
- Reserve boundary

Bull 2001



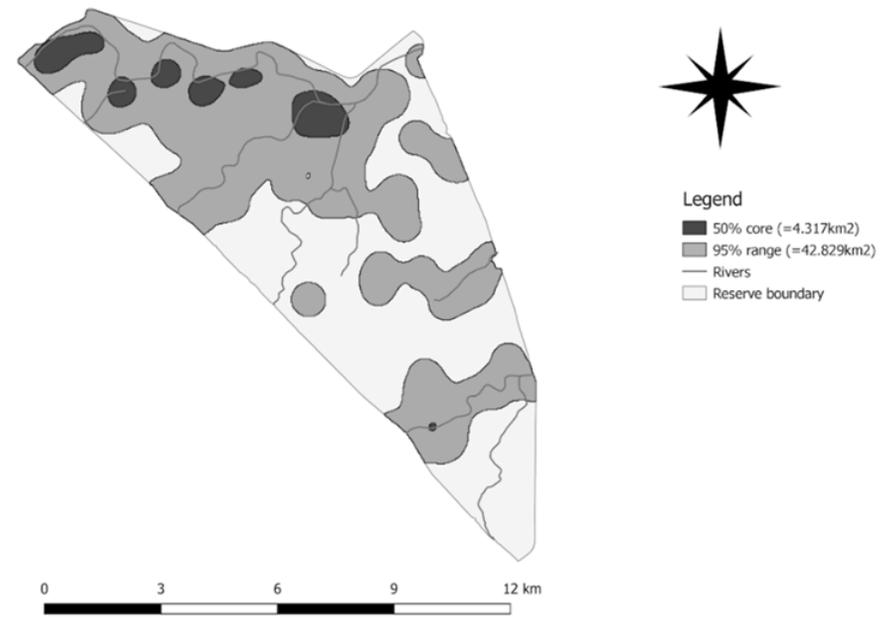
Legend

- 50% core (=4.973km<sup>2</sup>)
- 95% range (=32.246km<sup>2</sup>)
- Rivers
- Reserve boundary

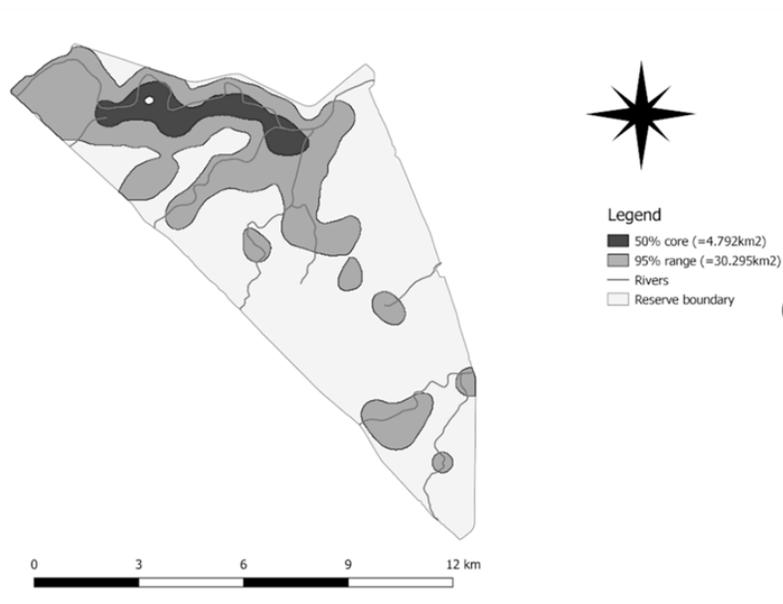
### Breeding Herd 2002



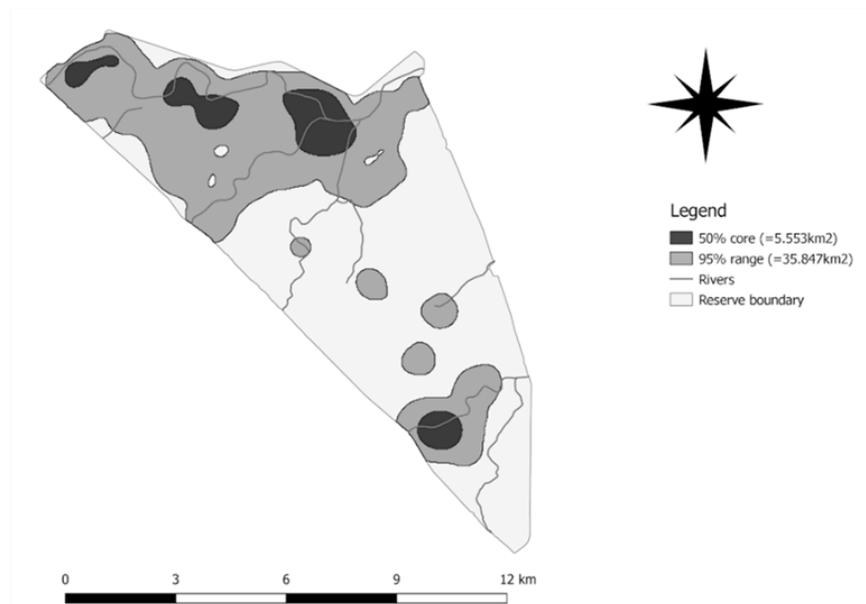
### Bulls 2002



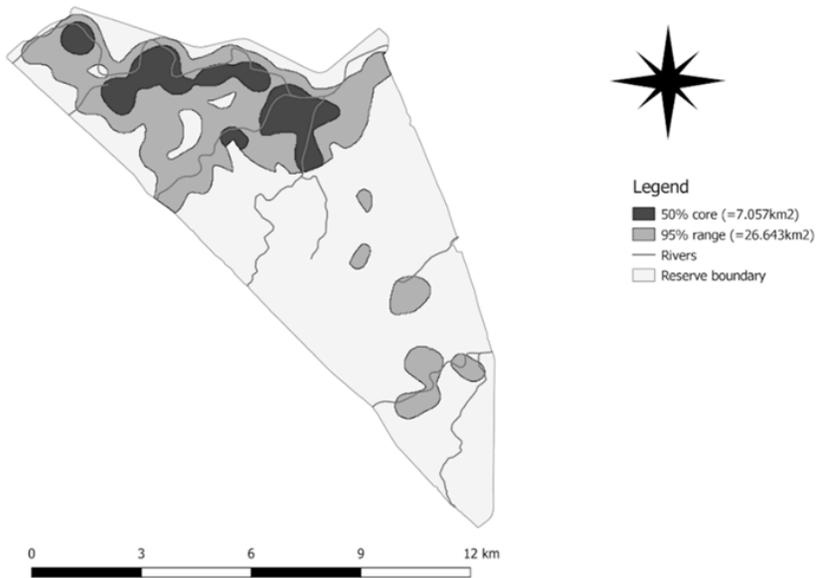
### Breeding Herd 2003



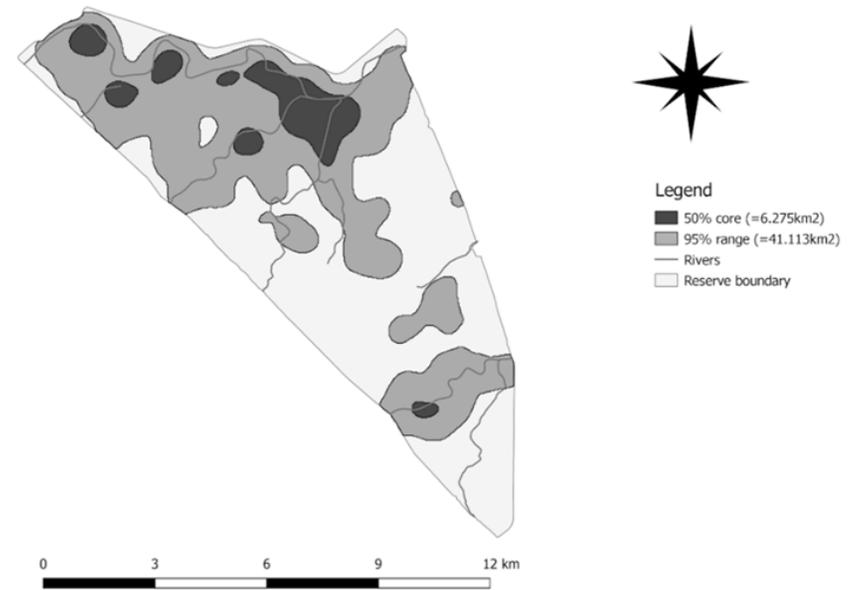
### Bulls 2003



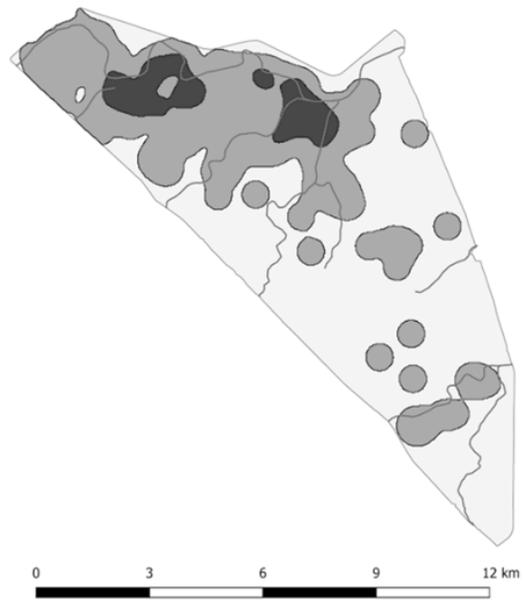
Breeding Herd 2004



Bulls 2004

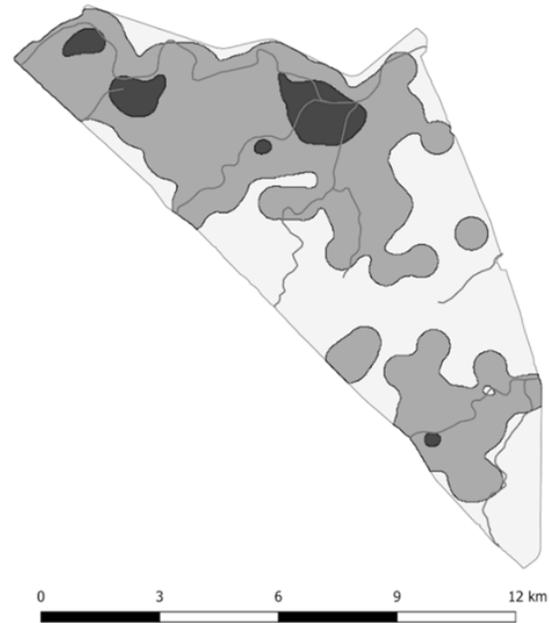


Breeding Herd 2005



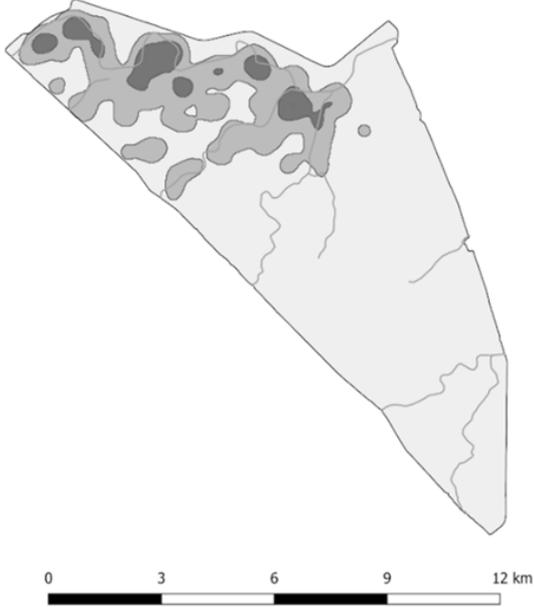
Legend  
■ 50% core (=5.044km)  
■ 95% range (=34.676l)  
— Rivers  
□ Reserve boundary

Bulls 2005

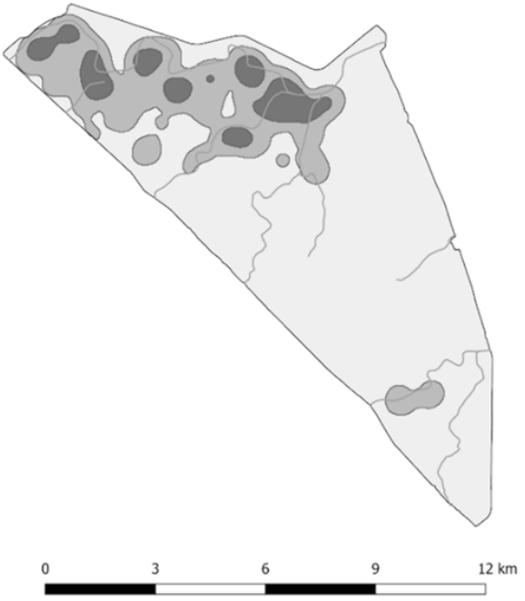


Legend  
■ 50% core (=4.495km<sup>2</sup>)  
■ 95% range (=45.869km<sup>2</sup>)  
— Rivers  
□ Reserve boundary

Breeding Herd 2006

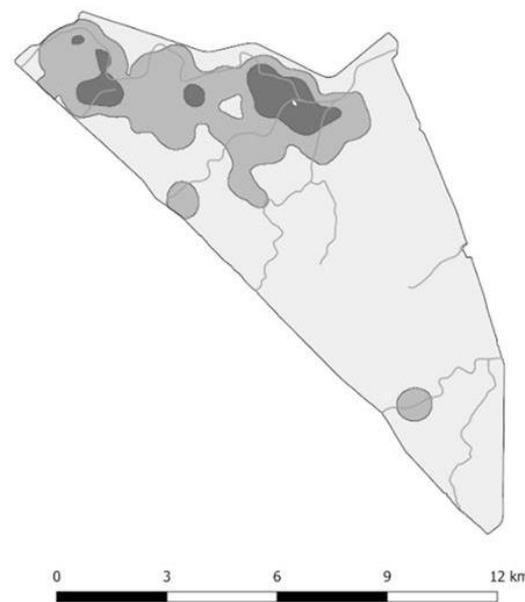
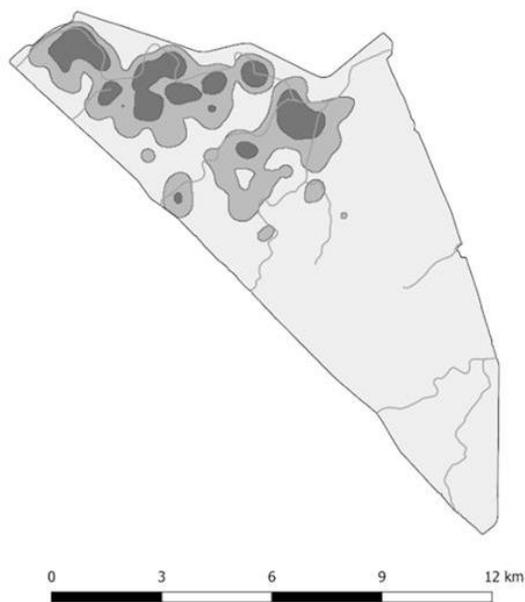


Bulls 2006

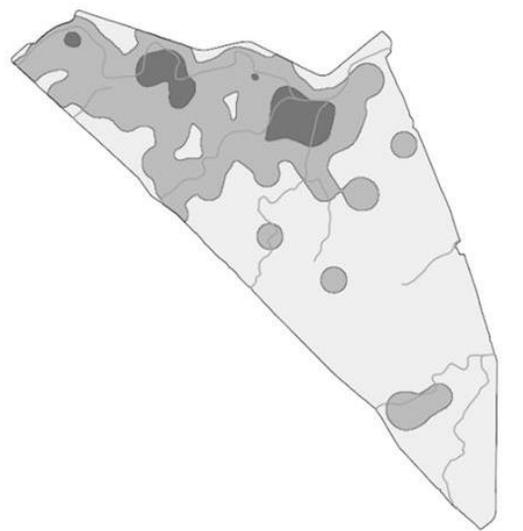


### Breeding Herd 2007

### Bulls 2007



## Breeding Herd 2008

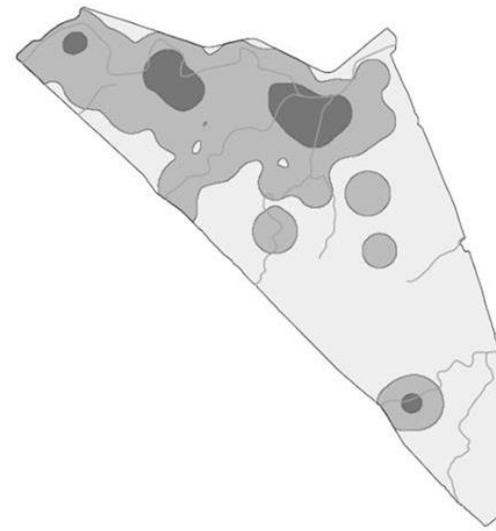


### Legend

- 50% core (=3.902km<sup>2</sup>)
- 95% range (=29.649km<sup>2</sup>)
- Rivers
- Reserve boundary



## Bulls 2008

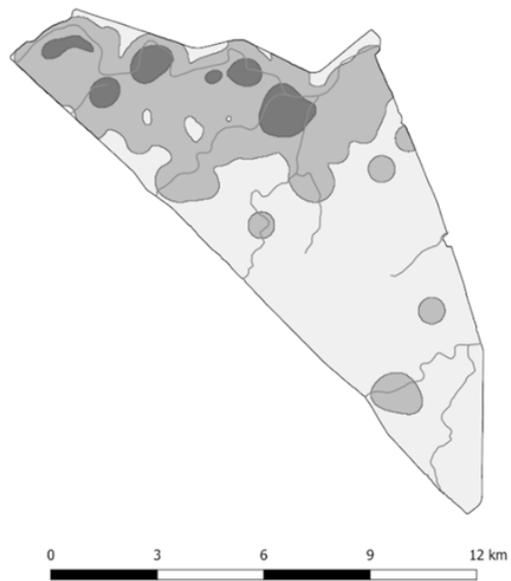


### Legend

- 50% core (=5.168km<sup>2</sup>)
- 95% range (=34.905km<sup>2</sup>)
- Rivers
- Reserve boundary



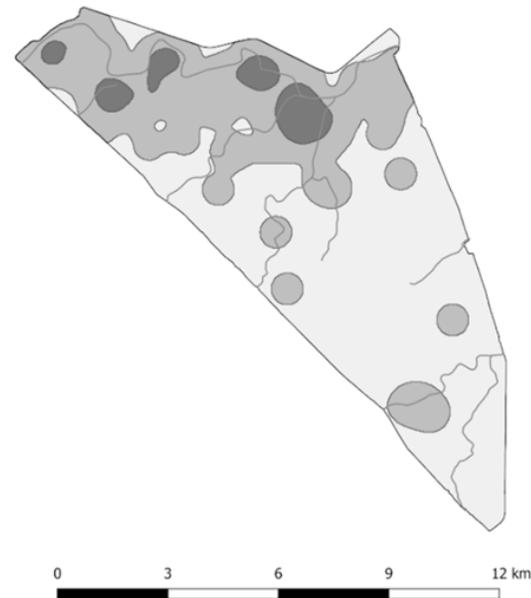
## Breeding Herd 2009



### Legend

- 50% core (=4.439km<sup>2</sup>)
- 95% range (=33.333km<sup>2</sup>)
- Rivers
- Reserve boundary

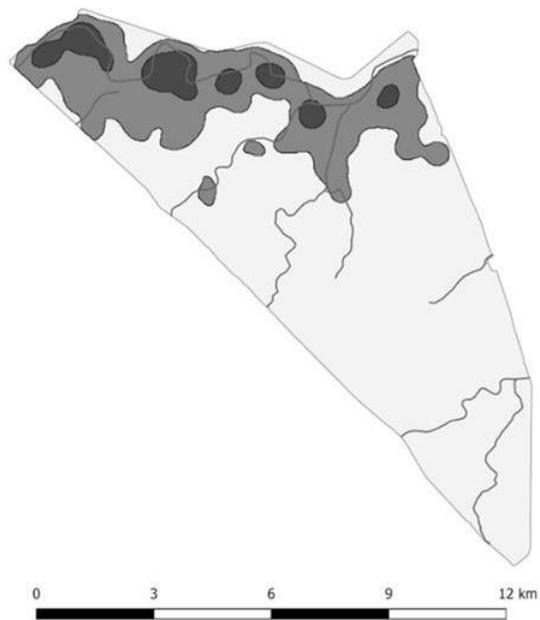
## Bulls 2009



### Legend

- 50% core (=4.352km<sup>2</sup>)
- 95% range (=33.307km<sup>2</sup>)
- Rivers
- Reserve boundary

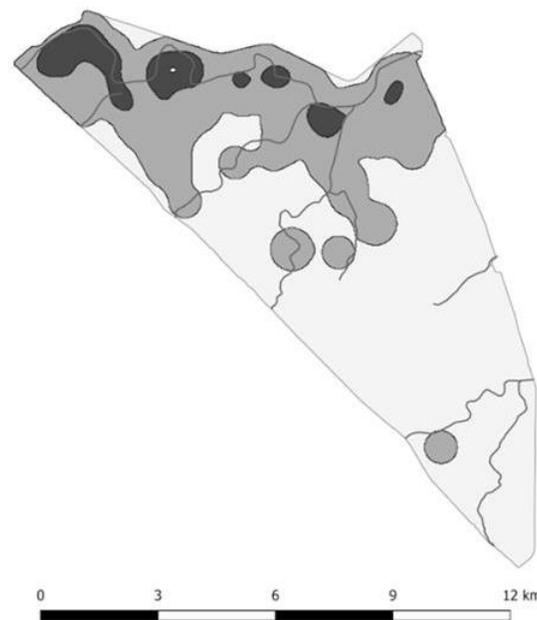
Breeding Herd 2010



Legend

- 50% core (=3.944km<sup>2</sup>)
- 95% range (=20.819km<sup>2</sup>)
- Rivers
- Reserve boundary

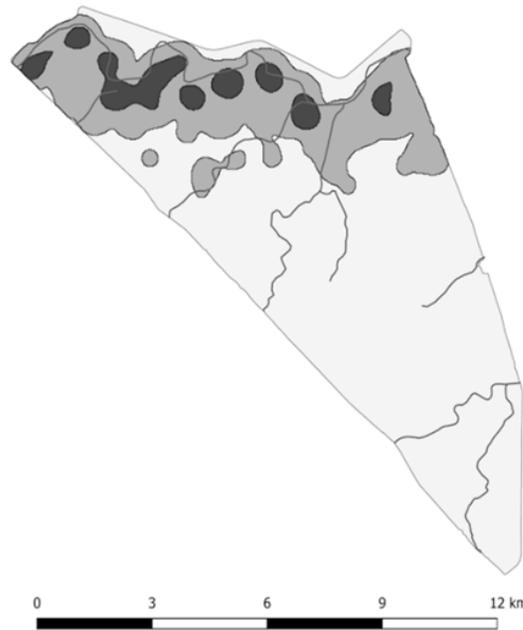
Bulls 2010



Legend

- 50% core (=4.851km<sup>2</sup>)
- 95% range (=31.555km<sup>2</sup>)
- Rivers
- Reserve boundary

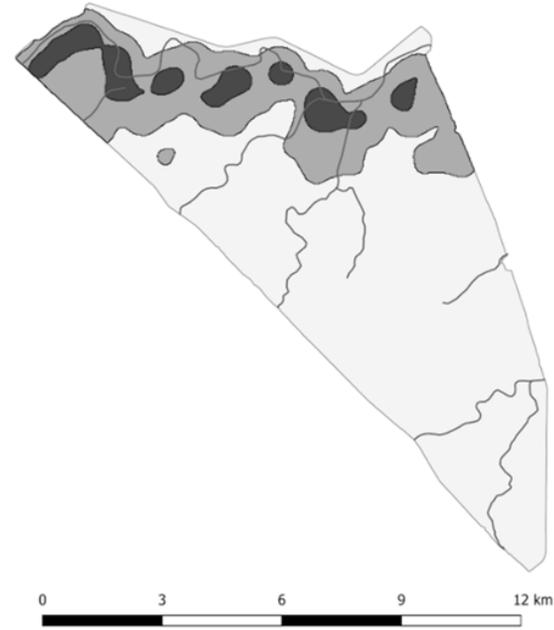
Breeding Herd 2011



Legend

- 50% core (=4.369km<sup>2</sup>)
- 95% range (=23.537km<sup>2</sup>)
- Rivers
- Reserve boundary

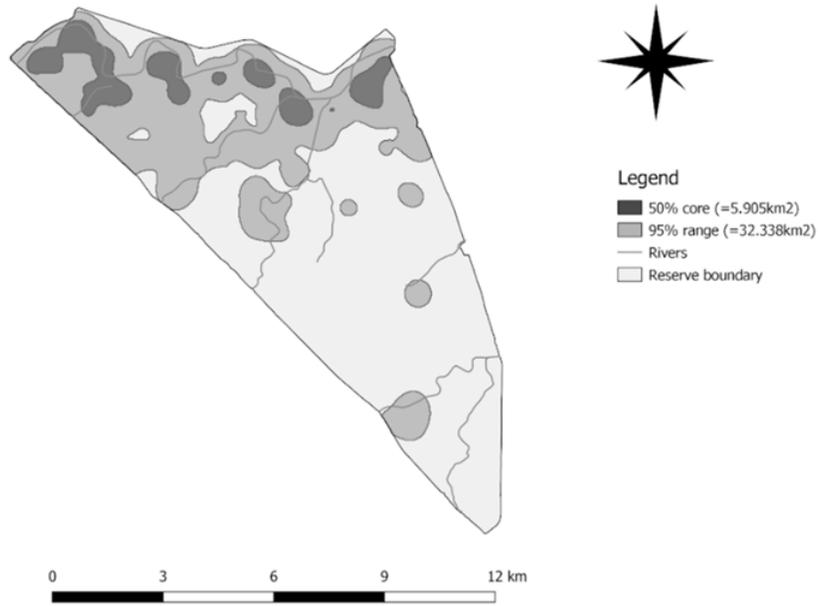
Bulls 2011



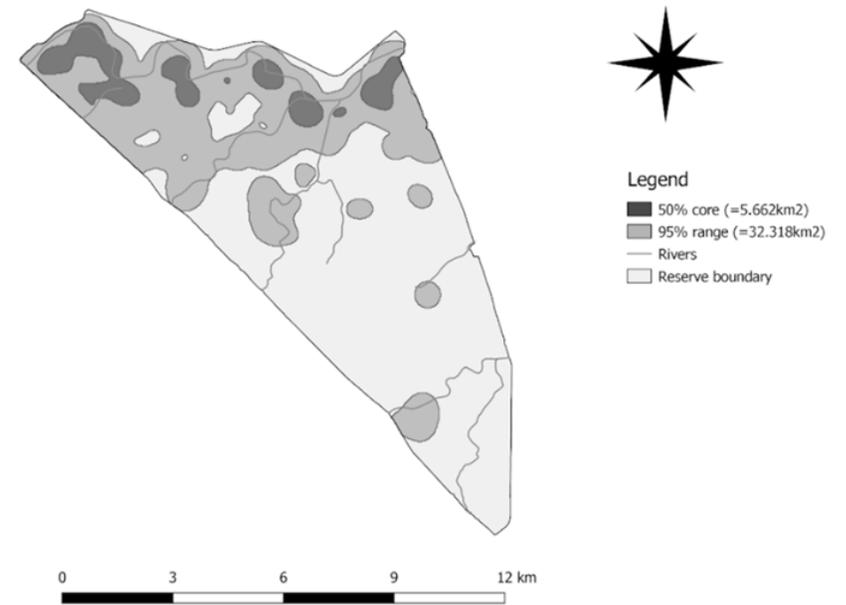
Legend

- 50% core (=4.924km<sup>2</sup>)
- 95% range (=22.603km<sup>2</sup>)
- Rivers
- Reserve boundary

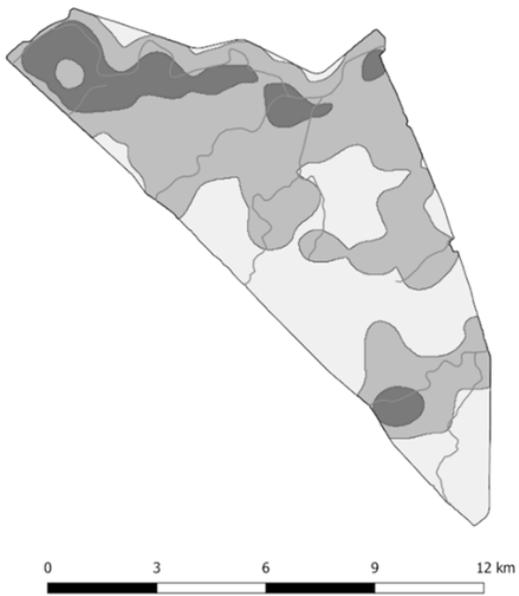
## Breeding Herd 2012



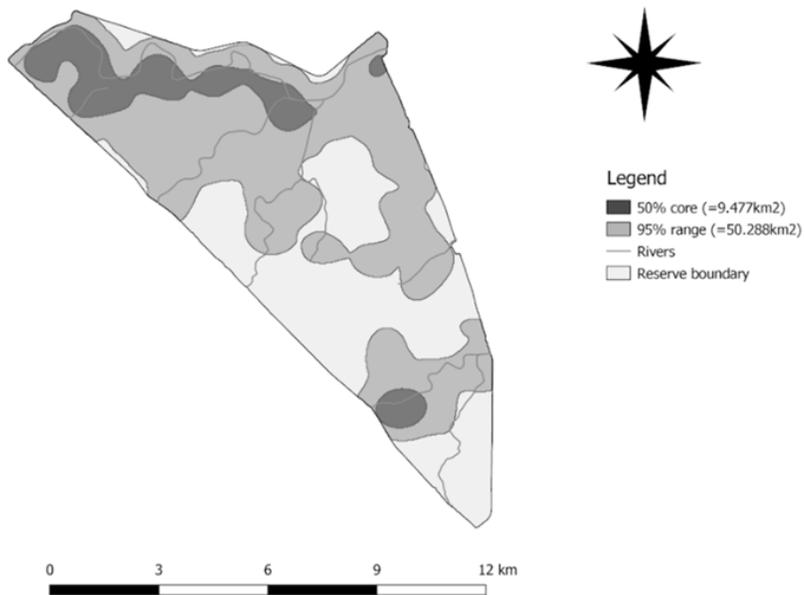
## Bulls 2012



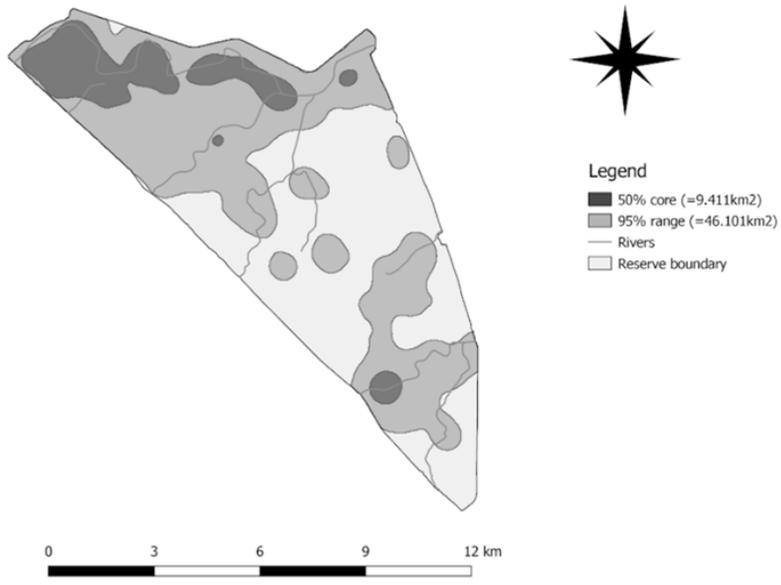
### Breeding Herd 2013



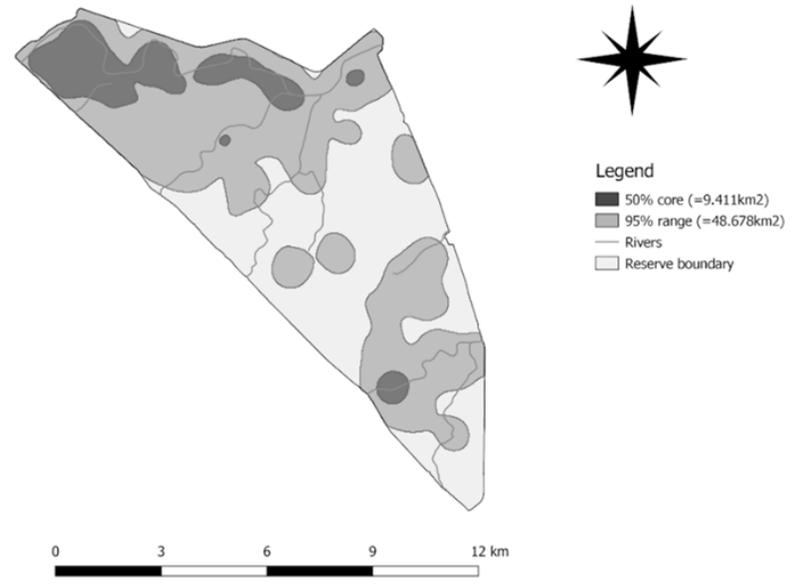
### Bulls 2013



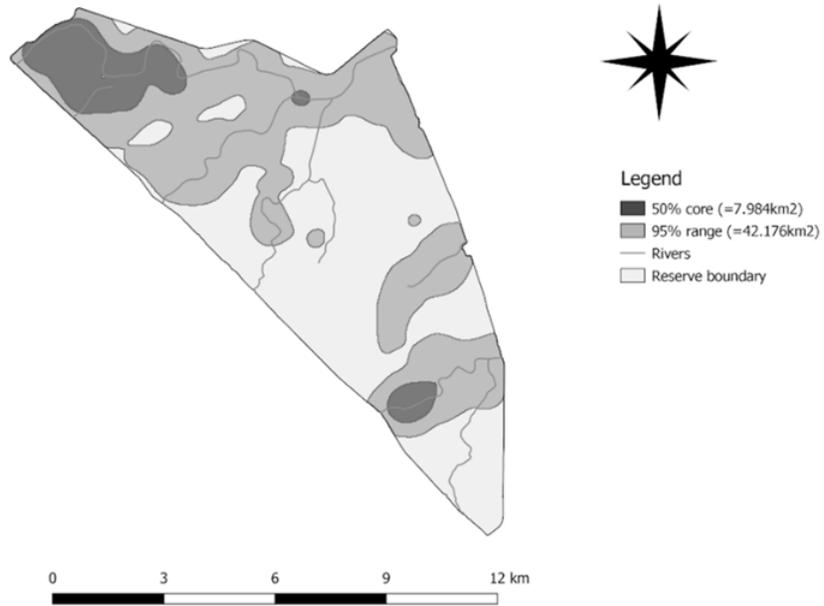
### Breeding Herd 2014



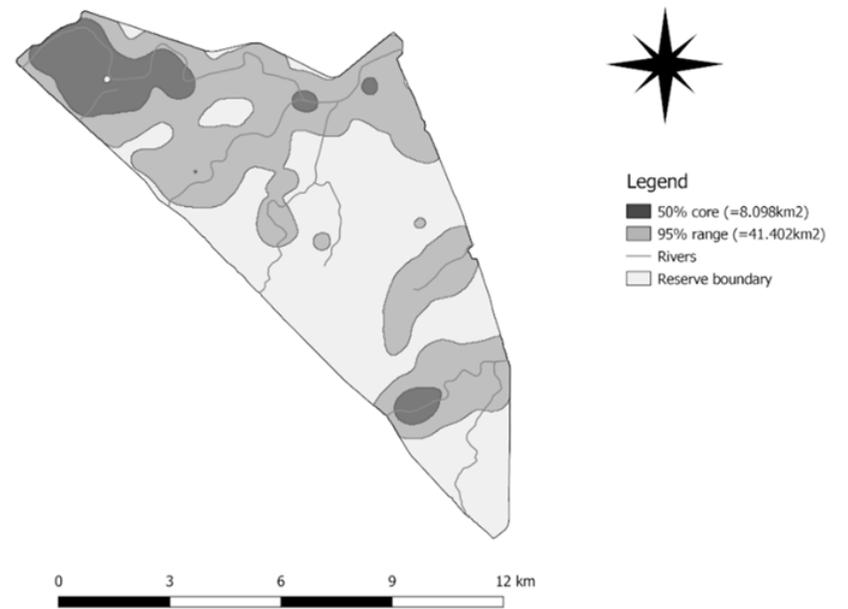
### Bulls 2014



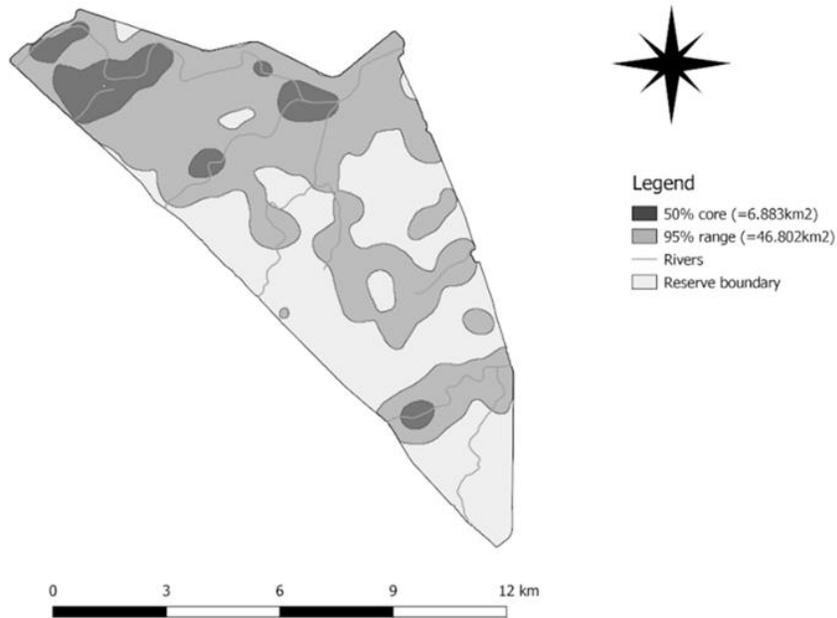
### Breeding Herd 2015



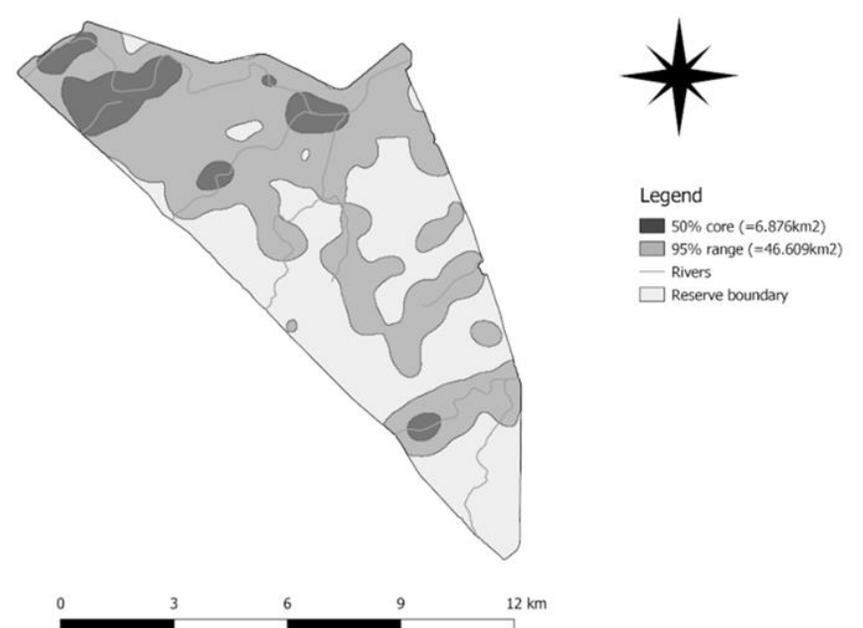
### Bulls 2015



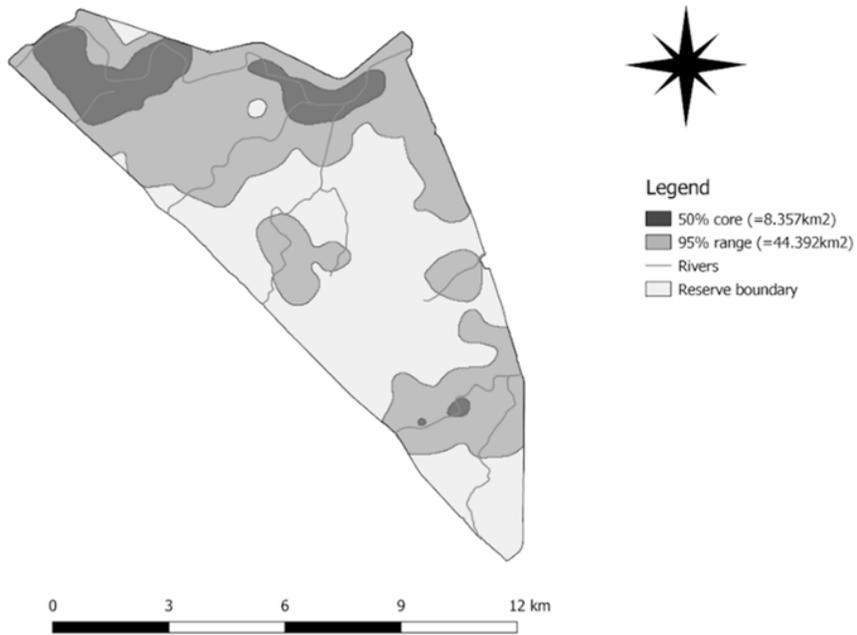
## Breeding Herd 2016



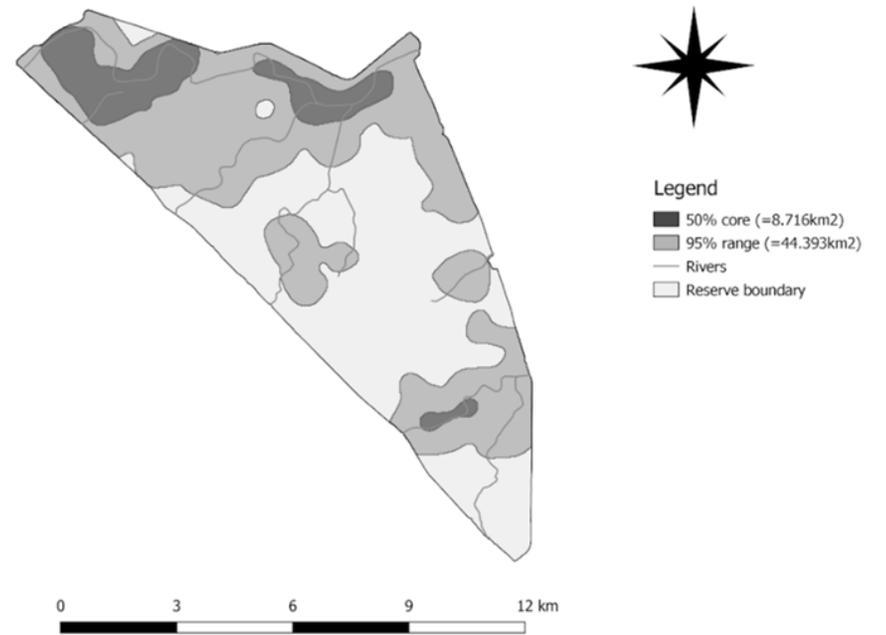
## Bulls 2016



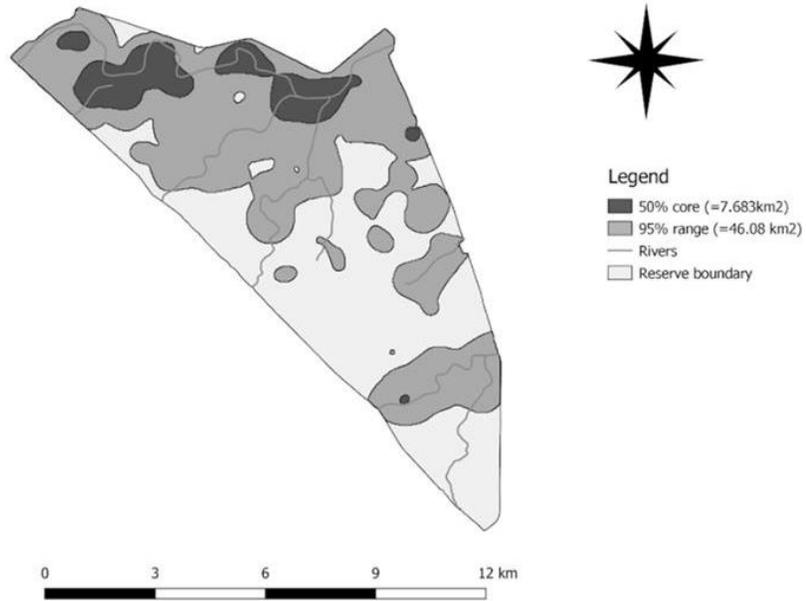
### Breeding Herd 2017



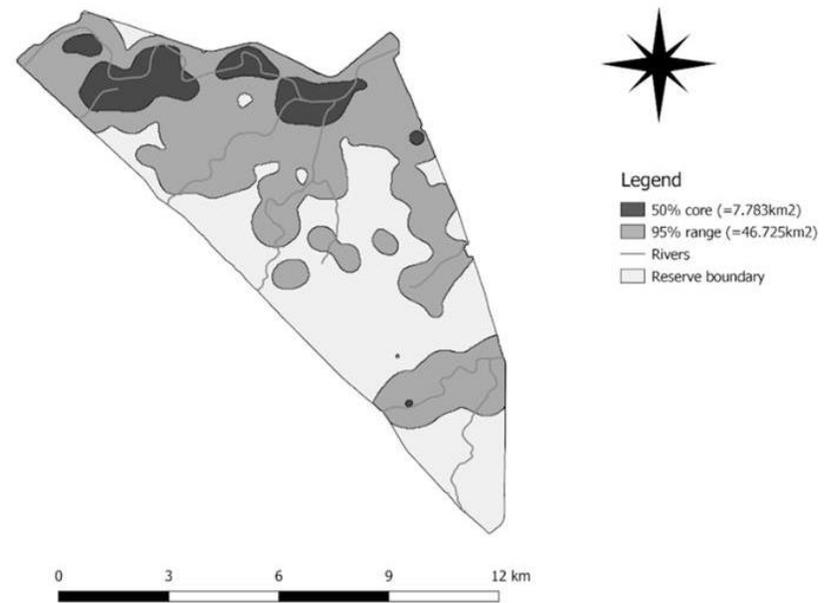
### Bulls 2017



## Breeding Herd 2018



## Bulls 2018



#### Appendix 4.

**Table A.4.1:** Raw data collected in 2018 for Chapter 2. PO = Pushed Over; BB = Branches Broken; MSB = Main Stem Broken; D =Debarking; C = Coppicing; S = Sprouting; R = Regrowth; FL = Fence Line

Species	Height (m)	Alive	PO	BB	MSB	D	C	S	R	FL
<i>Combretumapiculatum</i>	5	1	1	1	0	0	1	1	0	1
<i>Combretumapiculatum</i>	6.5	1	0	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6.5	1	0	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	1	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	1	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5.5	1	1	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	1
<i>Commiphoramollis</i>	7	1	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6.3	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	1	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Commiphoramollis</i>	8	1	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.5	1	0	1	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.7	1	0	0	0	1	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.8	1	0	1	0	0	0	0	0	0

<i>Commiphoramollis</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	0	0
<i>Sclerocaryabirrea</i>	12	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	8.5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	13	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	6	1	1	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	0

<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	11	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	0	1	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	0	1	1	0	0	1	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	0	1	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	6	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	0

<i>Combretumapiculatum</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	0	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7.5	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	1	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	0	0	0	0	0	0	0	0

<i>Combretumapiculatum</i>	7	0	1	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6.5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	1	0	0	0	1
<i>Sengalianigrescens</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	1	0	1	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	0	1	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	0

<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.5	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	5	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	6	1	1	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7.5	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	6	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Combretumimberbe</i>	7	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	0	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0

<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0
<i>Combretumimberbe</i>	5	1	0	1	0	0	1	0	0	0
<i>Sengalianigrescens</i>	8	0	0	1	0	0	0	0	0	0
<i>Combretumimberbe</i>	12	1	0	1	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5	1	1	1	0	0	1	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	1	1	0	0	1	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	0	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	0	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	1	0	0	0	1
<i>Combretumapiculatum</i>	6	0	1	0	0	0	0	0	0	1

<i>Combretumimberbe</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	5	0	1	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	0	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	10	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	0	1	0	0	0	0	1	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6.5	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	8	0	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	0	1	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	14	1	0	1	0	1	0	0	0	0	1

<i>Sengalianigrescens</i>	16	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	15	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	11	0	1	0	0	1	1	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	14	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	16	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	15	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	11	0	1	0	0	1	1	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	1

<i>Sclerocaryabirrea</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	1	1	0	0	0	1	0	1	
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5	0	1	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5.5	1	1	1	0	0	1	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	0	1	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	1	1	0	0	1	0	0	0	0

<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumimberbe</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Combretumimberbe</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	12	1	0	1	0	1	1	0	1	1	1
<i>Sclerocaryabirrea</i>	12	1	0	1	0	1	1	0	1	1	1
<i>Combretumapiculatum</i>	6.5	1	1	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.5	1	0	0	1	0	1	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	0	1	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	7	1	1	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5	1	0	0	1	1	1	0	0	0	0
<i>Sclerocaryabirrea</i>	6	0	1	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	5.5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	0	1	1	0	0	0	0	0	0	0

<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	18	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	0	1	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	0	0	1	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	12	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	0	1	1	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	1	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	1	1	0	0	0	1
<i>Combretumapiculatum</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	1	0	0	0	0	1

<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	1	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	1	1	0	0	1
<i>Combretumapiculatum</i>	9	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	1	0	0	1
<i>Sclerocaryabirrea</i>	5	1	0	1	1	0	1	0	0	1
<i>Sclerocaryabirrea</i>	11	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	0	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	1	1	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	1	1	1	0	0	0	0	1
<i>Sclerocaryabirrea</i>	10	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	11	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9.5	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8.5	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	10	1	0	1	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	10	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8.5	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	10	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8.5	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	7	0	1	0	0	0	0	0	0	1



<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Combretumimberbe</i>	9	1	1	1	0	1	0	0	0	0	0
<i>Commiphoramollis</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	1	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	0	1	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	11	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	1	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	7	1	0	1	1	0	0	1	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	1	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	1	0	0	0	0	0	0	1

<i>Sengalianigrescens</i>	6	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	1	0	0	0	1
<i>Combretumimberbe</i>	8	0	1	0	0	0	0	1	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	6	0	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	1	0	0	1
<i>Sengalianigrescens</i>	8	0	0	1	0	1	0	0	0	1
<i>Combretumapiculatum</i>	7	1	1	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	0	1	1	0	1	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5	1	1	0	0	0	0	1	0	1
<i>Sengalianigrescens</i>	6	1	0	1	0	0	1	0	0	1
<i>Sengalianigrescens</i>	8	0	0	1	0	1	0	0	0	1
<i>Combretumapiculatum</i>	7	1	1	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	0	1
<i>Combretumimberbe</i>	6	1	0	1	0	0	1	0	0	1
<i>Sengalianigrescens</i>	6	1	0	0	0	1	0	0	0	1
<i>Sengalianigrescens</i>	8	1	1	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	1	0	1	0	0	0	1
<i>Combretumimberbe</i>	14	1	0	1	0	0	0	0	0	1
<i>Sengalianigrescens</i>	13	1	0	1	0	1	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	1	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0

<i>Sengalianigrescens</i>	10	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	0	0	0
<i>Combretumimberbe</i>	7	0	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Combretumimberbe</i>	7	0	1	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	10	0	0	1	0	1	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	1	0	0	0	0	0
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Combretumapiculatum</i>	5	0	1	0	0	0	0	0	0	0	0
<i>Combretumimberbe</i>	6	0	0	1	1	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	7	1	0	1	0	0	1	0	0	0	1
<i>Combretumimberbe</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	8.5	1	0	1	0	0	1	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	0	1	0	0	0	1
<i>Sengalianigrescens</i>	9	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	1	0	0	0	1	1	0	0	1
<i>Combretumapiculatum</i>	7	1	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	0	0	1	0	0	1	1	1
<i>Sengalianigrescens</i>	9	1	0	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	1	0	0	1	1	1
<i>Sengalianigrescens</i>	8	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	9	1	0	0	0	0	1	0	0	0	1

<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	0	0	0	0	1	0	0	1
<i>Combretumapiculatum</i>	7	1	1	0	0	0	1	1	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	8	0	1	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	13	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	11	1	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	16	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	9	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	8	0	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	9	1	0	0	0	0	1	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	8	1	0	0	0	0	0	1	0	0	1
<i>Combretumapiculatum</i>	7	1	1	0	0	0	1	1	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	8	0	1	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	1	0	0	1	0	1
<i>Sengalianigrescens</i>	13	1	0	0	0	0	0	0	0	0	1

<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	11	1	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	16	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	10	1	1	0	0	0	1	0	1	1	1
<i>Combretumimberbe</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	12	1	1	0	0	0	1	0	0	0	1
<i>Combretumimberbe</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	11	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	9	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6	0	1	0	0	0	1	0	0	0	1
<i>Combretumapiculatum</i>	6	1	1	1	0	0	1	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9.5	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1

<i>Sclerocaryabirrea</i>	9	1	0	0	0	1	0	0	1	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	10	1	1	0	0	0	1	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9.5	1	0	0	0	1	0	0	1	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	1	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9.5	1	0	0	0	1	0	0	1	1
<i>Sengalianigrescens</i>	6	1	1	0	0	0	1	1	0	0
<i>Combretumapiculatum</i>	8	0	1	1	0	0	1	0	0	0
<i>Sengalianigrescens</i>	12	1	0	1	0	0	0	0	0	0
<i>Combretumimberbe</i>	11	1	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.5	1	0	0	0	0	0	0	0	0

<i>Sengalianigrescens</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Sengalianigrescens</i>	8	0	1	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	0	1	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	1	0	0	0	0	1
<i>Combretumapiculatum</i>	5.5	1	1	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	1	1	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8.5	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Commiphoramollis</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	9	1	0	0	0	0	0	0	0	0	1

<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	0	1
<i>Combretumimberbe</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	1	0	1	0	0	0	1	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	10	1	0	1	0	1	0	0	0	1	1
<i>Sclerocaryabirrea</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	11	0	1	1	0	1	0	0	0	1	1
<i>Sclerocaryabirrea</i>	13	1	0	1	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Combretumapiculatum</i>	9	1	1	0	0	0	0	1	0	0	1
<i>Sclerocaryabirrea</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	13	0	1	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sclerocaryabirrea</i>	14	1	0	0	0	1	0	0	0	1	1
<i>Sengalianigrescens</i>	11	1	0	0	0	1	0	0	0	1	1
<i>Sengalianigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Sengalianigrescens</i>	8	1	0	0	0	0	0	0	0	0	1

<i>Sengalia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
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**Table A.4.2:** Raw data collected in 2019 for Chapter 3. PO = Pushed Over; BB = Branches Broken; MSB = Main Stem Broken; D =Debarking; C = Coppicing; S = Sprouting; R = Regrowth; IP = Insect Presence

Species	Height (m)	A	PO	BB	MSB	D	C	S	R	IP
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.6	1	0	1	0	0	0	0	0	0
<i>Combretum imberbe</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	12	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	12	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.2	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8.8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.3	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.1	1	0	0	0	0	0	0	0	0



<i>Combretum apiculatum</i>	6.8	1	0	2	0	5	1	0	1	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	10	1	0	1	0	0	1	0	0	0
<i>Combretum apiculatum</i>	9.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	3	0	0	1	0	0	0
<i>Commiphora mollis</i>	7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10	1	0	3	0	5	1	0	0	0
<i>Commiphora mollis</i>	7.8	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	10	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	12	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	11.3	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.8	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.4	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.9	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.2	1	0	0	0	25	0	0	1	0
<i>Combretum apiculatum</i>	5.4	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.1	1	0	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	10.9	1	0	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	9.5	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	1

<i>Sclerocarya birrea</i>	4.5	1	0	4	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	1	0	1	0	1	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.1	1	0	1	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	7.7	1	0	4	1	0	1	0	0	1
<i>Combretum apiculatum</i>	4	1	0	2	1	0	1	0	0	1
<i>Sclerocarya birrea</i>	11.2	1	0	1	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	6	0	0	1	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	1	2	0	0	1	1	0	1
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.72	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.72	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.16	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	7.04	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.72	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.36	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	7.92	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	1	0	0	0	0	0	0
<i>Commiphora mollis</i>	7.92	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.92	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	3.96	1	0	6	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	1

<i>Commiphora mollis</i>	6.16	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6.16	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	7.04	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.28	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	2	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.4	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.2	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6.3	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.8	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.1	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.2	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.8	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	4	1	0	3	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	0

<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.3	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.1	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.2	1	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	1

<i>Combretum apiculatum</i>	6	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.2	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	1	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.8	1	0	2	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.5	1	0	0	0	0	0	0	1	0
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4.2	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	4.9	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7.9	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.2	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9.1	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.4	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.6	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.9	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0

<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.1	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	1	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.1	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.2	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.5	1	0	2	0	5	0	0	0	0	1
<i>Combretum apiculatum</i>	2.9	0	0	1	0	60	0	0	0	0	1
<i>Combretum apiculatum</i>	5.4	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	1	0	0	30	0	1	0	0	1

<i>Combretum apiculatum</i>	4.9	1	0	0	0	5	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.8	1	0	1	0	5	0	0	0	1
<i>Combretum apiculatum</i>	5	1	1	0	0	20	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	1	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	1	0	1
<i>Commiphora mollis</i>	4.7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.4	0	1	0	0	0	0	1	0	0
<i>Sclerocarya birrea</i>	5.4	1	0	3	0	0	0	0	0	0
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.1	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	1	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.7	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	2	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.7	1	0	3	0	0	0	0	0	1

<i>Combretum apiculatum</i>	4.5	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.6	1	1	2	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	1	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6.3	1	0	4	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8.3	1	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	3.9	1	0	1	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	9.6	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7.2	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.8	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9.2	1	0	1	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	7.2	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	11.6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	3	0	0	1	0	0	0	0

<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.1	1	0	2	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.8	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	4	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.3	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	3	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	0	0
<i>commiphora mollis</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	3	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	1	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	10.6	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	0	2	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.3	1	0	1	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	2	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	2	0	0	0	0	0	0	1

<i>Combretum apiculatum</i>	6.1	1	0	1	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.1	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.9	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	2	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.8	1	1	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.1	1	1	2	0	0	1	1	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	11	1	0	2	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	1	0	0	0	0
<i>Combretum apiculatum</i>	7.5	1	1	3	0	0	1	1	0	0	1
<i>Combretum apiculatum</i>	4.5	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	0	1	0	0	1	0	0	0	0
<i>Sclerocarya birrea</i>	9.4	1	0	3	0	0	1	0	0	0	1
<i>Sclerocarya birrea</i>	5.4	1	0	1	0	0	1	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	1	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	2	0	0	1	0	0	0	0
<i>Combretum apiculatum</i>	5.9	1	0	1	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	8	1	0	0	0	0	0	0	0	0	0

<i>Combretum apiculatum</i>	6	1	1	2	0	0	1	1	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.5	1	1	2	0	0	1	1	0	1
<i>Combretum apiculatum</i>	6.2	1	0	2	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8	1	0	5	0	0	1	0	0	0
<i>Commiphora mollis</i>	4.7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.9	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.8	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.3	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	12	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.3	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.8	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5.4	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5.3	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.7	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5.4	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	0	0	5	0	0	0	1
<i>Sclerocarya birrea</i>	12	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.8	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	10	1	1	1	0	0	0	1	0	1

<i>Commiphora mollis</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8.5	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.6	1	0	0	0	0	0	0	1	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	1	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	0	0	0	20	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	13	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.5	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.9	1	0	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	9	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	1	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0

<i>Commiphora mollis</i>	5.5	1	0	3	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.8	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.2	1	0	5	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	5	1	1	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	2	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10	0	0	4	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	1	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.1	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.9	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	1	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	6	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7	1	0	2	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7.6	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	3	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	6.5	1	0	2	0	0	0	0	0	0
<i>Combretum imberbe</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	8.1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	1	0	0	0	0	0	0
<i>Combretum imberbe</i>	6.5	1	0	1	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	1	1	0	0	0	1	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	6.7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	2	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.1	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	4.5	1	0	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6.2	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5	1	0	1	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5.1	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	4.1	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	4.6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.3	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.9	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.8	0	1	1	0	0	0	0	1	0	1
<i>Sclerocarya birrea</i>	7.5	1	0	2	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	7	1	0	0	0	0.2	0	0	0	1	1
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.6	1	0	0	0	0	0	0	0	0	0

<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10.6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.5	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8	1	0	1	0	0	1	0	0	0
<i>Combretum apiculatum</i>	7.2	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.9	1	0	2	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	3	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	4	1	0	5	0	0	1	0	0	0
<i>Combretum apiculatum</i>	5.2	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	7.5	1	0	2	0	0	1	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	4	0	0	1	0	0	0
<i>Commiphora mollis</i>	4	1	1	1	0	0	1	1	0	1
<i>Commiphora mollis</i>	5.1	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.1	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.9	1	0	0	0	10	0	0	0	1
<i>Commiphora mollis</i>	4.8	1	1	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	8	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	15	0	0	1	0
<i>Commiphora mollis</i>	6.2	1	0	0	0	30	0	0	1	0
<i>Commiphora mollis</i>	5.8	1	0	1	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	1

<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	8.8	1	0	4	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	8.6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	4	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	1	0	0	1	0	0	0	0
<i>Acacia nigrescens</i>	4.6	1	0	4	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.8	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	7.3	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	8.4	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4	1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	6	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4	1	1	2	0	0	1	1	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.8	1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	1	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.3	1	1	0	0	0	0	0	0	0	0

<i>Acacia nigrescens</i>	8	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5	1	0	2	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.4	1	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7	1	0	3	0	0	0	0	0	0
<i>Commiphora mollis</i>	5.7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.5	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4.6	0	1	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	8	1	0	1	0	0	1	0	0	0
<i>Acacia nigrescens</i>	5.4	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.4	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.16	1	0	2	0	0	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.5	1	1	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.5	1	1	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	5	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.28	1	1	2	0	0	0	0	0	0
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.28	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5	1	1	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	5	1	0	4	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	4.84	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	3.96	1	0	3	0	0	1	0	0	0

<i>Combretum apiculatum</i>	6.16	1	0	2	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.28	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.84	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	4.84	1	0	6	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.04	1	0	3	0	0.05	0	0	1	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	2	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7.48	1	0	1	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.28	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.84	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	4	0	0	1	0	0	0
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6.16	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.28	1	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.28	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	4.3	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.9	1	0	2	0	0	0	0	0	1
<i>Commiphora mollis</i>	5.1	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	5	0	0	1	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6	1	0	0	0	0	0	0	0	0

<i>Sclerocarya birrea</i>	<b>9</b>	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>5.5</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.4	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>7.5</b>	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>9</b>	1	0	1	0	0	0	0	0	0
<i>Combretum imberbe</i>	6.8	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	4.8	1	1	1	0	0	1	1	0	1
<i>Combretum apiculatum</i>	6.5	1	1	3	0	0	1	0	0	1
<i>Commiphora mollis</i>	<b>5</b>	1	1	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>5</b>	1	1	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.3	1	1	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.3	1	0	2	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.3	1	0	2	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>6</b>	1	0	3	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	8.1	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	4	0	0	0	0	0	1

<i>Combretum apiculatum</i>	6	1	0	1	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.4	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	1	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.7	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.1	1	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	2	0	0	1	0	0	1
<i>Commiphora mollis</i>	5	1	1	1	1	0	1	0	0	0
<i>Acacia nigrescens</i>	7	1	0	1	0	0	1	0	0	0
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	5.2	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4.2	1	0	0	0	0	0	0	0	0

<i>Commiphora mollis</i>	5	1	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.5	0	1	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9.5	1	0	4	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	6.8	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	1	2	0	0	1	1	0	1
<i>Combretum apiculatum</i>	6	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	2	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	4	1	0	3	0	0	1	0	0	0
<i>Combretum apiculatum</i>	8.3	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	4.5	1	0	7	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	1	1	0	0	1	1	0	0
<i>Sclerocarya birrea</i>	4.7	1	0	1	0	0	1	0	0	0
<i>Commiphora mollis</i>	5.1	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	1	3	0	0	1	1	0	1
<i>Combretum imberbe</i>	5.2	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	6.1	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1

<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.5	1	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	9.9	1	0	0	0	0.2	0	0	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	4	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10.1	1	0	2	0	0	1	0	0	0	0
<i>Sclerocarya birrea</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.5	1	0	2	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	8.8	1	0	3	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	1	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	7.2	1	0	2	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	<b>8</b>	1	0	3	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>8</b>	1	0	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.3	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.7	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>9</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.8	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.8	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.6	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>9</b>	1	0	4	0	0	1	0	0	1
<i>Combretum apiculatum</i>	4	1	1	0	0	0	1	1	0	0
<i>Combretum apiculatum</i>	5.2	1	0	1	0	0	1	0	0	0
<i>Acacia nigrescens</i>	<b>10</b>	1	0	2	0	0	1	0	0	0
<i>Acacia nigrescens</i>	8.8	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>6</b>	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>9</b>	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.1	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>6</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>11</b>	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	2	0	0	0	0	0	0

<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>10</b>	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.5	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	<b>5</b>	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.3	1	0	3	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>9</b>	1	0	3	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	3	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.1	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>8</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	1	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.3	1	0	2	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.1	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.8	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	8.4	1	0	2	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.4	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	1	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>7</b>	1	0	3	0	0	1	0	0	1
<i>Combretum apiculatum</i>	9.3	1	0	2	0	0	0	0	0	1

<i>Combretum apiculatum</i>	7.4	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	4.8	1	1	3	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	<b>9</b>	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	<b>4.5</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7.5</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.2	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.1	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.2	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>4.5</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.1	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.1	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.3	1	0	2	0	0	0	1	0	0	1

<i>Combretum imberbe</i>	7	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	11	1	0	0	0	0.7	0	0	1	1
<i>Acacia nigrescens</i>	7.9	1	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	2	0	0	1	0	0	0
<i>Acacia nigrescens</i>	10	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	6.8	1	0	2	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	10	1	0	3	0	0.2	1	0	1	1
<i>Combretum apiculatum</i>	7	1	0	2	1	0	1	0	0	1
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	12	1	0	1	0	0.5	1	0	1	1
<i>Acacia nigrescens</i>	11	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10.5	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	13	1	0	5	0	0.05	1	0	1	1
<i>Combretum apiculatum</i>	6.6	1	1	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	9	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	10	1	0	3	0	0	1	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	8	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.16	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	10	1	0	1	0	0	0	0	0	1
<i>Combretum imberbe</i>	10	1	0	1	0	0	0	0	0	0
<i>Combretum imberbe</i>	10.8	1	0	0	0	0	0	0	0	1

<i>Combretum imberbe</i>	<b>11</b>	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	<b>5</b>	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>4</b>	1	0	7	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	<b>5</b>	1	0	2	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	<b>6</b>	1	0	2	0	0	0	0	0	0
<i>Combretum apiculatum</i>	<b>6.5</b>	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.2	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	<b>8</b>	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>6</b>	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>7</b>	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	<b>4</b>	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	5	0	0.4	0	0	0	1
<i>Combretum apiculatum</i>	<b>5.5</b>	1	1	1	0	0	1	0	0	1
<i>Commiphora mollis</i>	<b>5</b>	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	4.4	1	0	3	0	0	1	0	0	1
<i>Commiphora mollis</i>	4.3	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.1	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>11</b>	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>6</b>	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	1	0	0	1	0	0	1
<i>Commiphora mollis</i>	<b>5</b>	1	1	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	<b>4</b>	1	1	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	<b>6</b>	1	0	6	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	5.8	1	0	4	0	0	1	0	0	1

<i>Sclerocarya birrea</i>	4.9	1	0	9	0	0	1	0	0	1
<i>Combretum apiculatum</i>	4.5	1	0	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.5	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6.5	1	0	5	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	7	1	0	3	0	0	1	0	0	1
<i>Commiphora mollis</i>	5.6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	1	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.3	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.2	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	6	1	0	7	0	0	1	0	0	1
<i>Commiphora mollis</i>	5.8	1	0	2	0	0	1	0	0	0
<i>Combretum apiculatum</i>	6.4	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6	1	0	2	0	0	1	0	0	1
<i>Commiphora mollis</i>	5.5	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.4	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.5	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.6	1	0	1	0	0	1	0	0	0
<i>Acacia nigrescens</i>	7.3	1	1	3	0	0	1	1	0	1
<i>Combretum apiculatum</i>	5	1	0	3	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.5	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.4	1	0	1	0	0	1	0	0	1
<i>Commiphora mollis</i>	5.72	1	0	0	0	0.25	0	0	0	1
<i>Sclerocarya birrea</i>	5.5	1	0	0	0	0	0	0	0	0

<i>Acacia nigrescens</i>	6.16	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>6.5</b>	1	0	1	0	0	1	0	0	0	0
<i>Acacia nigrescens</i>	7.04	1	0	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.16	1	0	0	0	0	1	0	0	0	0
<i>Acacia nigrescens</i>	7.04	1	0	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.92	1	1	5	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.04	1	0	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.04	1	0	0	0	0	0	0	0	0	0
<i>Combretum imberbe</i>	6.16	1	0	2	0	0	1	0	0	0	0
<i>Acacia nigrescens</i>	7.48	1	0	1	0	0	1	0	0	0	0
<i>Sclerocarya birrea</i>	7.92	1	0	2	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.8	1	1	3	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.8	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.04	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.04	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.92	0	1	5	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>5</b>	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	5	0	0	0	0	1
<i>Commiphora mollis</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	<b>4</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	7.4	1	0	1	0	0	1	0	0	0	0
<i>Sclerocarya birrea</i>	<b>7</b>	1	0	1	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	<b>5</b>	1	0	0	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	<b>5</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.6	1	1	1	0	0	1	1	0	0	1
<i>Commiphora mollis</i>	<b>5</b>	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	7.3	1	0	11	0	0	1	0	0	0	1
<i>Commiphora mollis</i>	5.1	1	0	0	0	0	0	0	0	0	0

<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	9.2	1	0	1	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8.9	1	0	3	0	0	1	0	0	0	1
<i>Sclerocarya birrea</i>	9.6	1	0	4	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	2	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	6.1	1	0	2	0	0	1	0	0	0	0
<i>Combretum apiculatum</i>	6.7	1	1	2	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	8.2	1	0	0	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.5	1	1	0	0	0	1	0	0	0	1
<i>Sclerocarya birrea</i>	9.2	1	0	0	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	7	1	1	0	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	1	0	0.95	0	0	0	0	0
<i>Combretum apiculatum</i>	7.6	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	1	0	0	0	0	1	0	0	0
<i>Combretum apiculatum</i>	5.1	1	0	2	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	2	0	0	0	0	0	0	0
<i>Commiphora mollis</i>	6.3	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	2	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	1	0	0	0	0	0	0	1
<i>Commiphora mollis</i>	4.8	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.4	1	0	0	0	0	0	0	0	0	0

<i>Acacia nigrescens</i>	7.7	1	0	3	0	0	0	0	0	0
<i>Combretum imberbe</i>	7	1	0	3	1	0	1	1	0	1
<i>Combretum apiculatum</i>	5.8	1	1	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8.2	1	0	3	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	3	0	50	1	0	1	1
<i>Acacia nigrescens</i>	8	1	0	0	0	25	1	0	0	0
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9.2	1	0	6	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.2	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.8	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.2	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.3	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8.1	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.9	1	0	1	0	0.1	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	1	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	8.5	1	0	1	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	12	1	0	2	0	0.4	1	0	1	1
<i>Combretum apiculatum</i>	5	1	0	1	0	0	1	0	0	1

<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	5	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.5	1	0	5	0	0	1	0	0	0
<i>Acacia nigrescens</i>	6.9	1	0	5	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	1	0	0	1	0	0	0
<i>Combretum imberbe</i>	9.6	1	0	4	0	0	1	0	0	1
<i>Acacia nigrescens</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	6.6	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6	1	1	1	0	0	1	1	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.3	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.9	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.7	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.4	1	0	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	9	1	0	8	1	0	1	1	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	<b>11</b>	1	0	0	0	0.4	0	0	1	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.3	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	5.9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.2	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.9	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	4.6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9.1	1	0	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.2	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.8	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.2	1	0	1	0	0.5	0	0	1	1
<i>Combretum apiculatum</i>	7.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.1	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6	1	1	1	0	0	1	1	0	1
<i>Sclerocarya birrea</i>	<b>6</b>	1	0	1	0	0	0	0	0	0
<i>Combretum apiculatum</i>	<b>10</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7.5</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>9.5</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.6	1	0	0	0	0	0	0	0	0

<i>Acacia nigrescens</i>	<b>10</b>	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.7	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	5.1	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.9	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>9</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.9	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.9	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>9</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	6.9	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.3	1	1	1	0	0	1	0	0	0	1
<i>Acacia nigrescens</i>	<b>11</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	5.8	1	1	1	0	0	1	0	0	0	0
<i>Acacia nigrescens</i>	7.9	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>9</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>12</b>	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>9</b>	1	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7.7	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	4.4	1	0	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.5	1	0	2	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	2	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.2	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.1	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	8.2	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.7	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10	1	0	0	0	0.1	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.8	0	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	0	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	12	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	2	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.7	1	0	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	8.8	1	0	0	0	0	0	0	0	0	1

<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>10</b>	1	0	0	0	0.05	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>8</b>	1	0	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.9	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>11</b>	1	0	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	1	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>9</b>	1	0	1	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>8</b>	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	4.6	1	1	0	1	0	1	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	<b>9</b>	1	0	0	0	0	1	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.3	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8.5</b>	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8.5	1	0	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	10.3	1	0	0	0	0	0	0	0	0
<i>Sclerocarya birrea</i>	4.1	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	5.1	0	1	1	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	6	0	1	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.4	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.2	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7	1	1	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	8	1	1	0	0	0	1	1	0	1
<i>Combretum apiculatum</i>	6.8	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	8.8	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.5	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5	1	1	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6.7	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	4.5	0	1	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	11	1	0	0	0	0.4	0	0	1	1
<i>Acacia nigrescens</i>	6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	9	1	0	0	0	0	0	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0.3	1	0	1	1
<i>Acacia nigrescens</i>	8	0	1	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	10	1	0	6	0	0.05	1	0	0	1
<i>Acacia nigrescens</i>	10	1	0	3	0	0.1	1	0	0	0
<i>Acacia nigrescens</i>	8	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.9	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.2	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8.8	1	0	2	0	0	0	0	0	1

<i>Acacia nigrescens</i>	6.5	1	1	2	1	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8.4	1	0	1	0	0.5	0	0	1	1
<i>Acacia nigrescens</i>	<b>6</b>	1	0	2	1	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>9</b>	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>12</b>	1	0	4	0	0.4	1	0	0	1
<i>Acacia nigrescens</i>	<b>12</b>	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	5.9	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.2	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.5	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>11</b>	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	5.6	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4	1	0	2	1	0	1	0	0	1
<i>Combretum apiculatum</i>	5.5	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5.6	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>7</b>	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	5.4	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.1	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	5.7	1	0	5	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0.05	0	0	1	1
<i>Acacia nigrescens</i>	5.9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	2	0	0	0	0	0	1

<i>Acacia nigrescens</i>	7.5	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8	1	0	3	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6	1	0	2	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.3	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	2	1	0	0	0	0	1
<i>Combretum apiculatum</i>	6.6	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.3	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	5	1	0	3	0	0	1	0	0	1
<i>Combretum apiculatum</i>	6	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6.9	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	4.7	1	0	4	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	9	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	6	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	4.8	1	0	0	0	0	1	0	0	1
<i>Combretum apiculatum</i>	8.1	1	0	1	0	0	0	0	0	1
<i>Combretum apiculatum</i>	8	1	0	0	0	0	0	0	0	0
<i>Combretum apiculatum</i>	9	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	9.9	1	0	2	0	0	0	0	0	1
<i>Combretum imberbe</i>	14	1	0	3	0	0	0	0	0	1
<i>Combretum imberbe</i>	11.3	1	0	3	0	0	0	0	0	1
<i>Acacia nigrescens</i>	12.3	1	0	3	0	30	0	0	0	0
<i>Combretum imberbe</i>	12	1	0	4	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8.9	1	0	1	0	70	0	0	1	1
<i>Combretum imberbe</i>	15	1	0	4	0	0	1	0	0	0
<i>Combretum imberbe</i>	11.1	1	0	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.5	1	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	10.3	1	0	0	0	80	0	0	1	1
<i>Combretum imberbe</i>	8.8	1	0	5	0	0	1	0	0	1
<i>Acacia nigrescens</i>	8.9	1	0	1	0	5	1	0	0	1
<i>Acacia nigrescens</i>	8.3	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	1	2	0	0.1	0	0	1	1
<i>Combretum apiculatum</i>	<b>4.5</b>	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	4.3	0	1	2	0	0.3	0	0	0	1
<i>Combretum apiculatum</i>	5.9	1	0	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>10</b>	1	1	3	0	0	1	0	0	0
<i>Acacia nigrescens</i>	6.7	1	0	3	0	0	1	0	0	0
<i>Acacia nigrescens</i>	<b>7</b>	1	0	2	0	0.05	0	0	1	1
<i>Combretum apiculatum</i>	5.6	1	1	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	6.2	0	1	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.4	1	0	2	0	0	1	0	0	0
<i>Acacia nigrescens</i>	7.2	1	1	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	6.3	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>7.5</b>	1	0	2	0	0	0	0	0	0
<i>Acacia nigrescens</i>	<b>8</b>	1	0	1	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7.3	1	1	4	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>7</b>	1	0	4	1	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>10</b>	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	7.2	1	1	4	0	0	1	0	0	1
<i>Combretum apiculatum</i>	5.2	1	1	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	7.2	1	1	2	0	0	0	0	0	1
<i>Acacia nigrescens</i>	8.4	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	0	0	0.6	1	0	1	1
<i>Acacia nigrescens</i>	6.5	1	0	4	0	0.25	1	0	0	1
<i>Acacia nigrescens</i>	<b>12</b>	1	0	0	0	0.1	0	0	1	1
<i>Combretum imberbe</i>	<b>10</b>	1	0	3	0	0	1	0	0	1

<i>Sclerocarya birrea</i>	5.4	1	0	3	0	0	1	0	0	0
<i>Sclerocarya birrea</i>	6.6	1	0	7	0	0	1	0	0	1
<i>Sclerocarya birrea</i>	12	1	0	2	0	0.35	1	0	1	0
<i>Combretum imberbe</i>	10	1	0	4	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8	1	0	3	0	0.5	1	0	0	1
<i>Acacia nigrescens</i>	4.9	1	1	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	6	1	1	2	0	0	1	1	0	1
<i>Acacia nigrescens</i>	6	1	0	3	0	0	1	0	0	0
<i>Acacia nigrescens</i>	7	1	0	2	0	0.8	0	0	0	1
<i>Combretum apiculatum</i>	7	1	1	2	0	0	1	1	0	1
<i>Acacia nigrescens</i>	8	0	1	4	0	0.25	0	0	0	1
<i>Combretum apiculatum</i>	4	1	0	2	0	0	1	1	0	1
<i>Combretum apiculatum</i>	5	1	0	2	0	0	0	0	0	1
<i>Combretum imberbe</i>	6	1	0	4	0	0	1	0	0	1
<i>Acacia nigrescens</i>	9.2	0	0	6	0	0.9	1	0	0	1
<i>Combretum imberbe</i>	15	1	0	5	0	0	1	0	0	1
<i>Combretum imberbe</i>	5.3	1	0	3	0	0	1	0	0	1
<i>Acacia nigrescens</i>	10	1	0	3	0	0	1	0	0	0
<i>Acacia nigrescens</i>	10	1	0	2	0	0	0	0	0	0
<i>Acacia nigrescens</i>	10	1	0	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	12.1	1	0	2	0	0.5	1	0	1	1
<i>Combretum imberbe</i>	7	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1
<i>Combretum imberbe</i>	7	1	1	2	0	0	1	0	0	1
<i>Acacia nigrescens</i>	5.9	1	0	0	0	0	0	0	0	1
<i>Acacia nigrescens</i>	10	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	9	1	0	0	0	0	0	0	0	0
<i>Acacia nigrescens</i>	8.3	1	1	1	0	0	0	0	0	0
<i>Acacia nigrescens</i>	7	1	0	0	0	0	0	0	0	1

<i>Acacia nigrescens</i>	4.7	1	1	1	0	0	1	0	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	1	0	0	0.25	0	0	0	0
<i>Acacia nigrescens</i>	4.5	1	0	0	1	0	1	0	0	1
<i>Acacia nigrescens</i>	4.9	1	0	0	0	0	0	0	0	1
<i>Combretum apiculatum</i>	<b>5</b>	1	1	2	0	0	1	1	0	1
<i>Acacia nigrescens</i>	<b>8</b>	1	0	2	0	0	1	0	0	1